



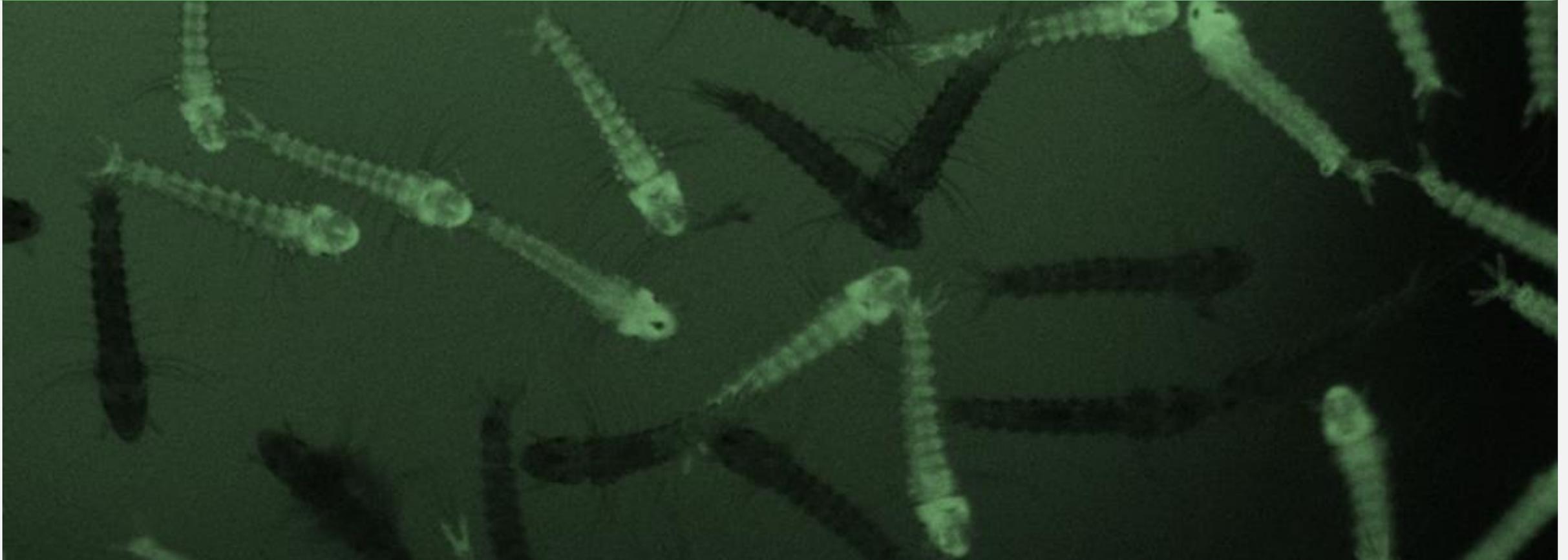
Development of Genetic Sexing Strains for the application of the Sterile Insect Technique in Aedes albopictus and Aedes aegypti

Célia Lutrat

celia.lutrat@outlook.com



INTRODUCTION



INTRODUCTION

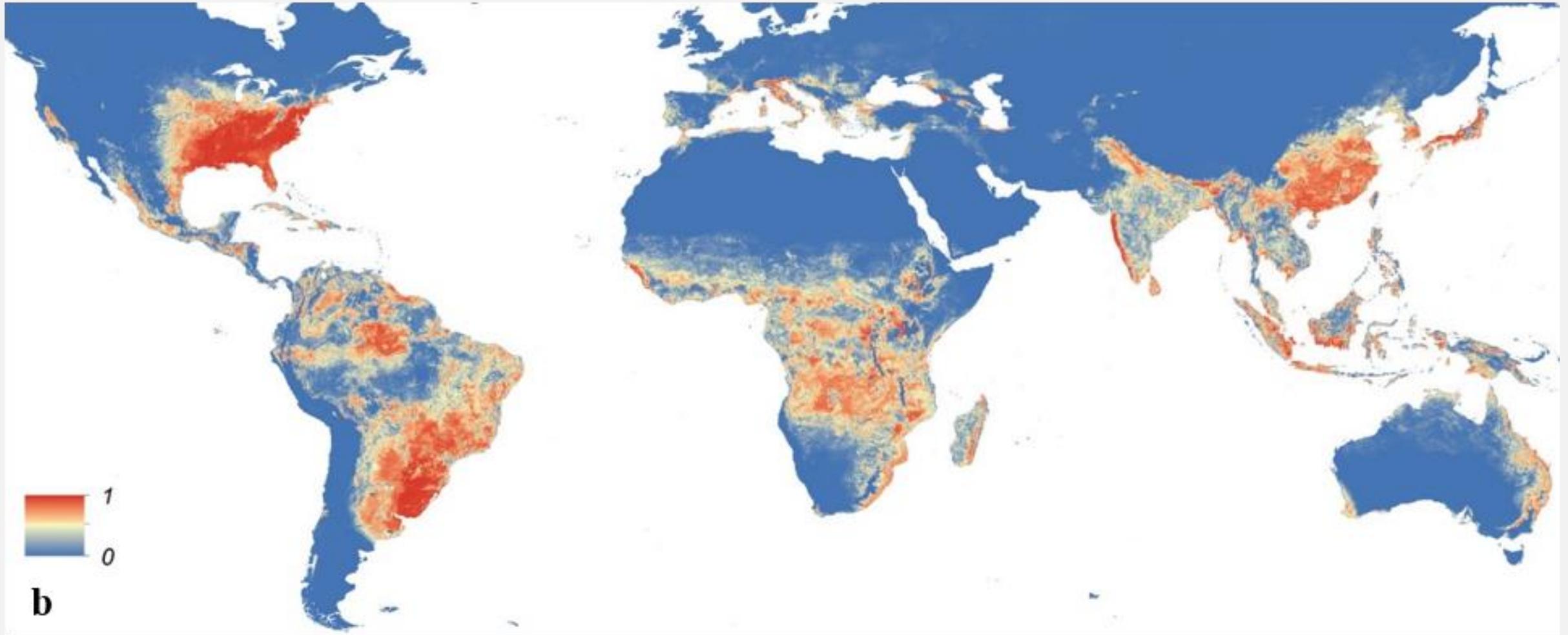
Aedes albopictus



Jim Occi, BugPics, Bugwood.org

INTRODUCTION

World distribution of *Aedes albopictus*

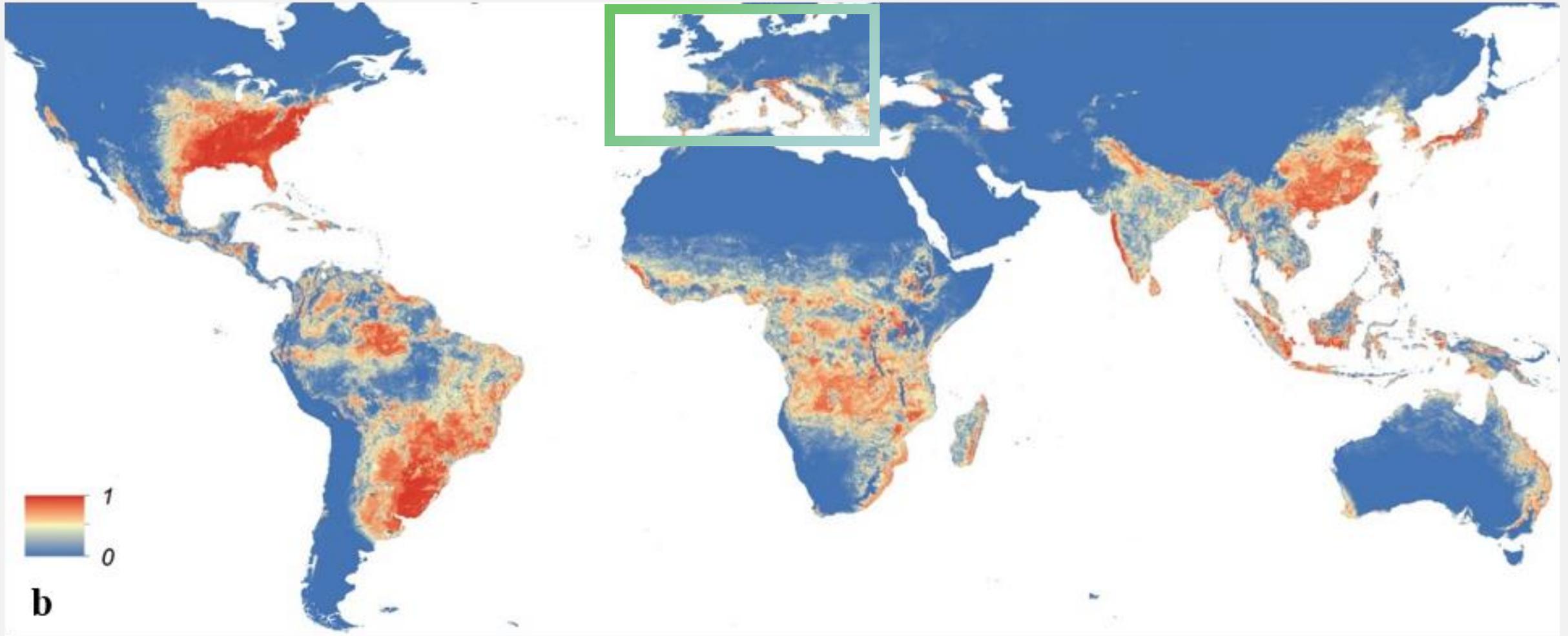


Kraemer et al. 2015

Reiter 1998

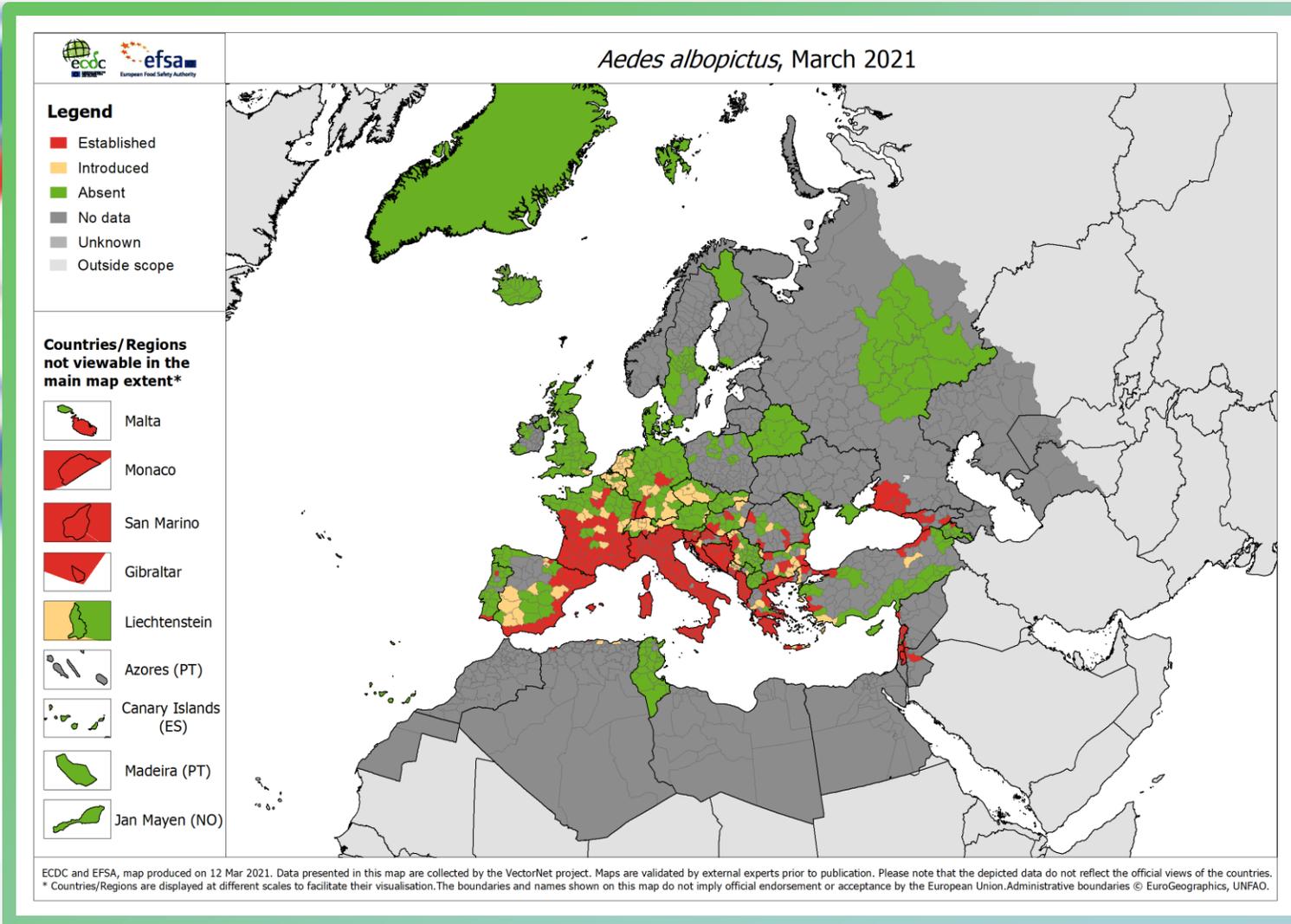
INTRODUCTION

World distribution of *Aedes albopictus*



INTRODUCTION

World distribution of *Aedes albopictus*



INTRODUCTION

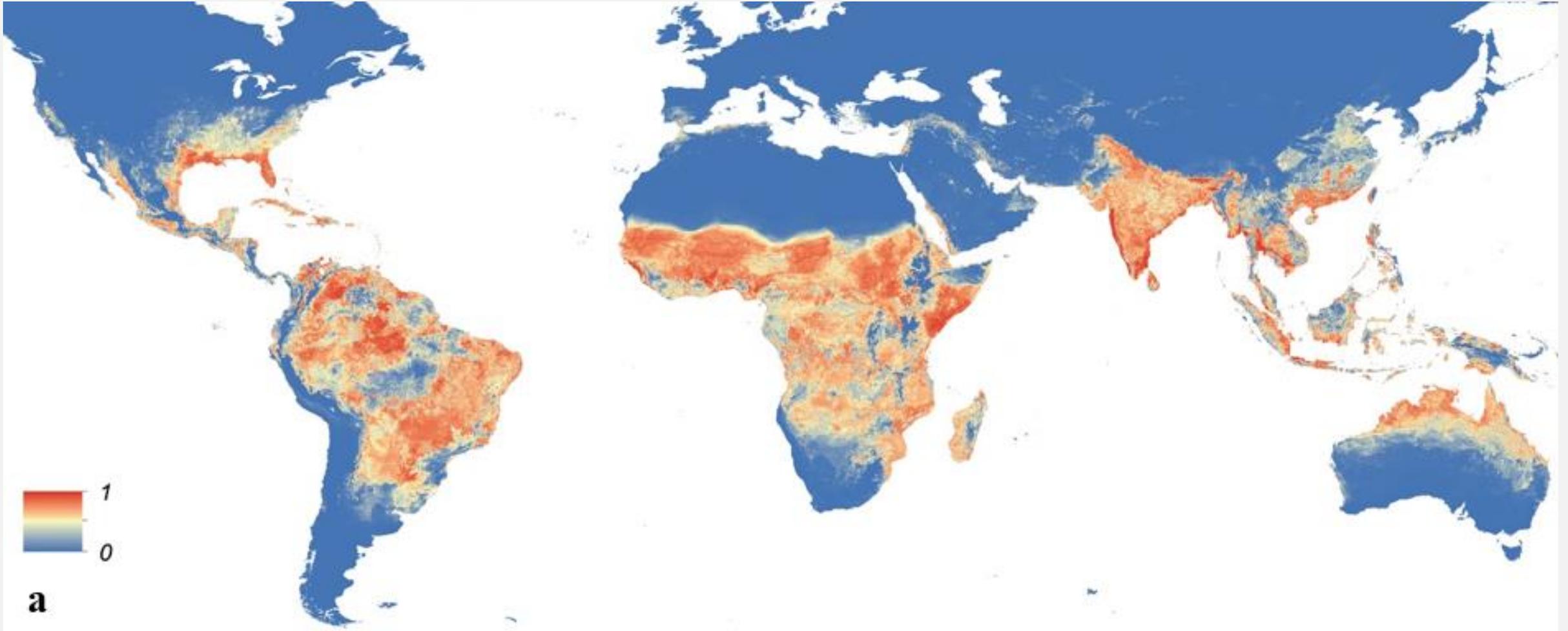
Aedes aegypti



James Gathany - PHIL, CDC

INTRODUCTION

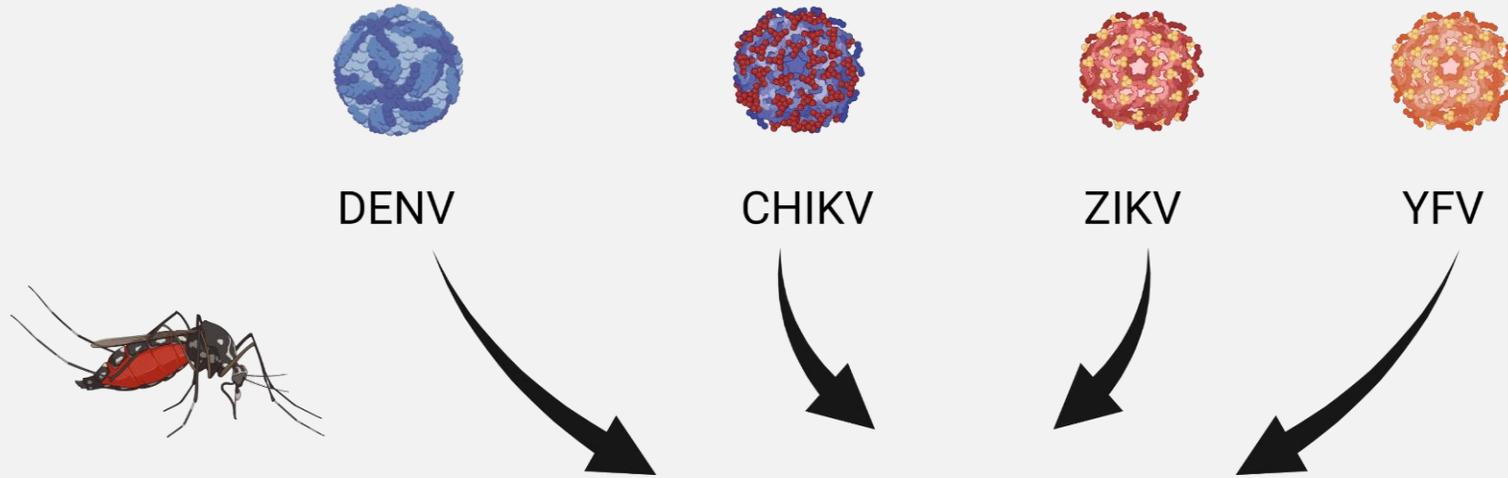
World distribution of *Aedes aegypti*



Kraemer et al. 2015
Soghigian et al. 2020

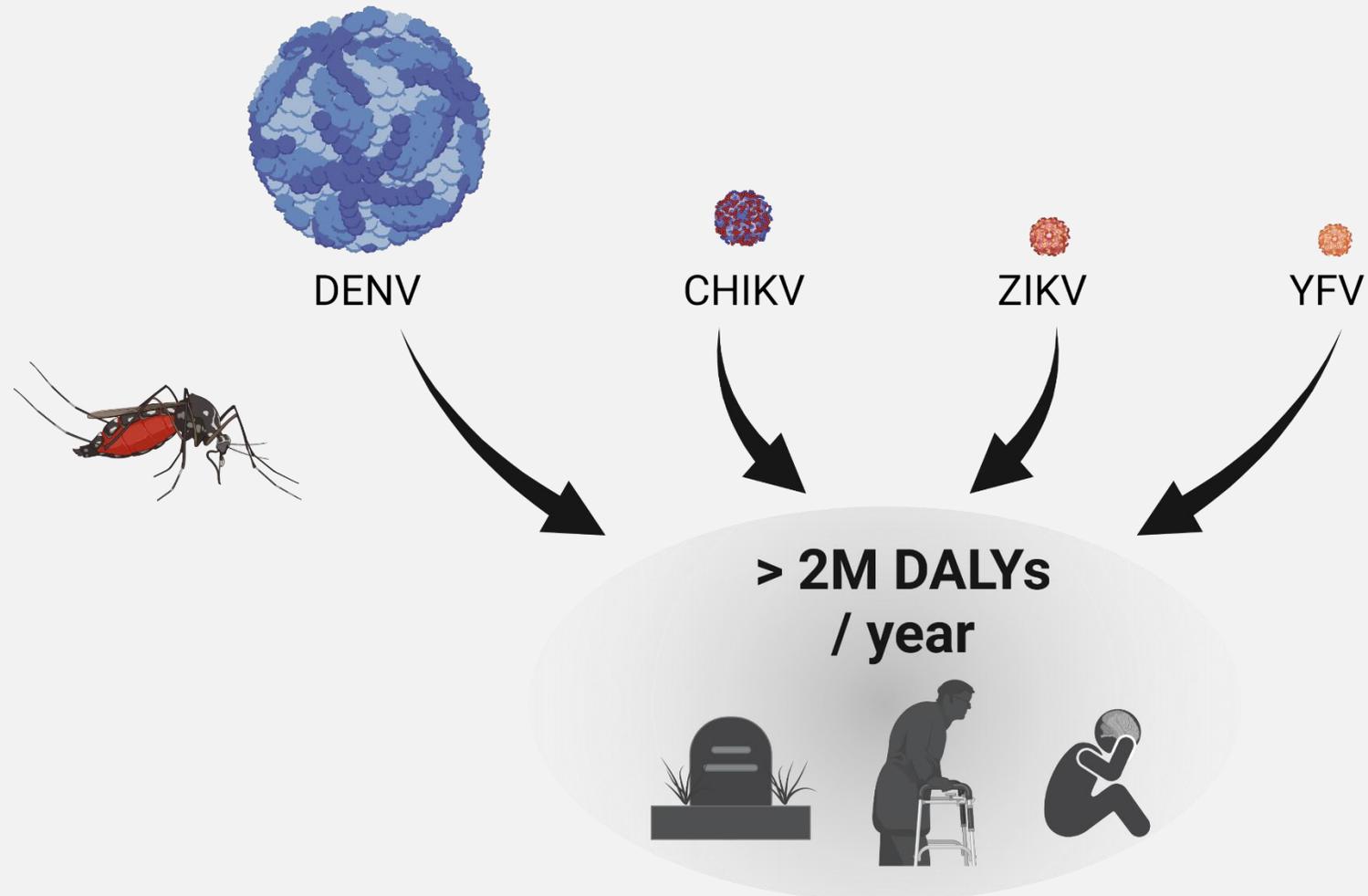
INTRODUCTION

Aedes-borne viruses cause severe illnesses costing over 2M DALYs annually



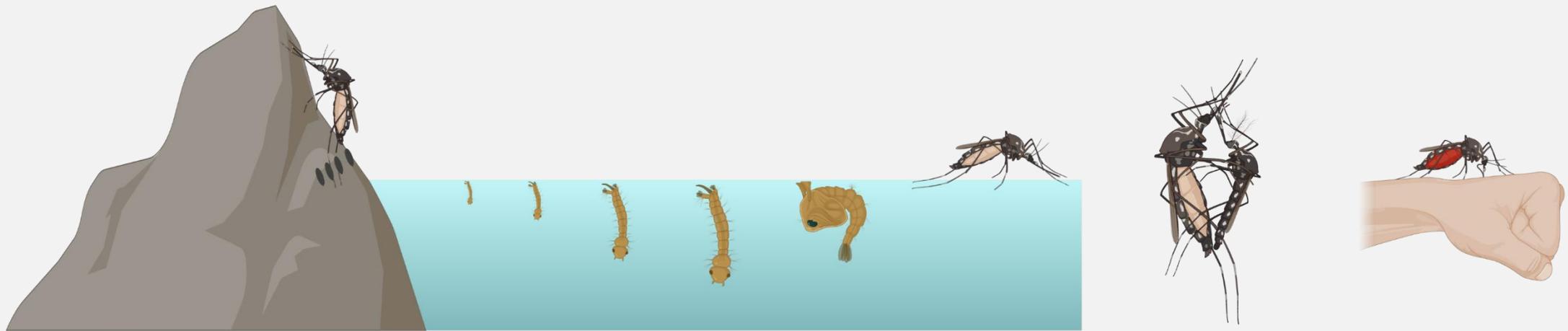
INTRODUCTION

Aedes-borne viruses cause severe illnesses costing over 2M DALYs annually



INTRODUCTION

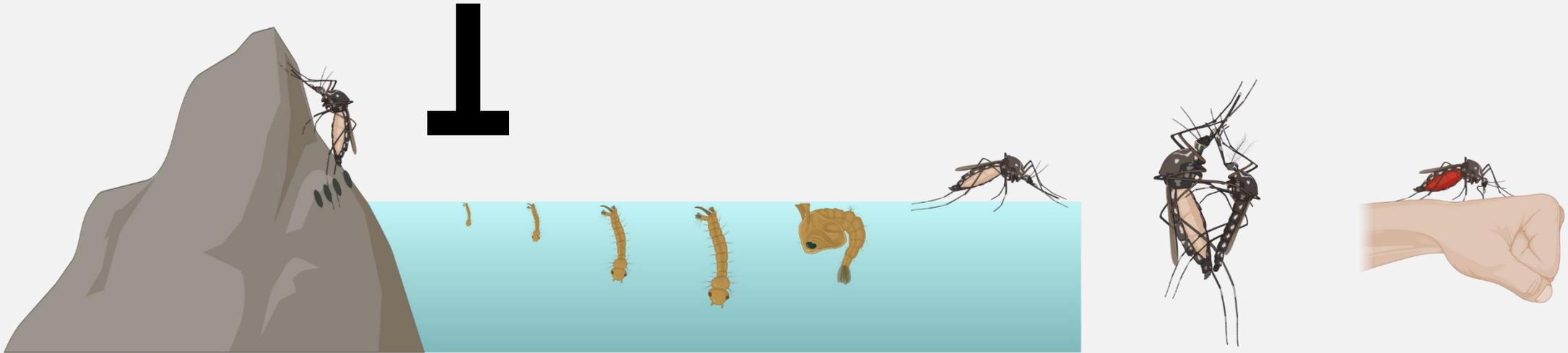
Life cycle of *Aedes* mosquitoes



INTRODUCTION

Current vector control against *Aedes* mosquitoes

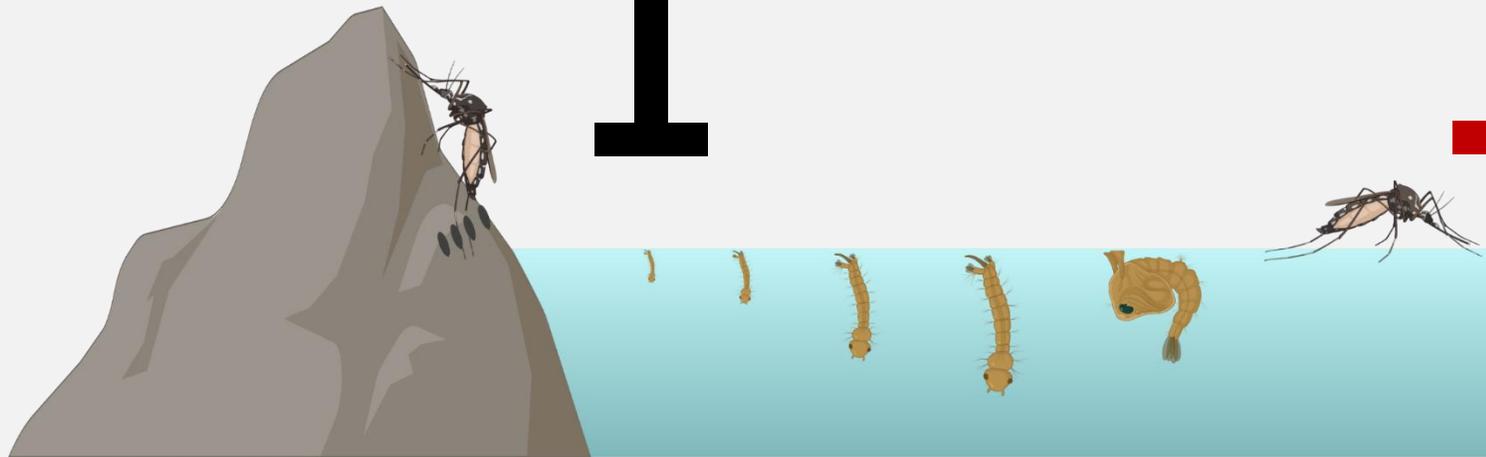
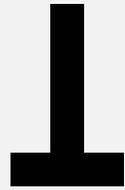
Mechanical suppression
+ Larvicides



INTRODUCTION

Current vector control against *Aedes* mosquitoes

Mechanical suppression
+ Larvicides

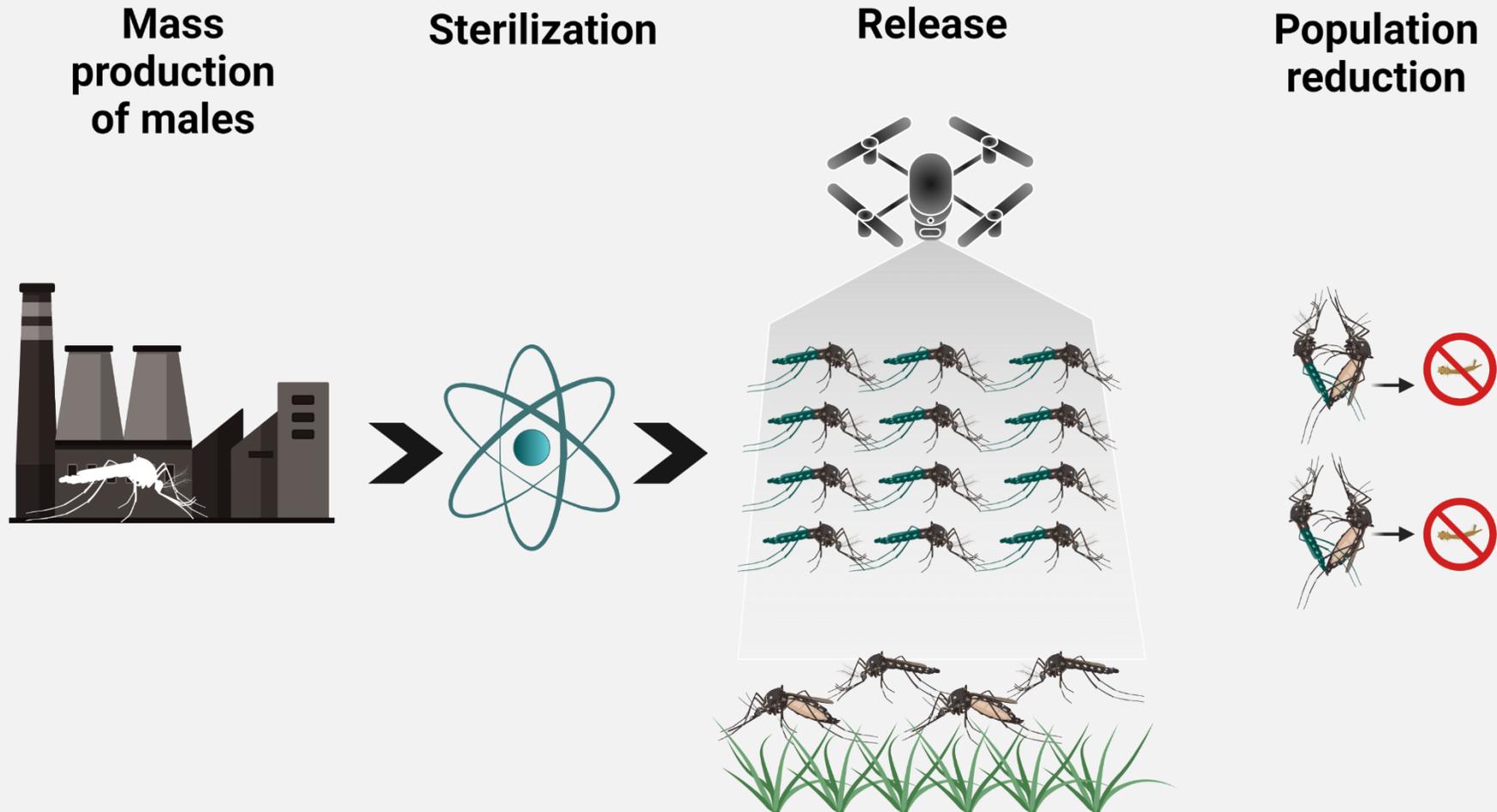


Insecticide
spraying



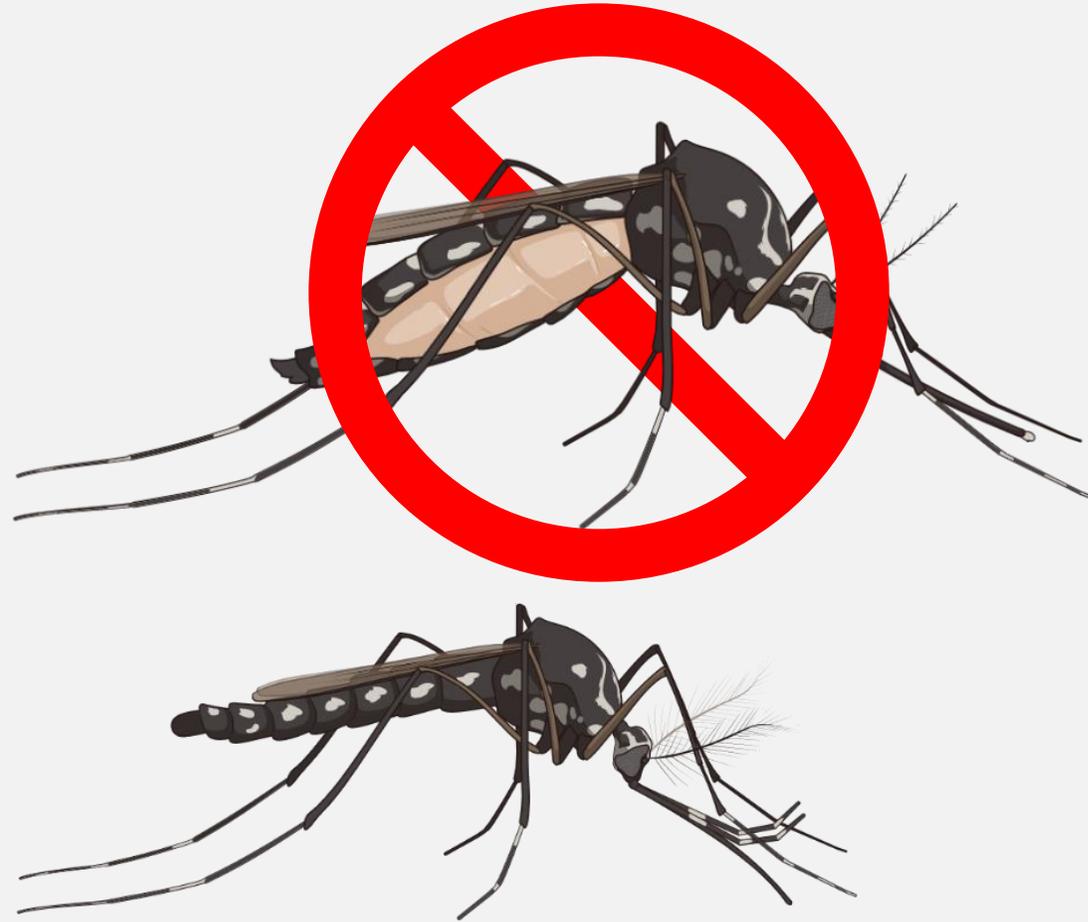
INTRODUCTION

The Sterile Insect Technique (SIT) is a promising alternative



INTRODUCTION

Sex separation is the main bottleneck to *Aedes* SIT



INTRODUCTION

Aedes sex dimorphism



INTRODUCTION

Current sexing methods exploit the size dimorphism at the pupal stage



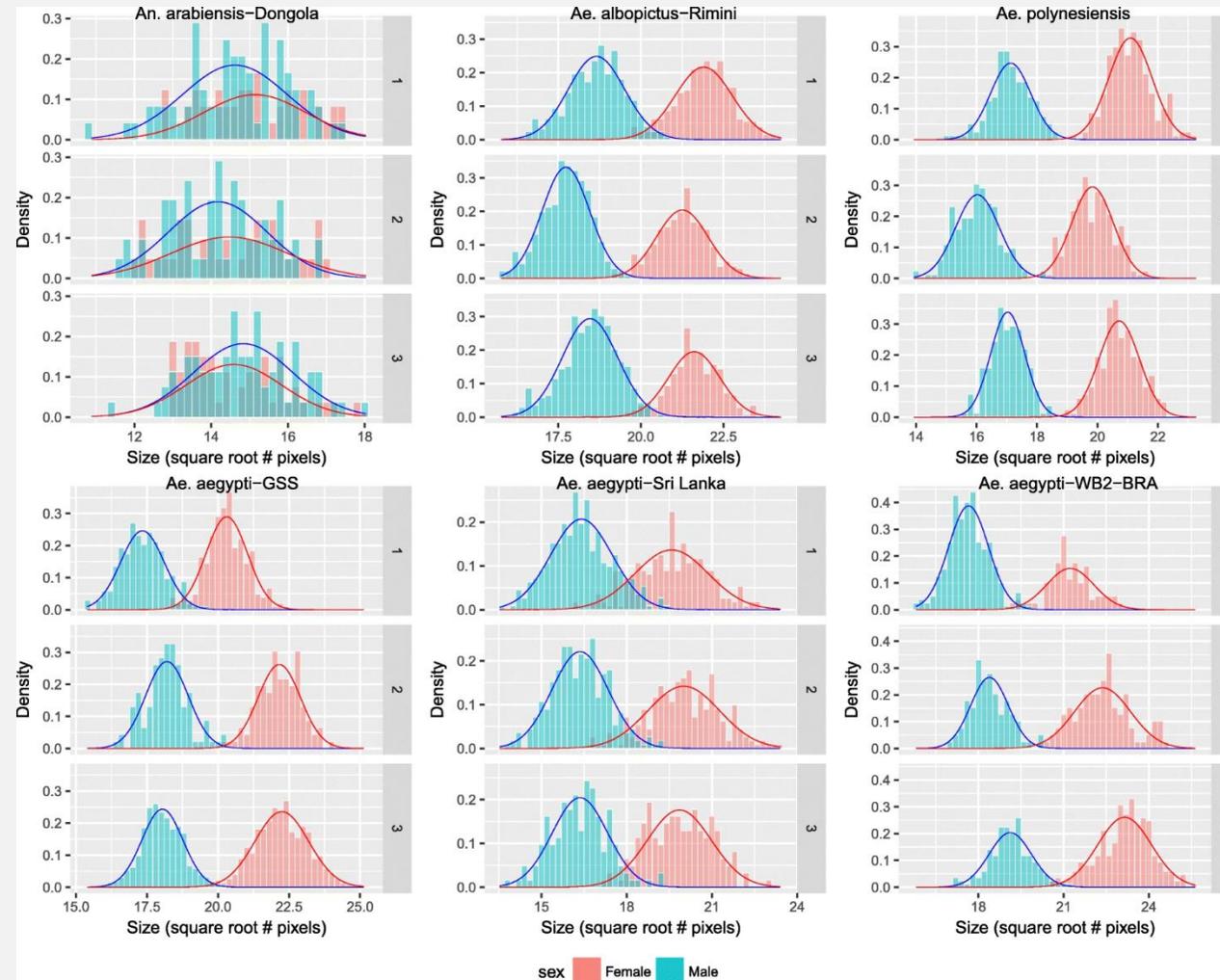
INTRODUCTION

Current sexing methods exploit the size dimorphism at the pupal stage



INTRODUCTION

Current sexing methods exploit the size dimorphism at the pupal stage



INTRODUCTION



30% male recovery
 $\leq 1\%$ female contamination
1800 pupae /min

INTRODUCTION



30% male recovery
 $\leq 1\%$ female contamination
1800 pupae /min



Up to 70% male recovery
 $\leq 1\%$ female contamination
? pupae /min

OBJECTIVE:

Improve the cost-efficiency ratio
of *Aedes* sex separation



OBJECTIVE

Comparison of the sexing methods developed during the last 15 years

Trends in Parasitology 

Volume 35, Issue 8, August 2019, Pages 649-662

Review

Sex Sorting for Pest Control: It's Raining Men!

Célia Lutrat ^{1, 2, 3}, David Giesbrecht ⁴, Eric Marois ⁵, Steve Whyard ⁴, Thierry Baldet ^{1, 2}, Jérémy Bouyer ^{1, 6} 



OBJECTIVE

7 criteria for a good sexing method

Sorting stage

OBJECTIVE

7 criteria for a good sexing method

Sorting stage

Male recovery

OBJECTIVE

7 criteria for a good sexing method

Sorting stage

Male recovery

Female contamination

OBJECTIVE

7 criteria for a good sexing method

Sorting stage

Male recovery

Female contamination

Sorting speed

OBJECTIVE

7 criteria for a good sexing method

Sorting stage

Male recovery

Female contamination

Sorting speed

Initial investment

OBJECTIVE

7 criteria for a good sexing method

Sorting stage

Male recovery

Female contamination

Sorting speed

Initial investment

Consumable cost

OBJECTIVE

7 criteria for a good sexing method

Sorting stage

Male recovery

Female contamination

Sorting speed

Initial investment

Consumable cost

Acceptability

OBJECTIVE

3 strategies identified

Conditional female death /
conditional masculinization



OBJECTIVE

3 strategies identified

Conditional female death /
conditional masculinization



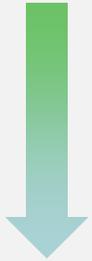
Converting females into
males by RNAi



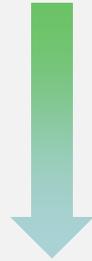
OBJECTIVE

3 strategies identified

Conditional female death /
conditional masculinization



Converting females into
males by RNAi

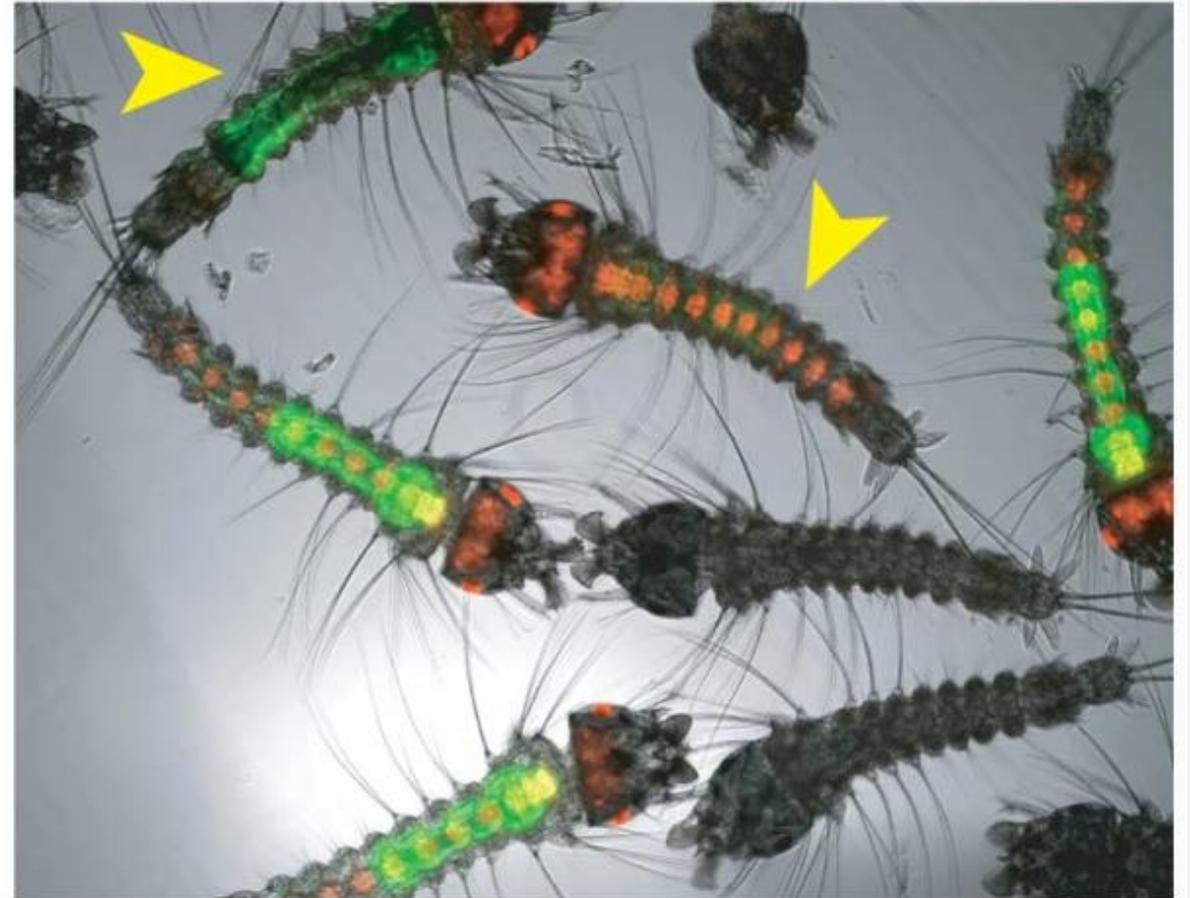
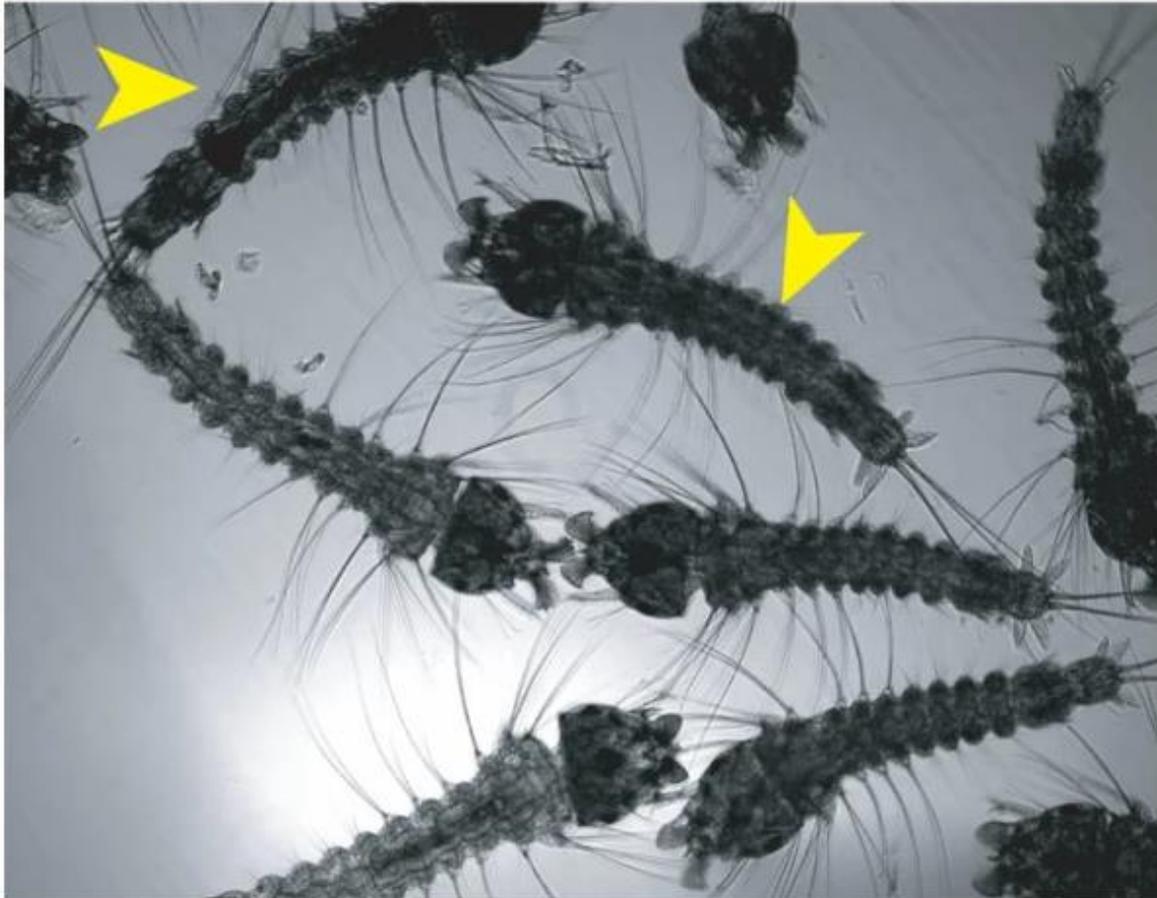


Sex-specific expression of a
fluorescence marker



OBJECTIVE

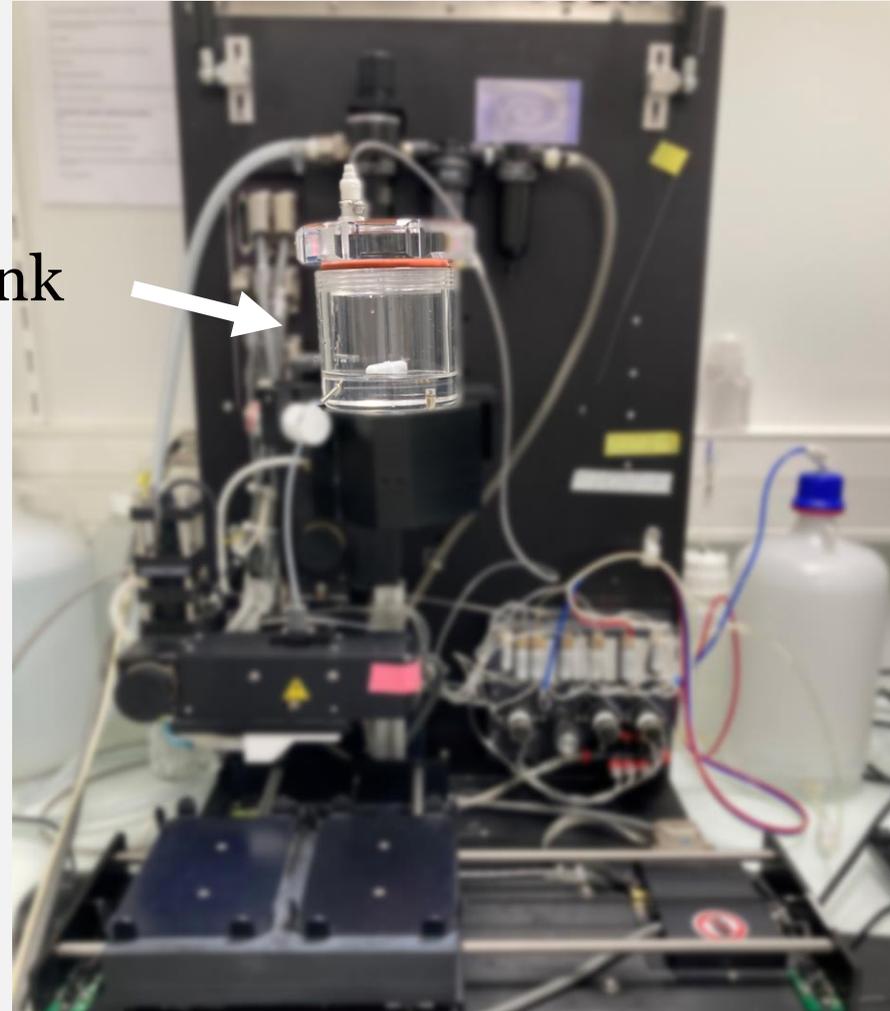
This approach has already been developed in *Anopheles* mosquitoes



OBJECTIVE

Neonate larvae can be sorted based on fluorescence using a COPAS device

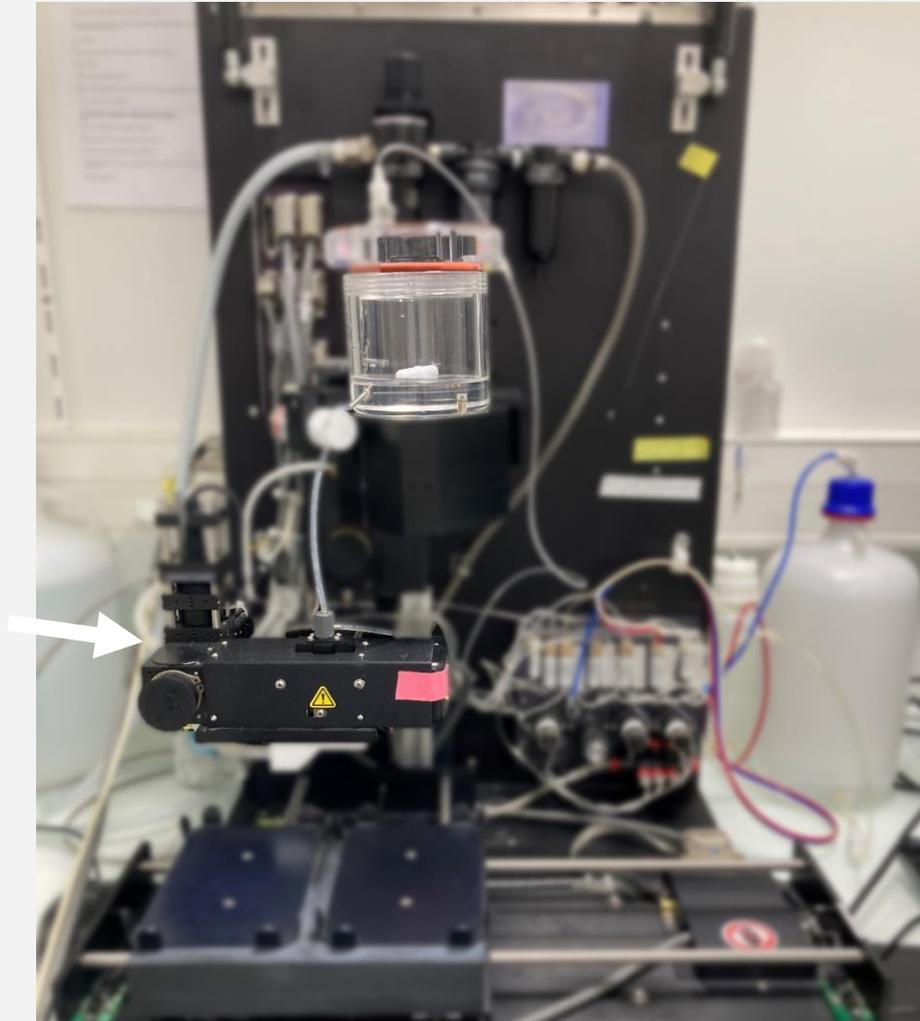
Sample tank



OBJECTIVE

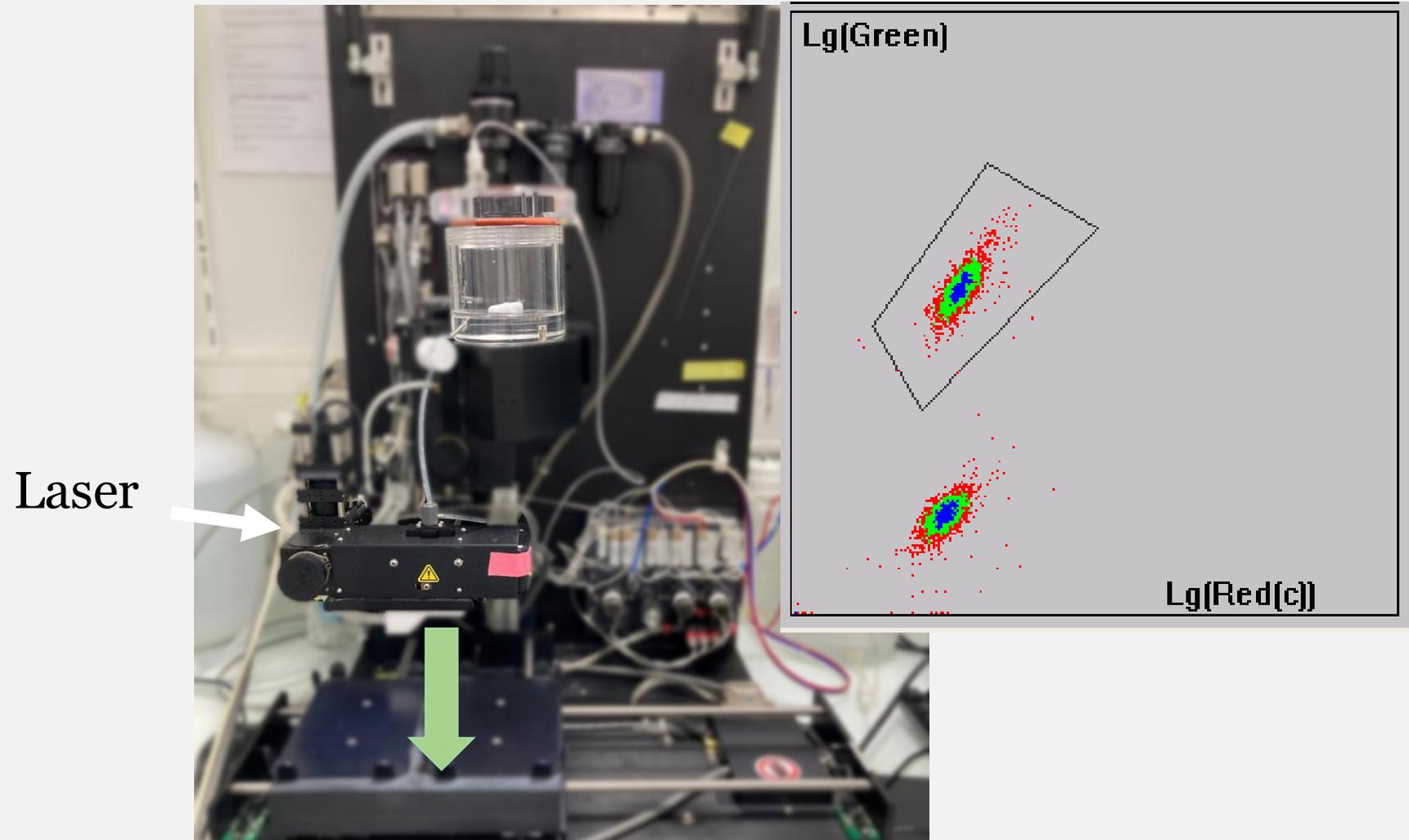
Neonate larvae can be sorted based on fluorescence using a COPAS device

Laser



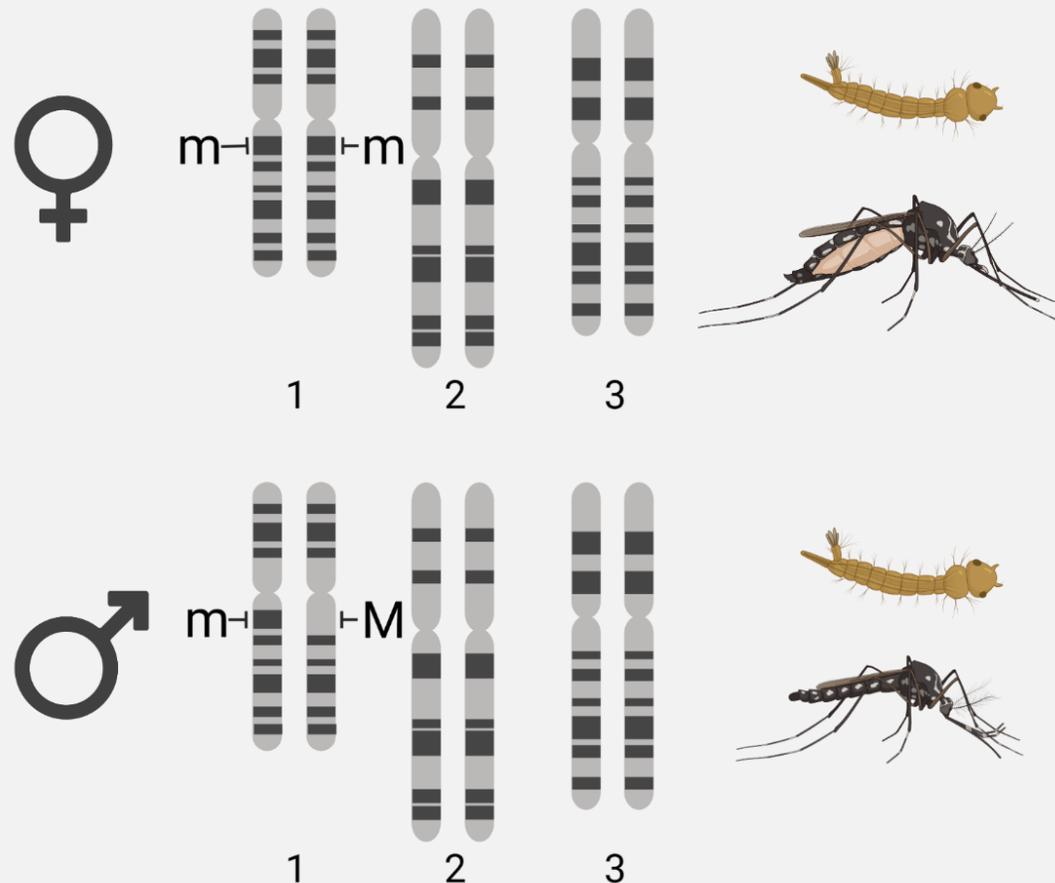
OBJECTIVE

Neonate larvae can be sorted based on fluorescence using a COPAS device



OBJECTIVE

Developing Genetic Sexing Strains carrying a sex-specific fluorescent markers



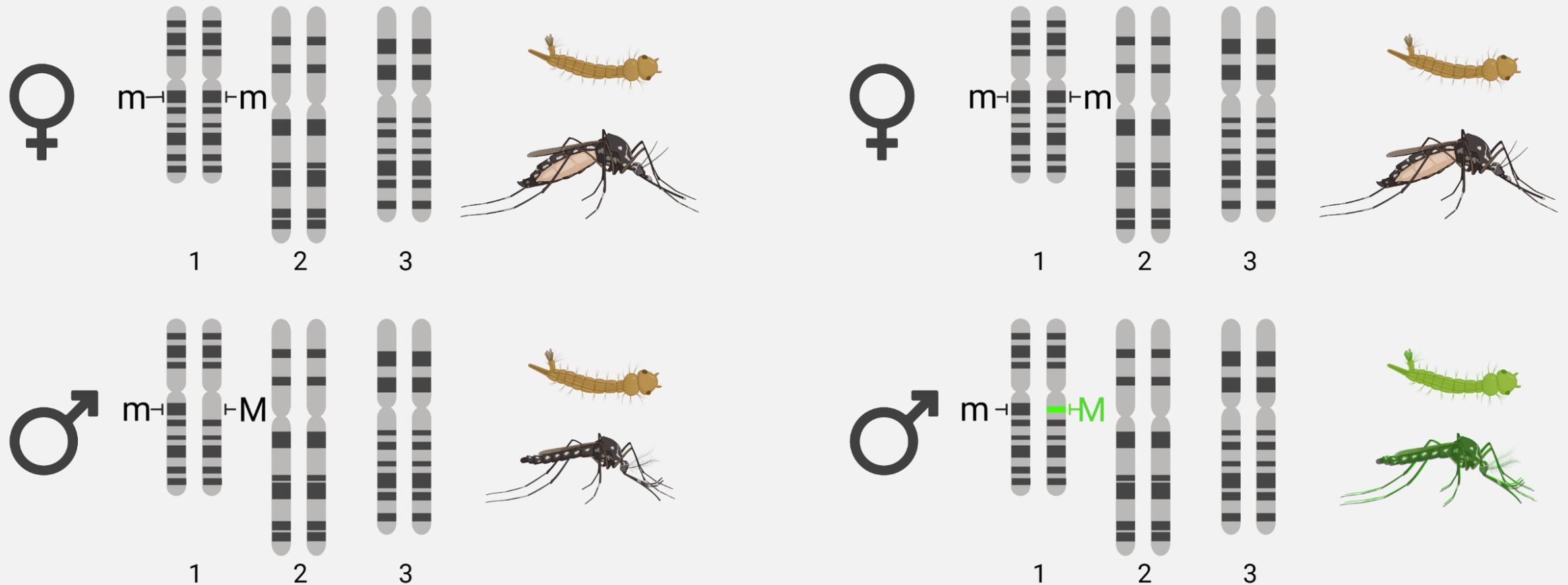
OBJECTIVE

Developing Genetic Sexing Strains carrying a sex-specific fluorescent markers



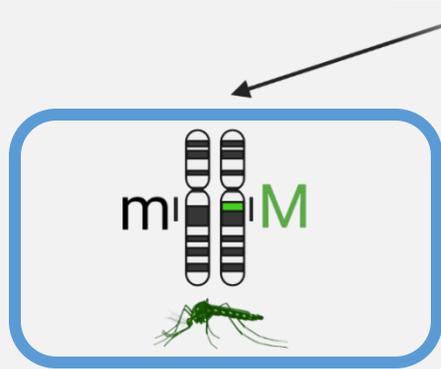
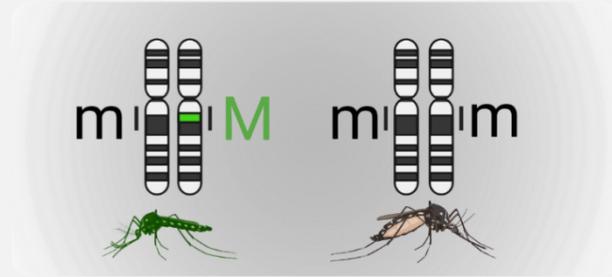
OBJECTIVE

Developing Genetic Sexing Strains carrying a sex-specific fluorescent markers



OBJECTIVE

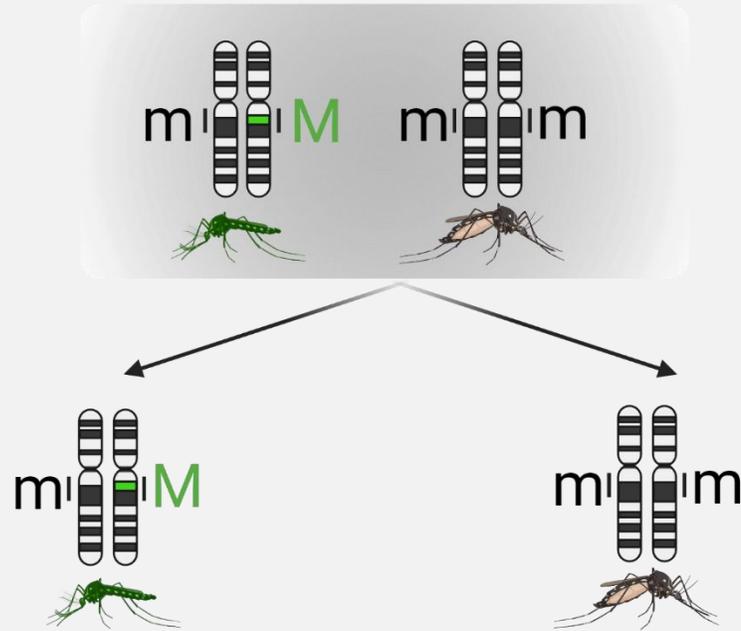
2 GSS allow 2 uses: transgenic or non-transgenic



Transgenic males

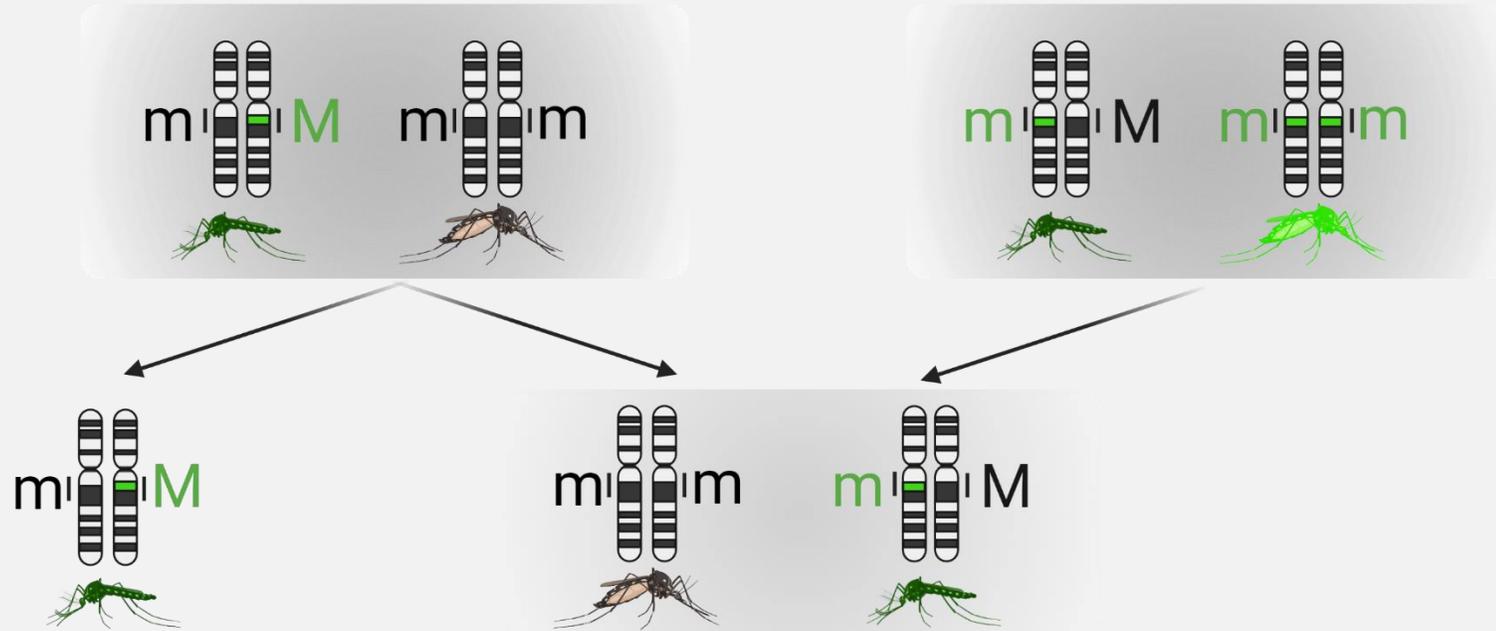
OBJECTIVE

2 GSS allow 2 uses: transgenic or non-transgenic



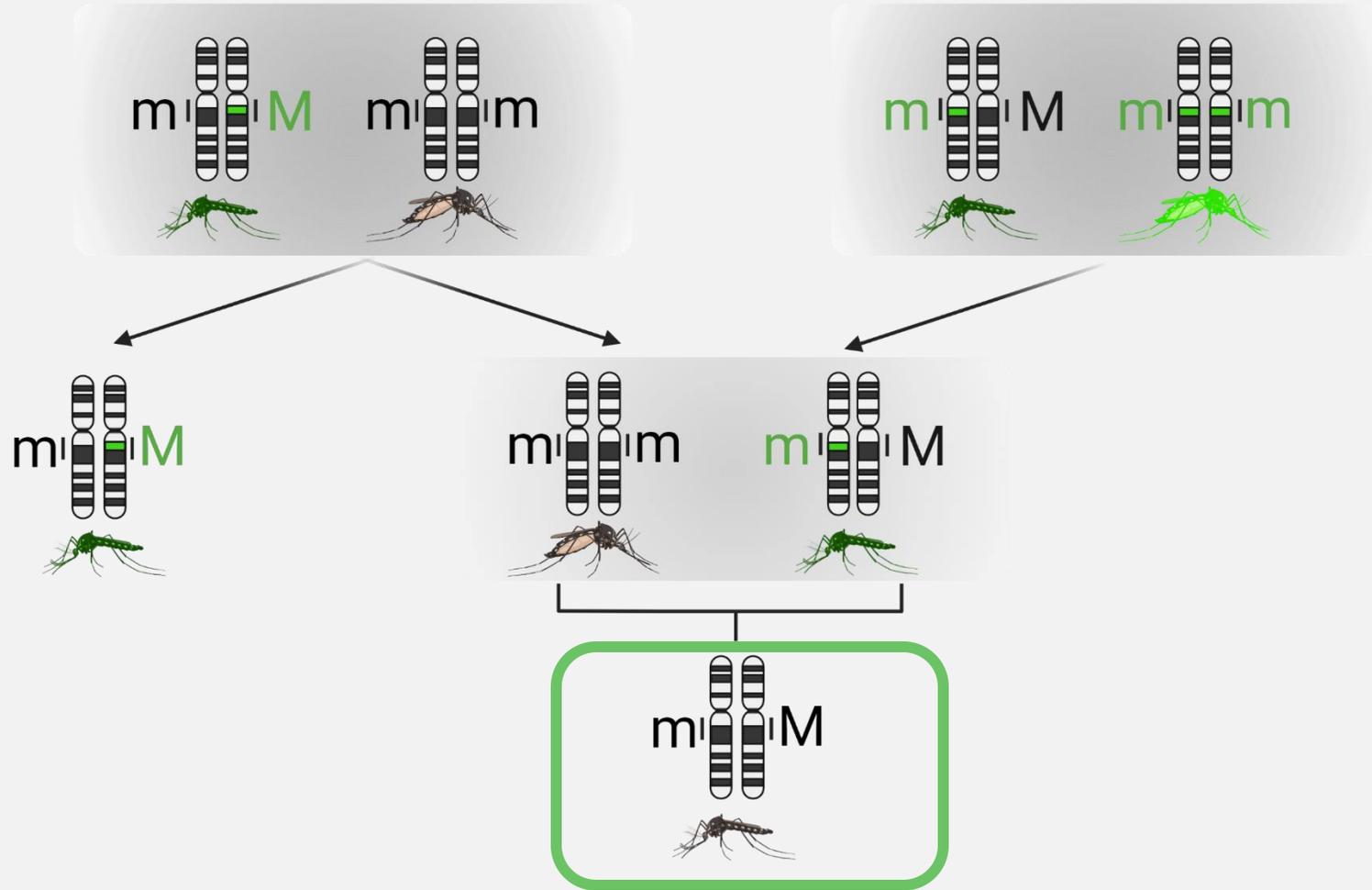
OBJECTIVE

2 GSS allow 2 uses: transgenic or non-transgenic



OBJECTIVE

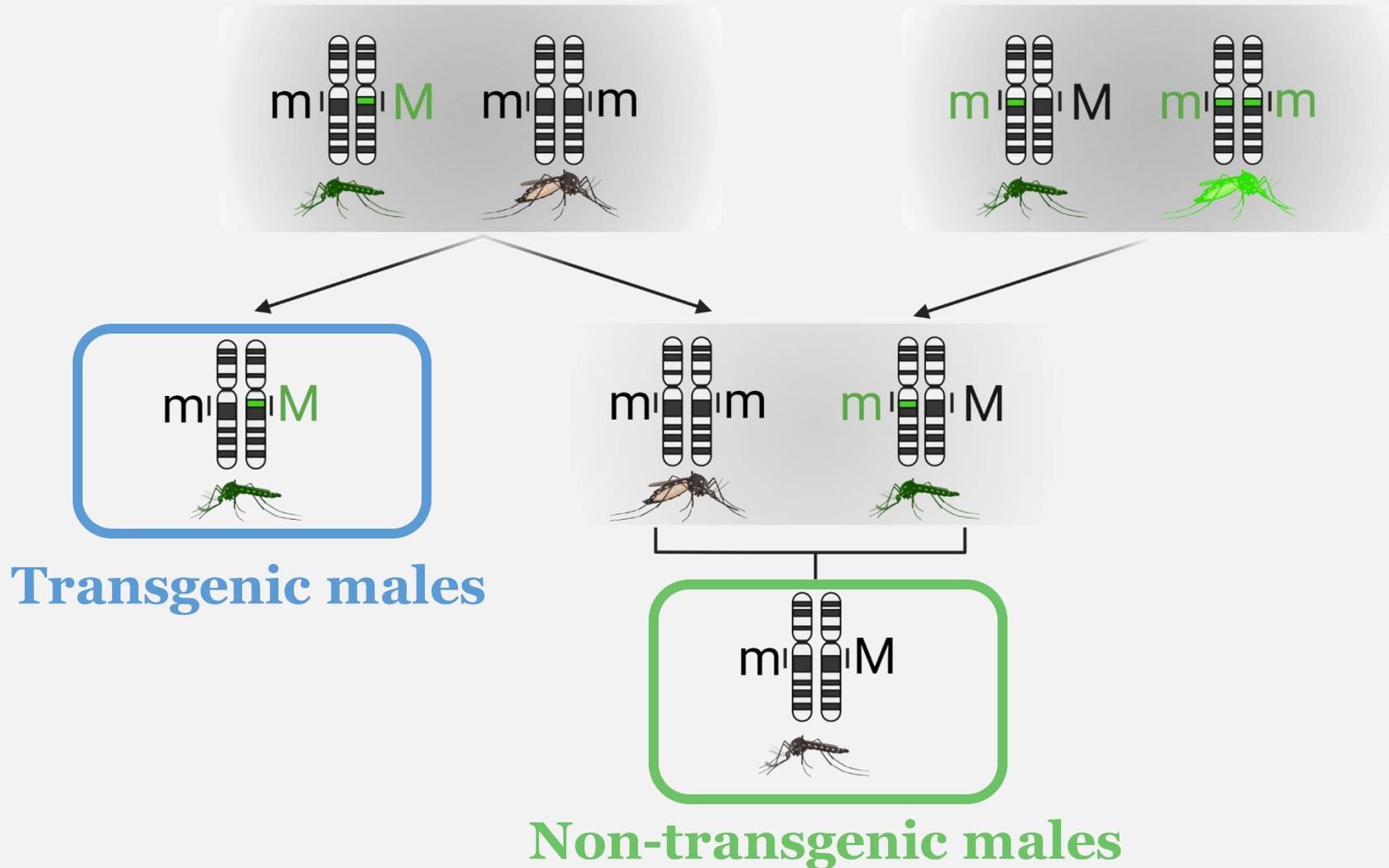
2 GSS allow 2 uses: transgenic or non-transgenic



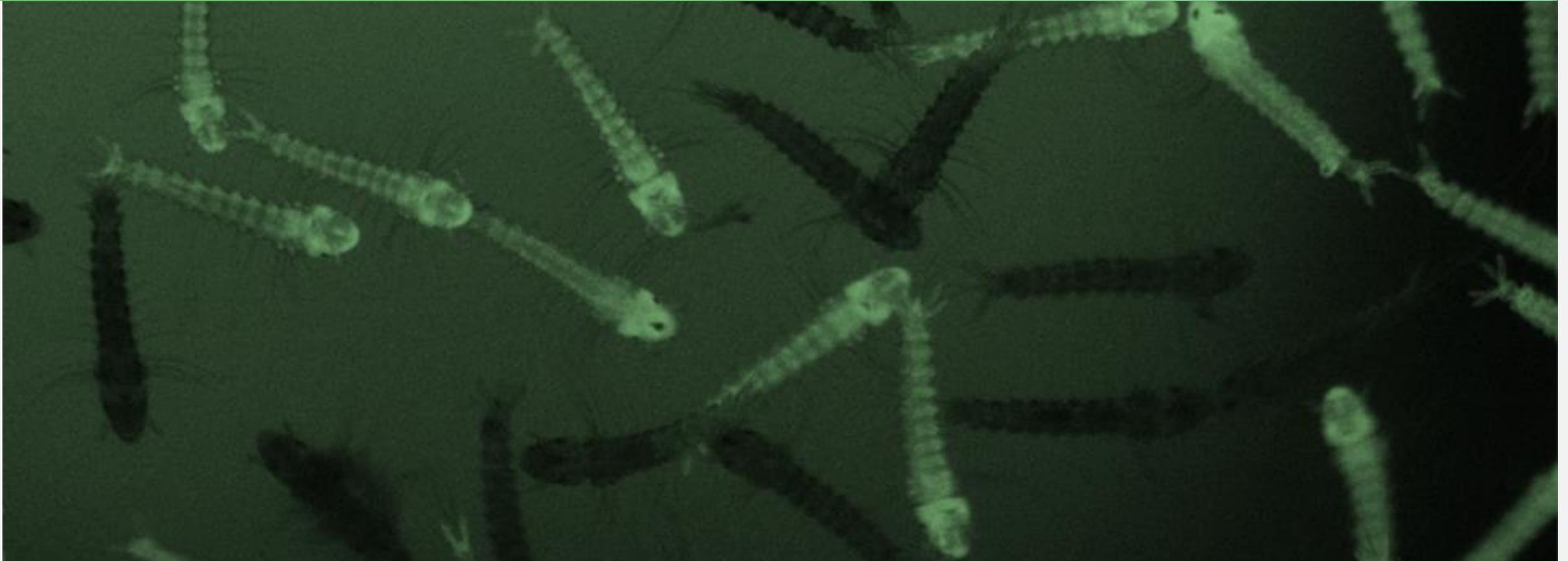
Non-transgenic males

OBJECTIVE

2 GSS allow 2 uses: transgenic or non-transgenic

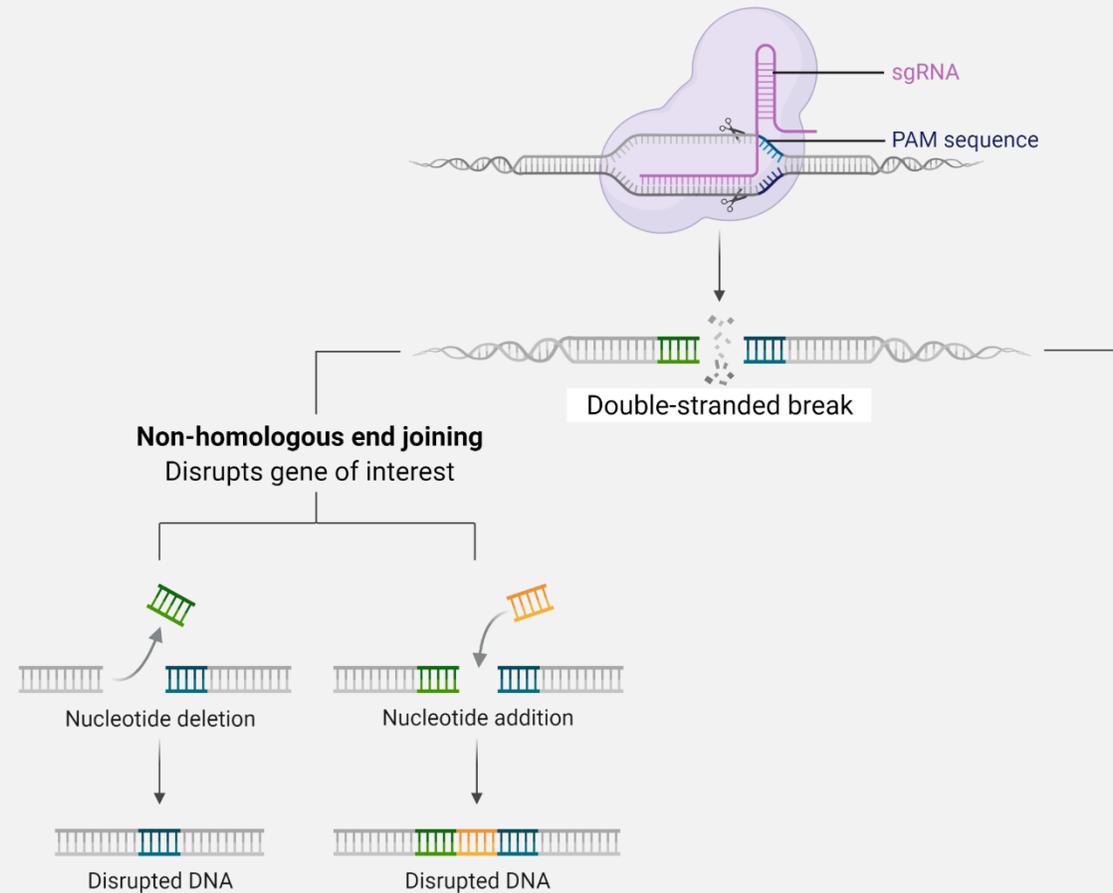


METHODS



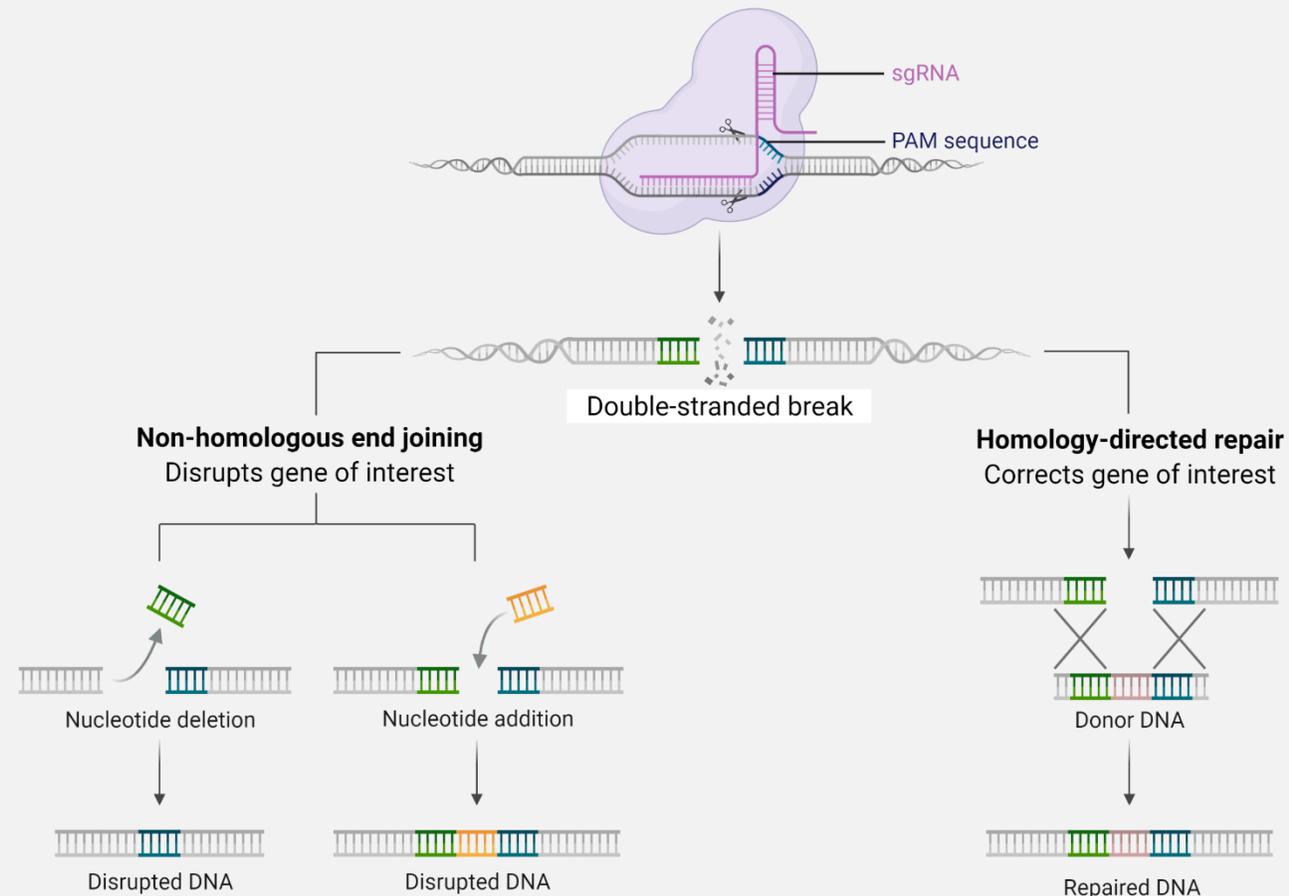
METHODS

CRISPR-Cas9 allows targeted genome editing



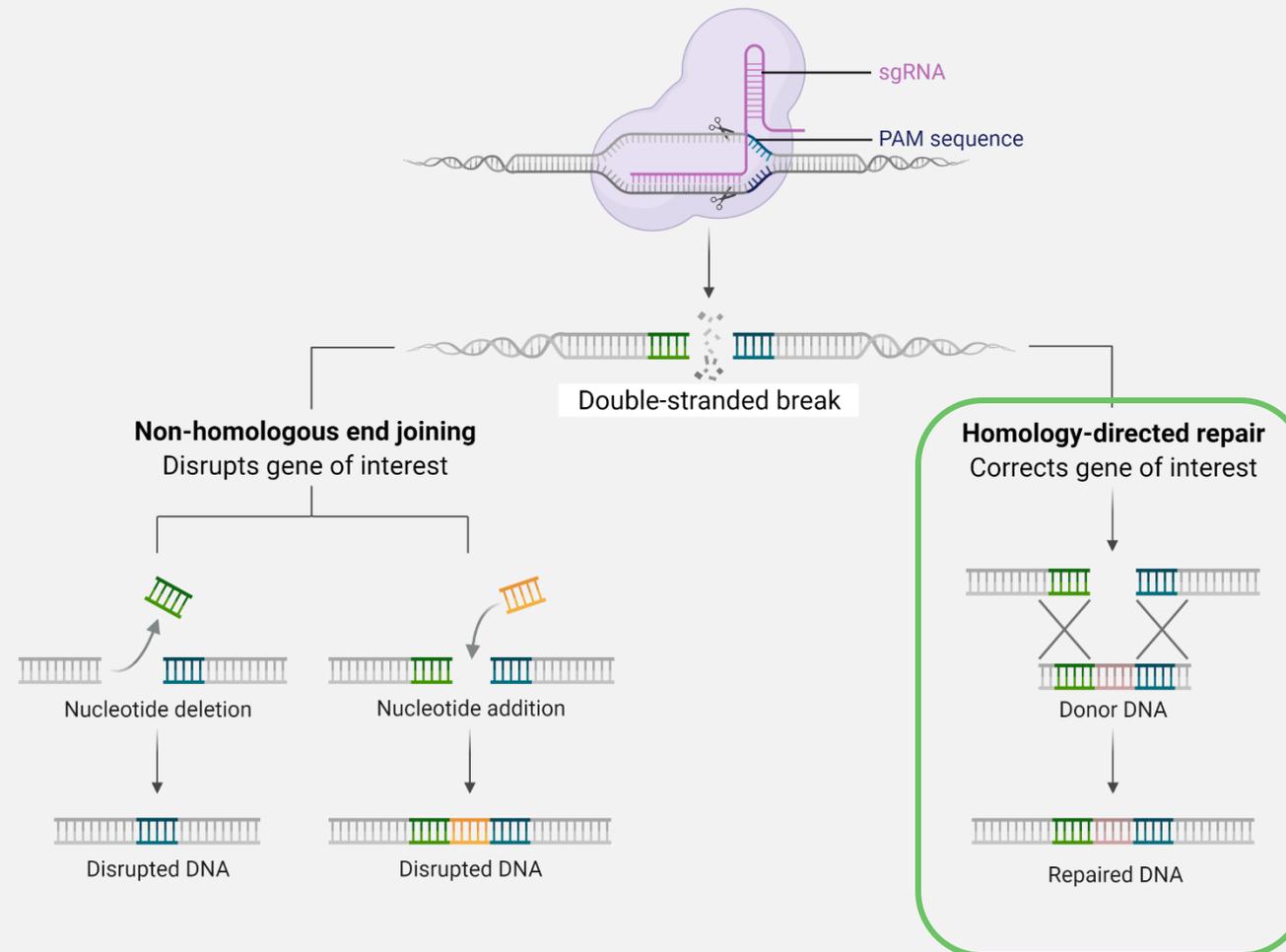
METHODS

CRISPR-Cas9 allows targeted genome editing



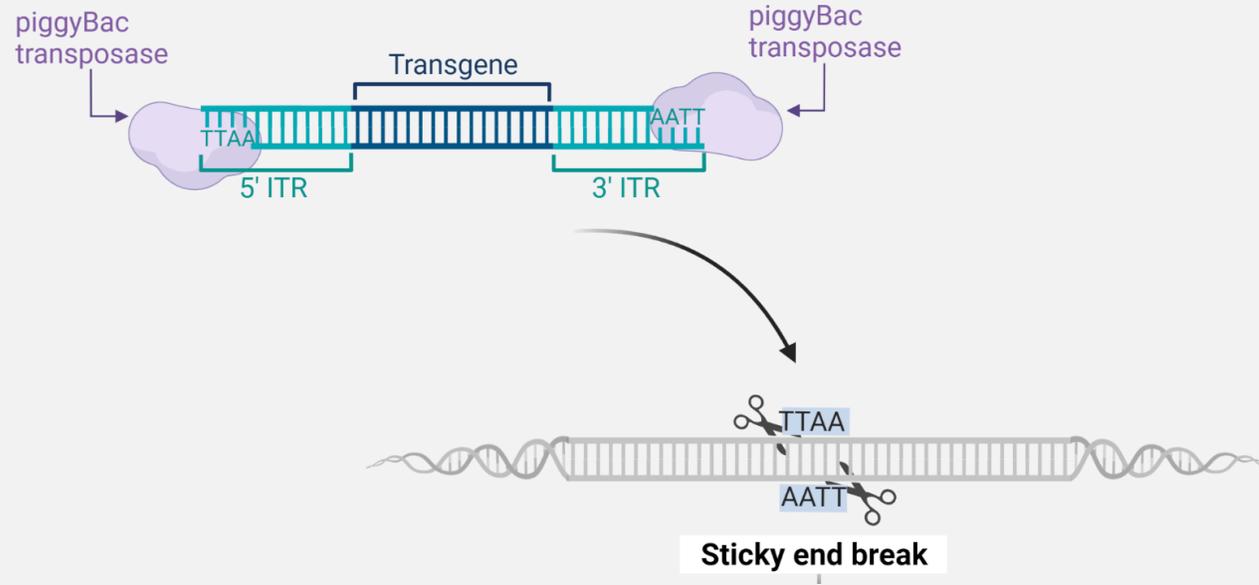
METHODS

CRISPR-Cas9 allows targeted genome editing



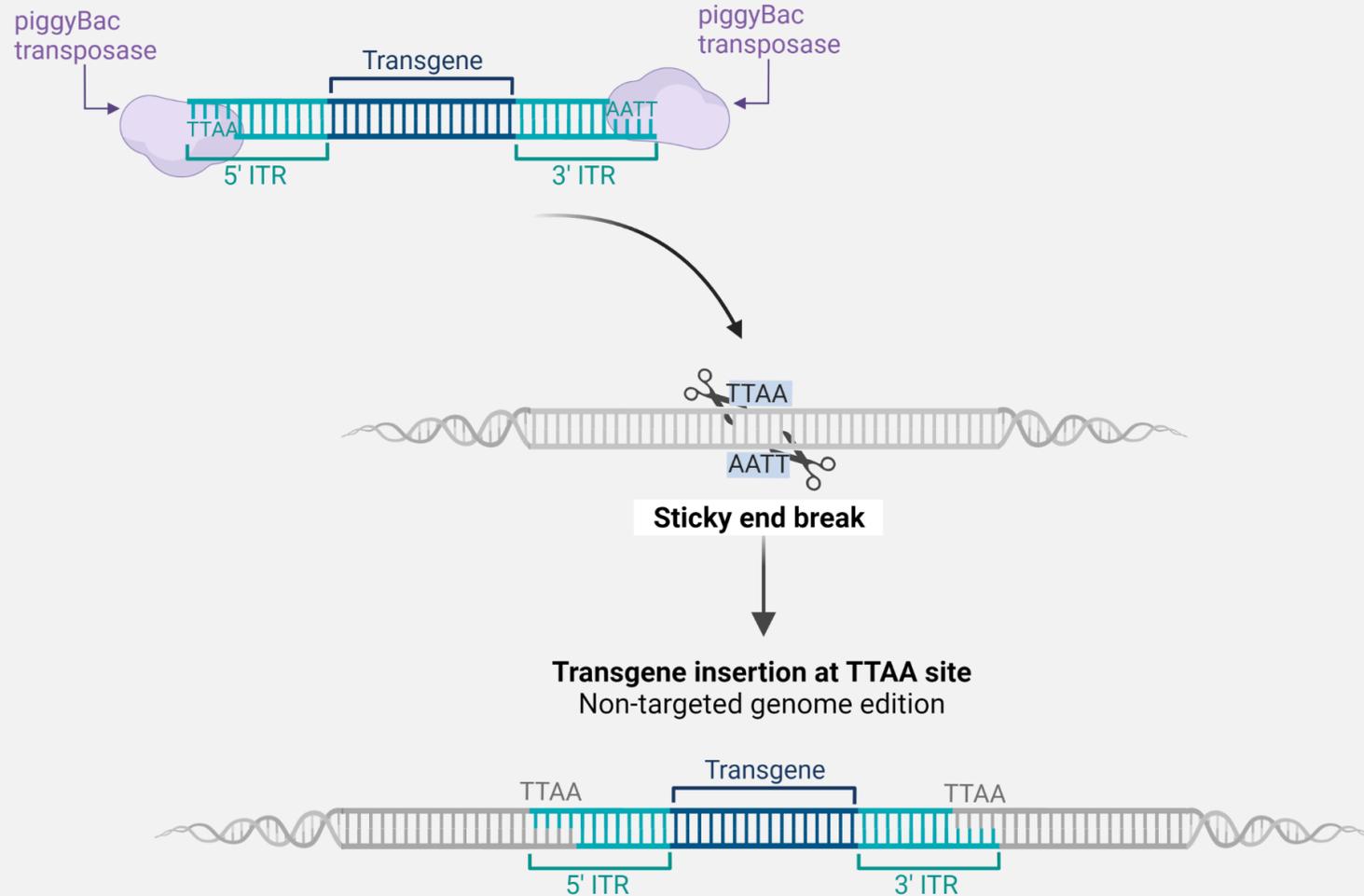
METHODS

piggyBac allows non-targeted genome editing



METHODS

piggyBac allows non-targeted genome editing



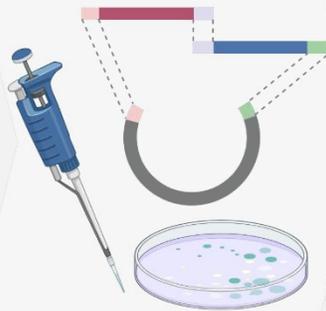
METHODS

Development of transgenic lines

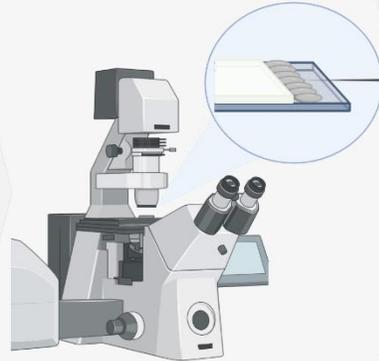
Design



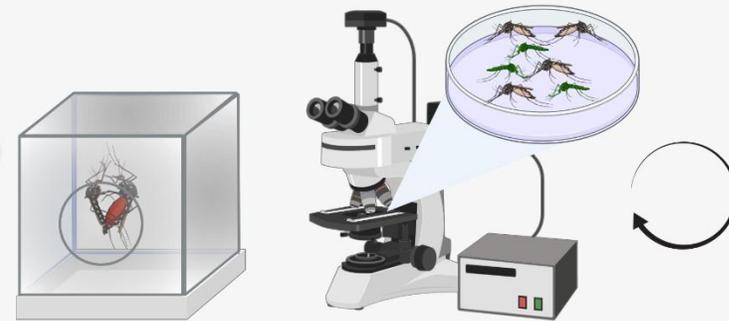
Cloning



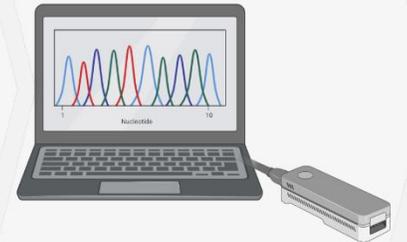
Microinjection



Crossing, Screening, Repeat



Sequencing / PCR



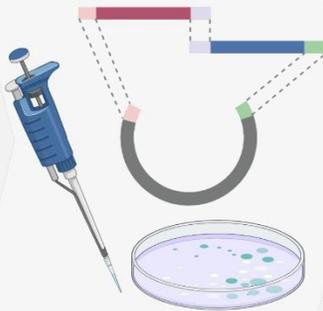
METHODS

Development of transgenic lines

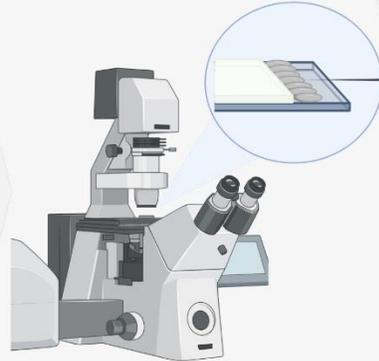
Design



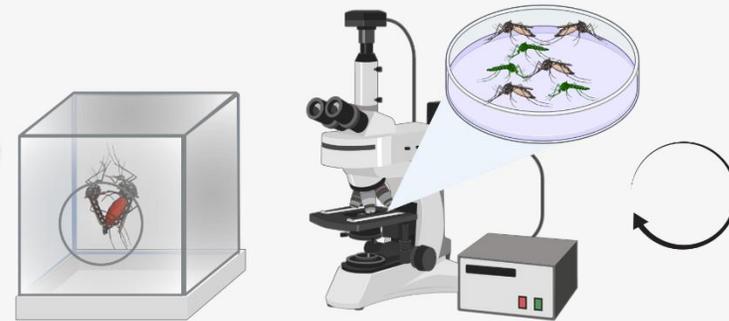
Cloning



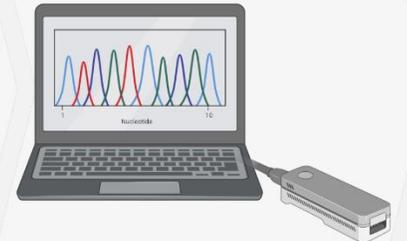
Microinjection



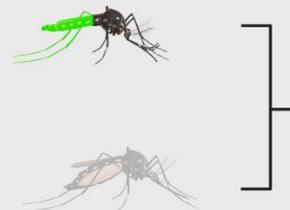
Crossing, Screening, Repeat



Sequencing / PCR



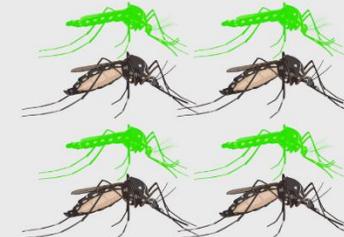
G0
Transient
expression



G1
Transgenic
individuals



G2
Transgene
segregation



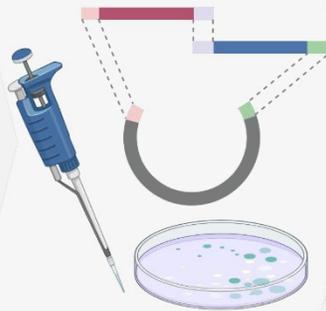
METHODS

Development of transgenic lines

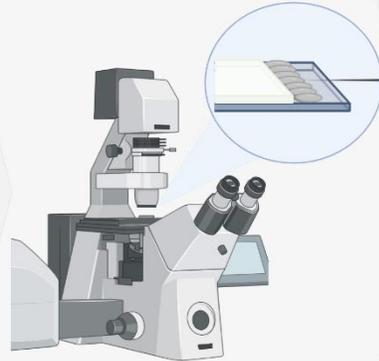
Design



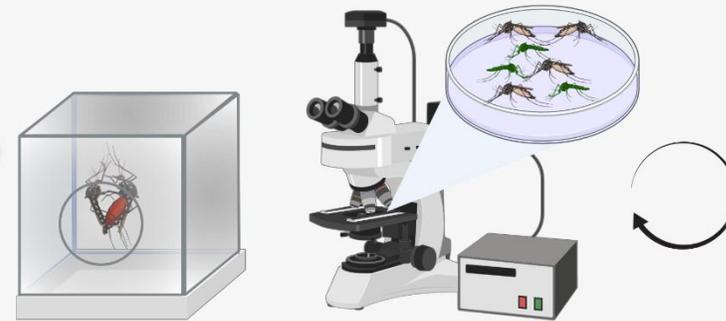
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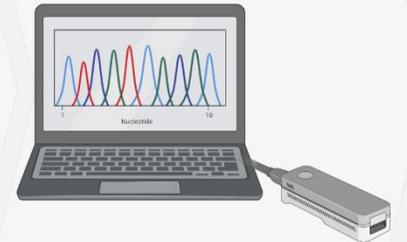
Microinjection



Crossing, Screening, Repeat



Sequencing / PCR



RESULTS

Developing GSS in *Aedes aegypti*

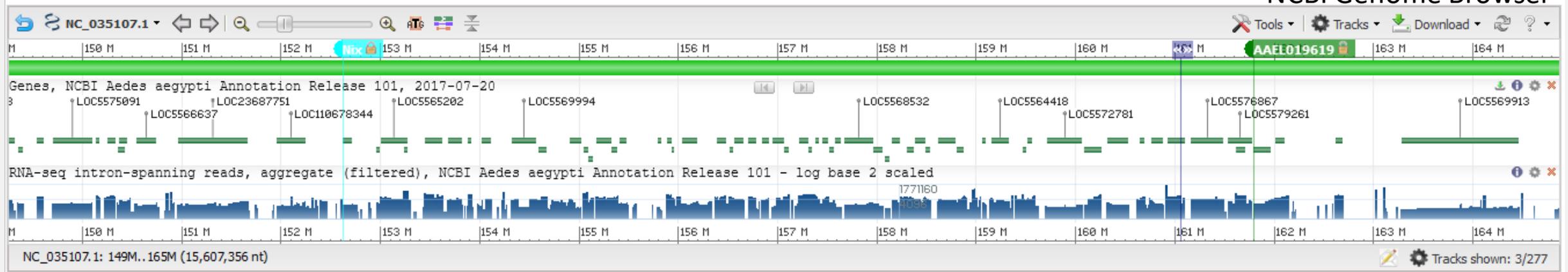


RESULTS – *Aedes Aegypti*

Target identification

Chromosome I

NCBI Genome Browser

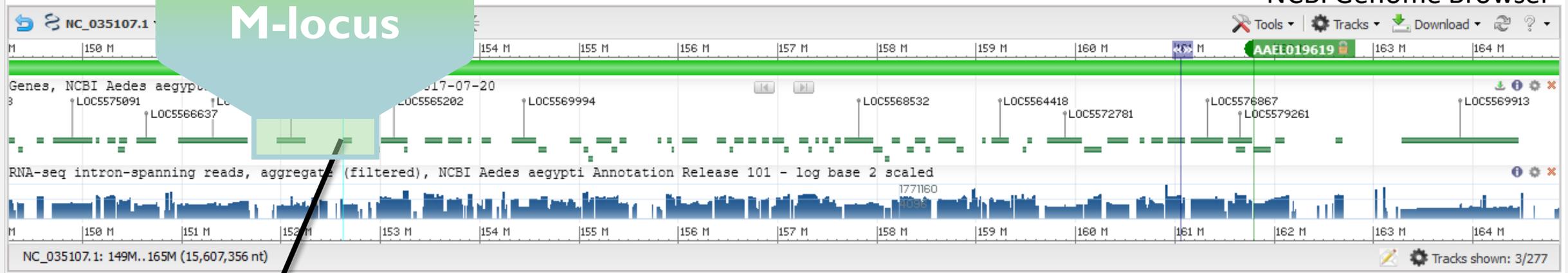


RESULTS – *Aedes Aegypti*

Target identification

Chromosome I

NCBI Genome Browser



M-locus

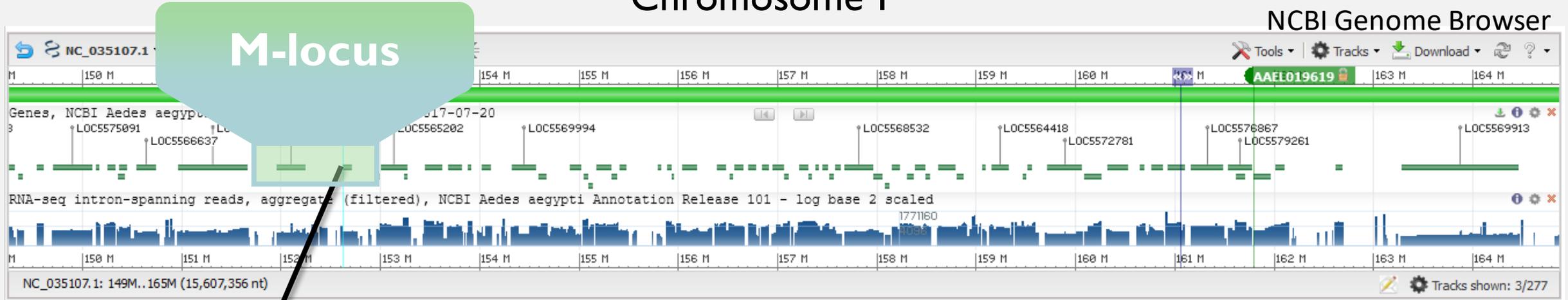
M-locus:
Nix

RESULTS – *Aedes Aegypti*

Target identification

Chromosome I

NCBI Genome Browser



M-locus

M-locus:
Nix

- in the intron

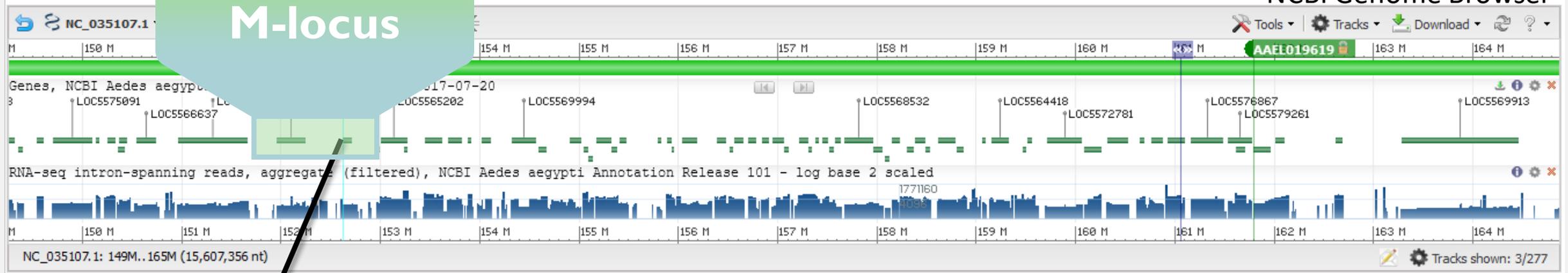


RESULTS – *Aedes Aegypti*

Target identification

Chromosome I

NCBI Genome Browser



M-locus

M-locus:
Nix

- in the intron
- as an artificial intron

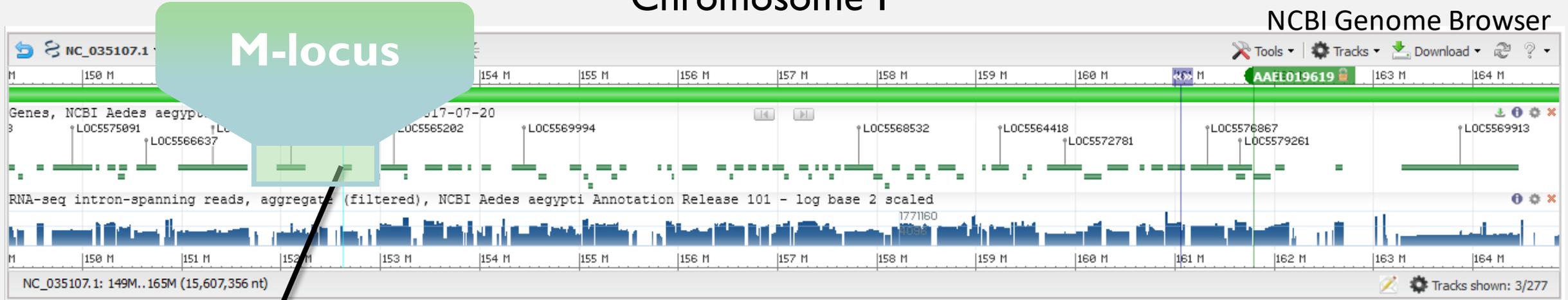


RESULTS – *Aedes Aegypti*

Target identification

Chromosome I

NCBI Genome Browser



M-locus

M-locus:
Nix

- in the intron
- as an artificial intron



Masculinization gene

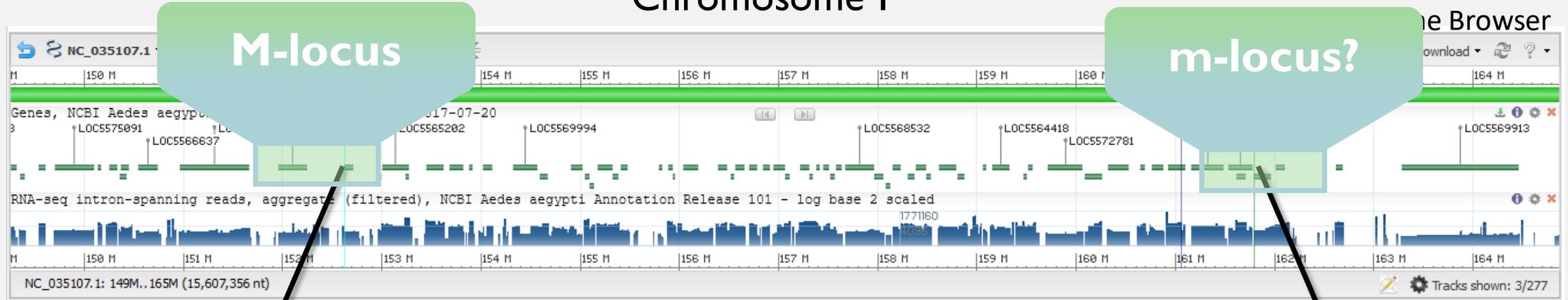
Central to the M-locus

Highest linkage between
maleness and fluorescence

RESULTS – *Aedes Aegypti*

Target identification

Chromosome I



M-locus:
Nix

- in the intron
- as an artificial intron

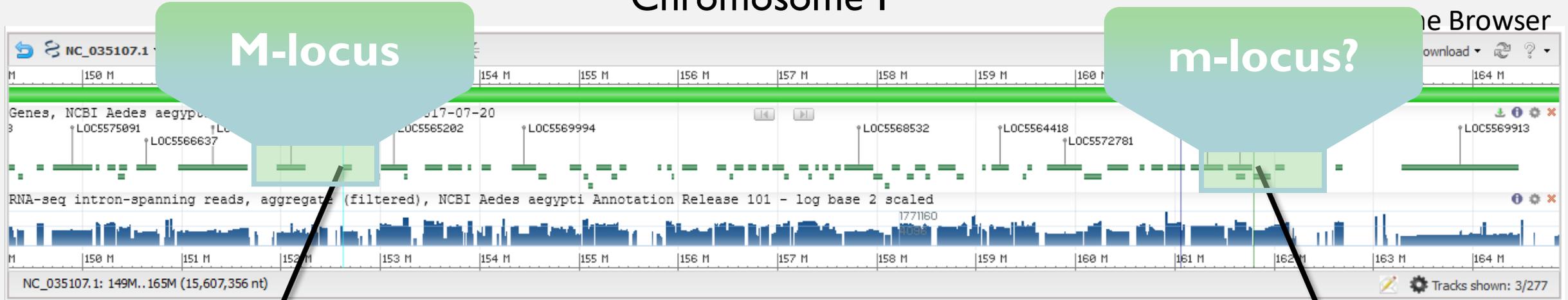
Gene linked to the m-locus
(chromosome quotient
analysis, A. Fontaine)
Probably misassembled
on the M-bearing
chromosome I

m-locus:
AAEL019619
- in an intron
- as an artificial intron

RESULTS – *Aedes Aegypti*

Target identification

Chromosome I



M-locus:
Nix

- in the intron
- as an artificial intron

m-locus:
AAEL019619

- in an intron
- as an artificial intron

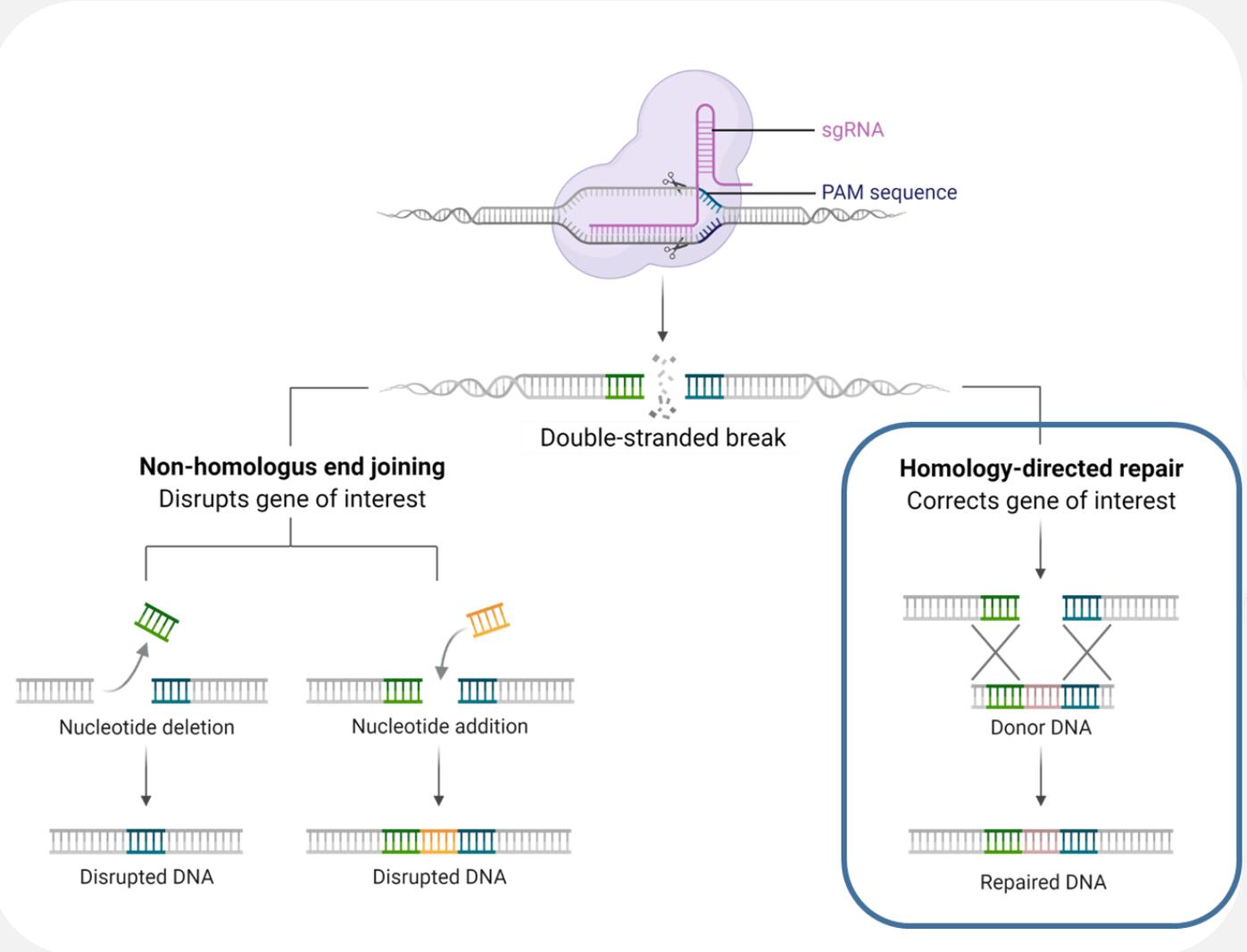
RESULTS – *Aedes Aegypti*

CRISPR-Cas9 approach

Design

Cloning

Microinjection



RESULTS – *Aedes Aegypti*

CRISPR-Cas9
approach



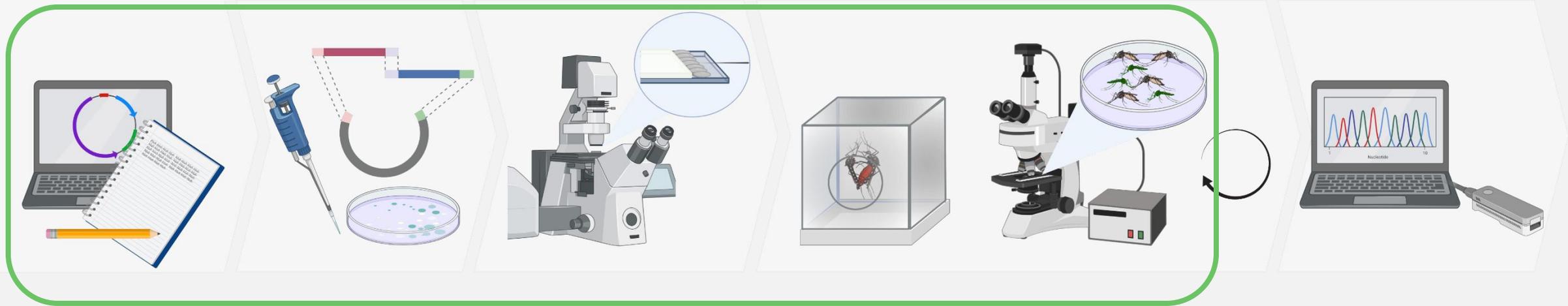
Design

Cloning

Microinjection

Crossing, Screening, Repeat

Sequencing / PCR



sgRNAs
+ repair plasmids

RESULTS – *Aedes Aegypti*

CRISPR-Cas9
approach

200 Cas9-
expressing eggs

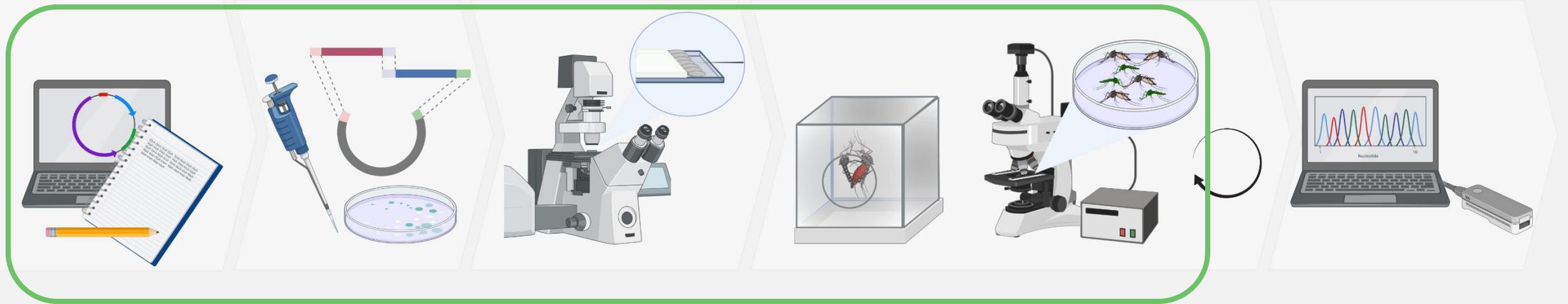
Design

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↑
sgRNAs
+ repair plasmids

RESULTS – *Aedes Aegypti*

CRISPR-Cas9
approach

200 Cas9-
expressing eggs

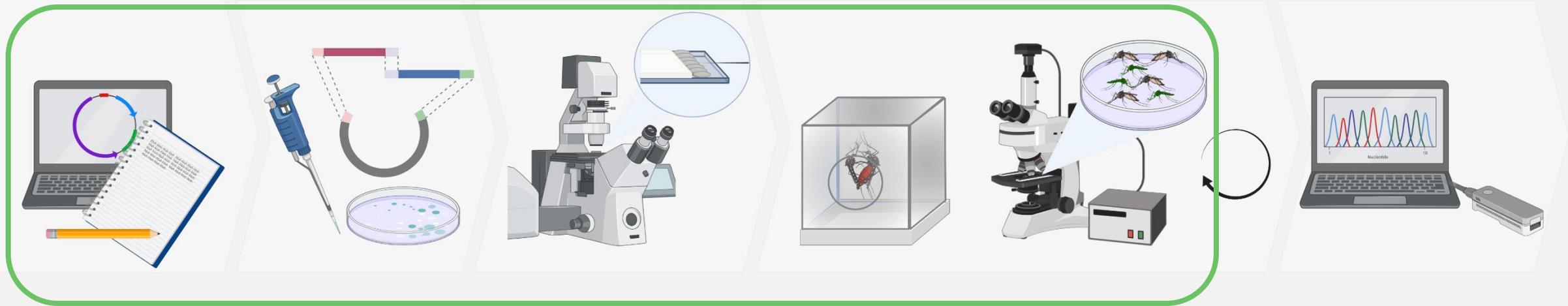
Design

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Sequencing / PCR



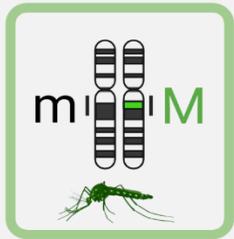
↑
sgRNAs
+ repair plasmids

↑
1 transgenic larva
in pooled GI!

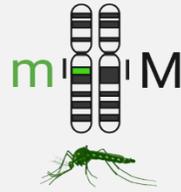
RESULTS – *Aedes Aegypti*

Interpretation

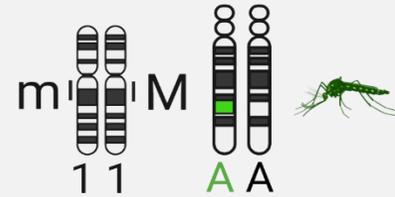
M-linked insertion



m-linked insertion



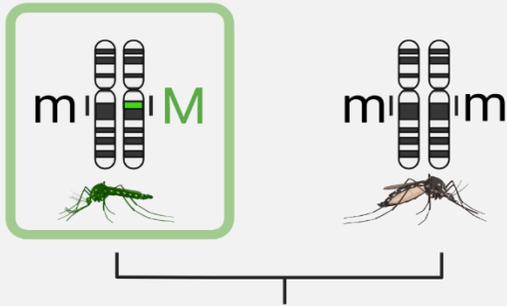
Autosomal insertion



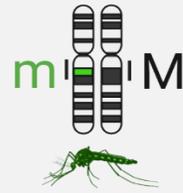
RESULTS – *Aedes Aegypti*

Interpretation

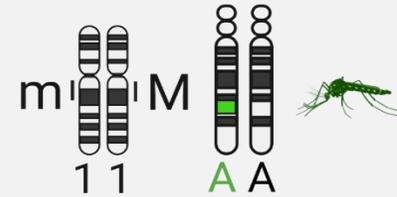
M-linked insertion



m-linked insertion



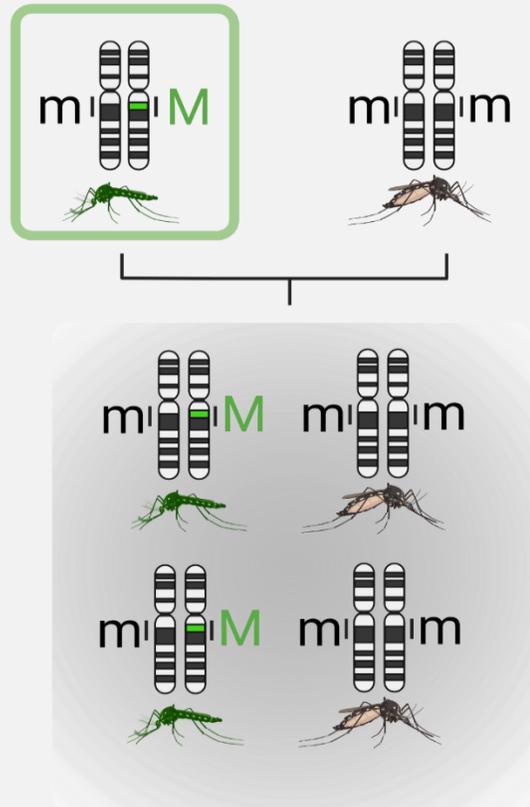
Autosomal insertion



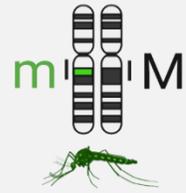
RESULTS – *AEDES AEGYPTI*

Interpretation

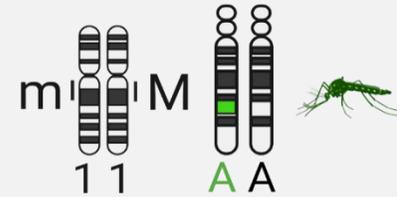
M-linked insertion



m-linked insertion



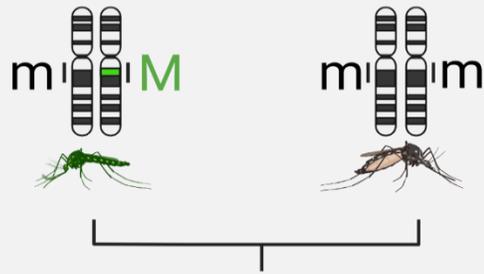
Autosomal insertion



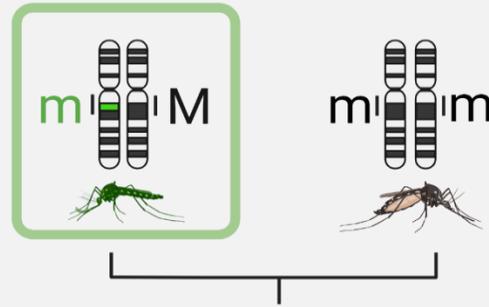
RESULTS – *Aedes Aegypti*

Interpretation

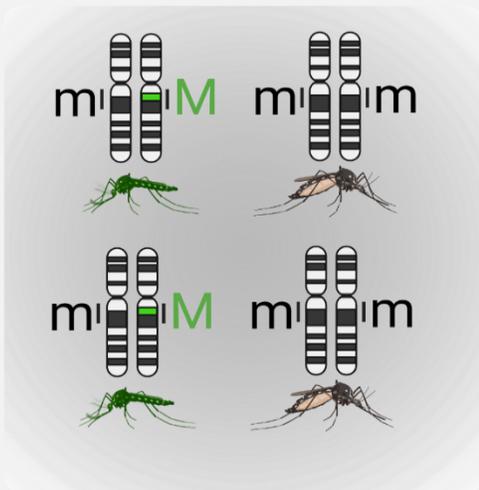
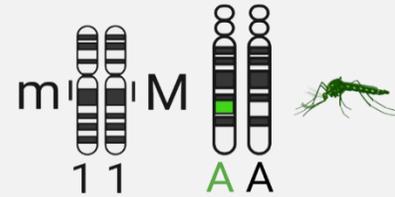
M-linked insertion



m-linked insertion



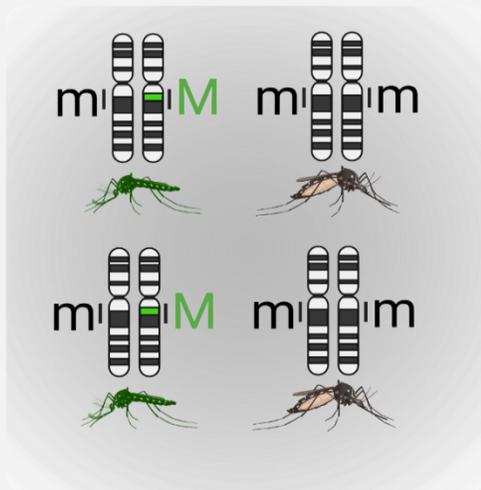
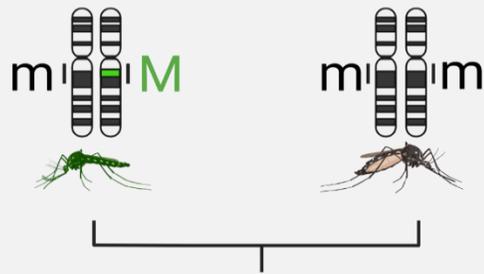
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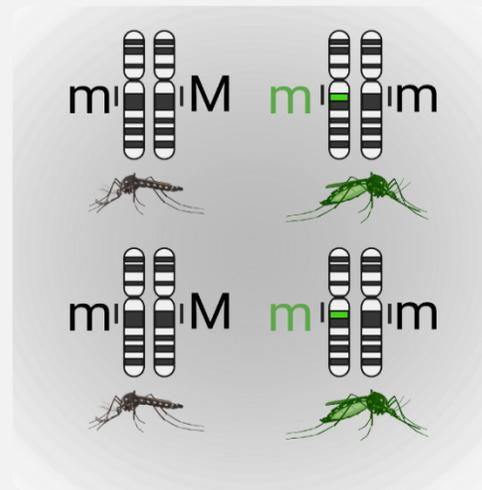
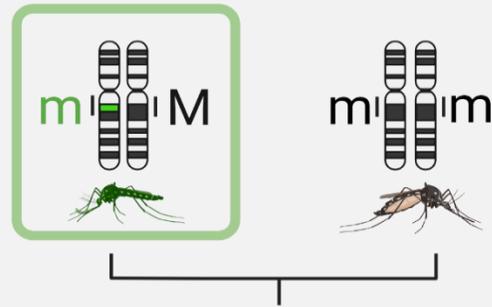
RESULTS – *AEDES AEGYPTI*

Interpretation

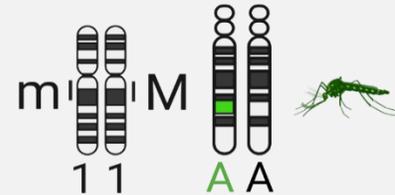
M-linked insertion



m-linked insertion



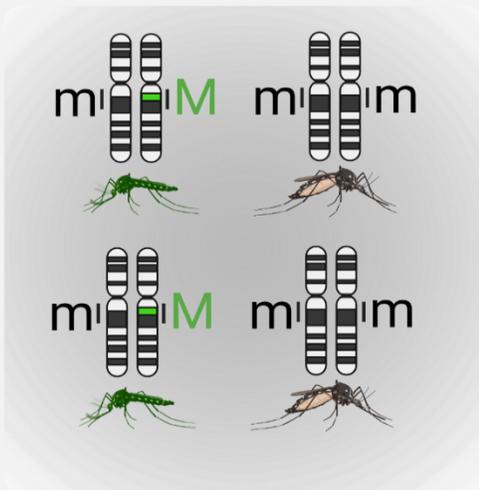
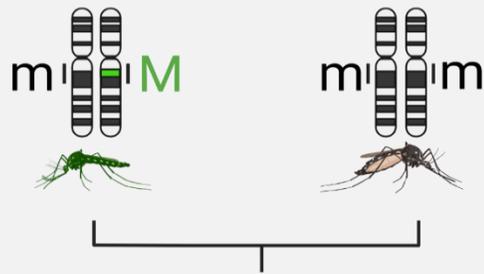
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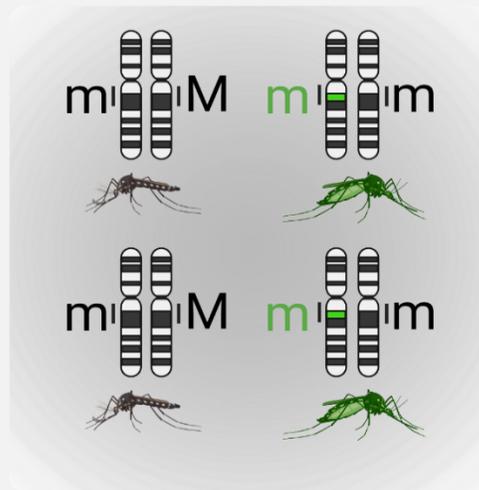
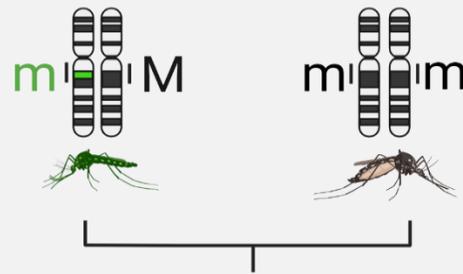
RESULTS – *AEDES AEGYPTI*

Interpretation

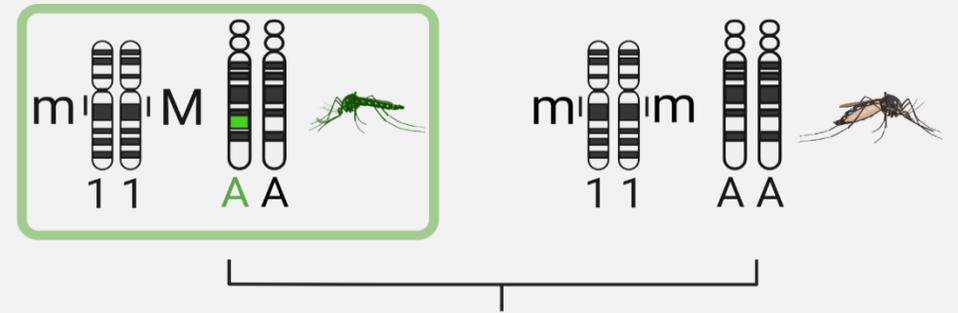
M-linked insertion



m-linked insertion



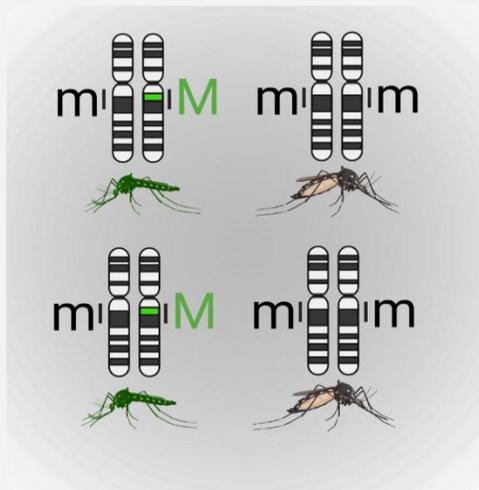
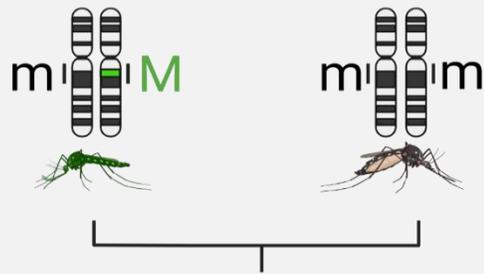
Autosomal insertion



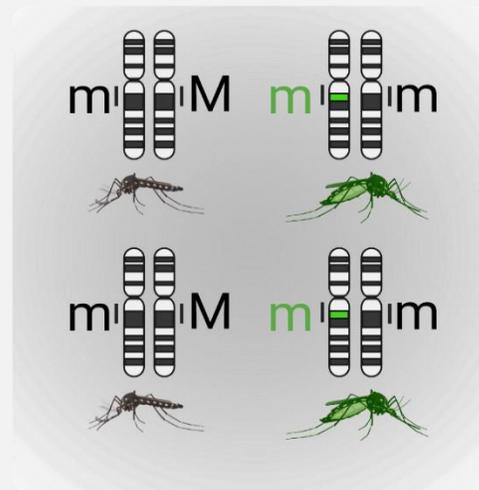
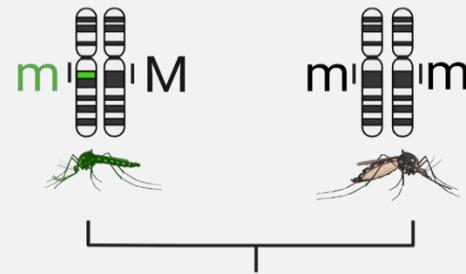
RESULTS – *Aedes Aegypti*

Interpretation

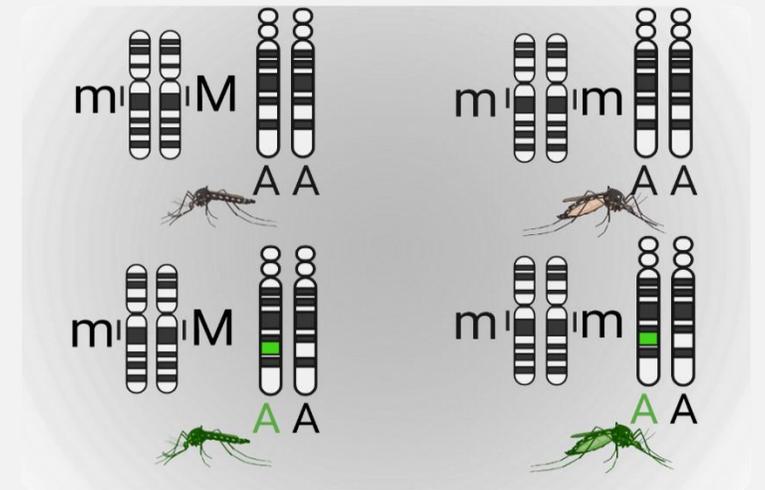
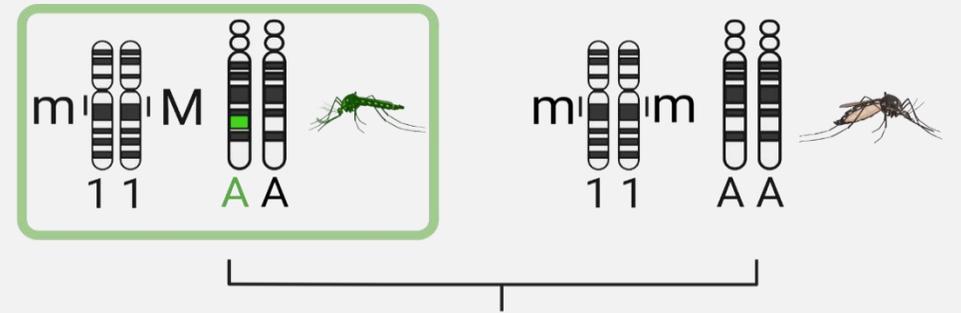
M-linked insertion



m-linked insertion



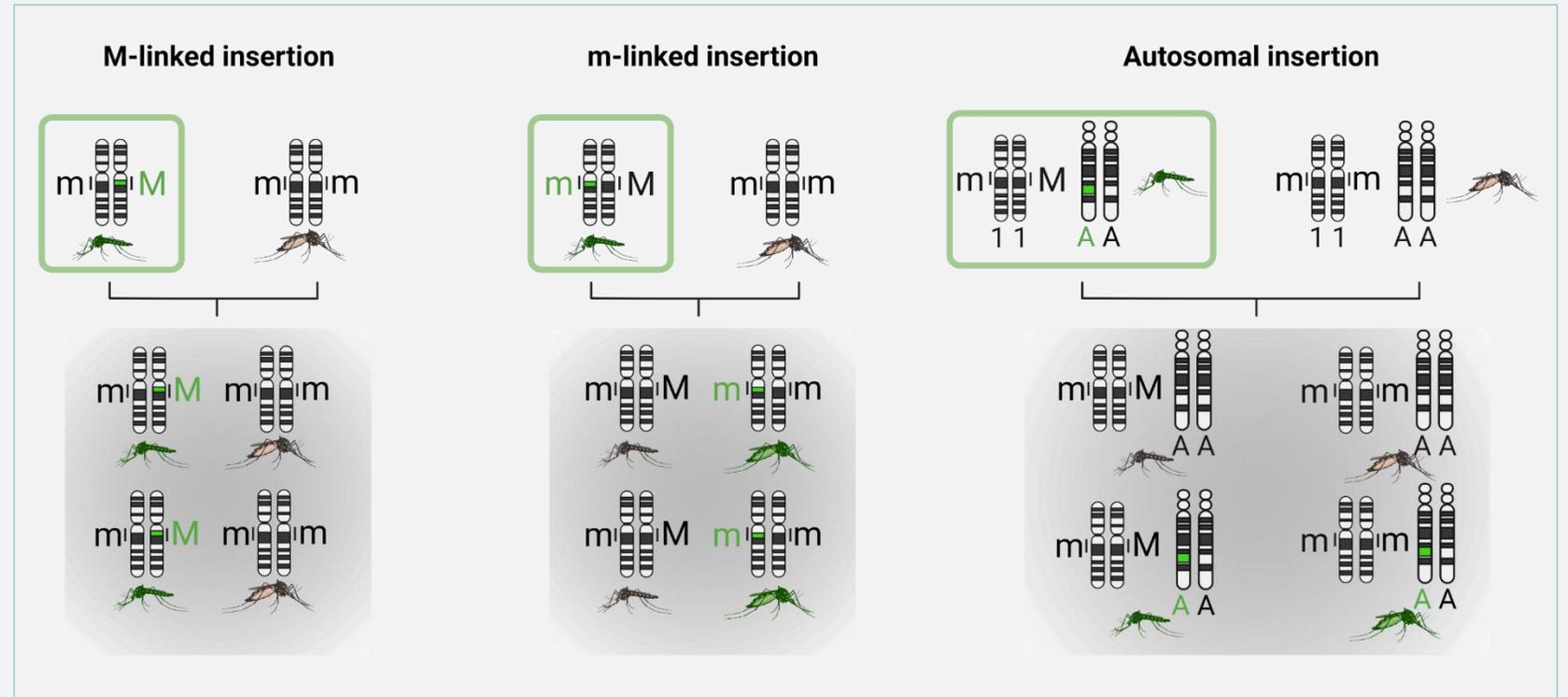
Autosomal insertion



RESULTS – *Aedes Aegypti*

Interpretation

Offspring
phenotype



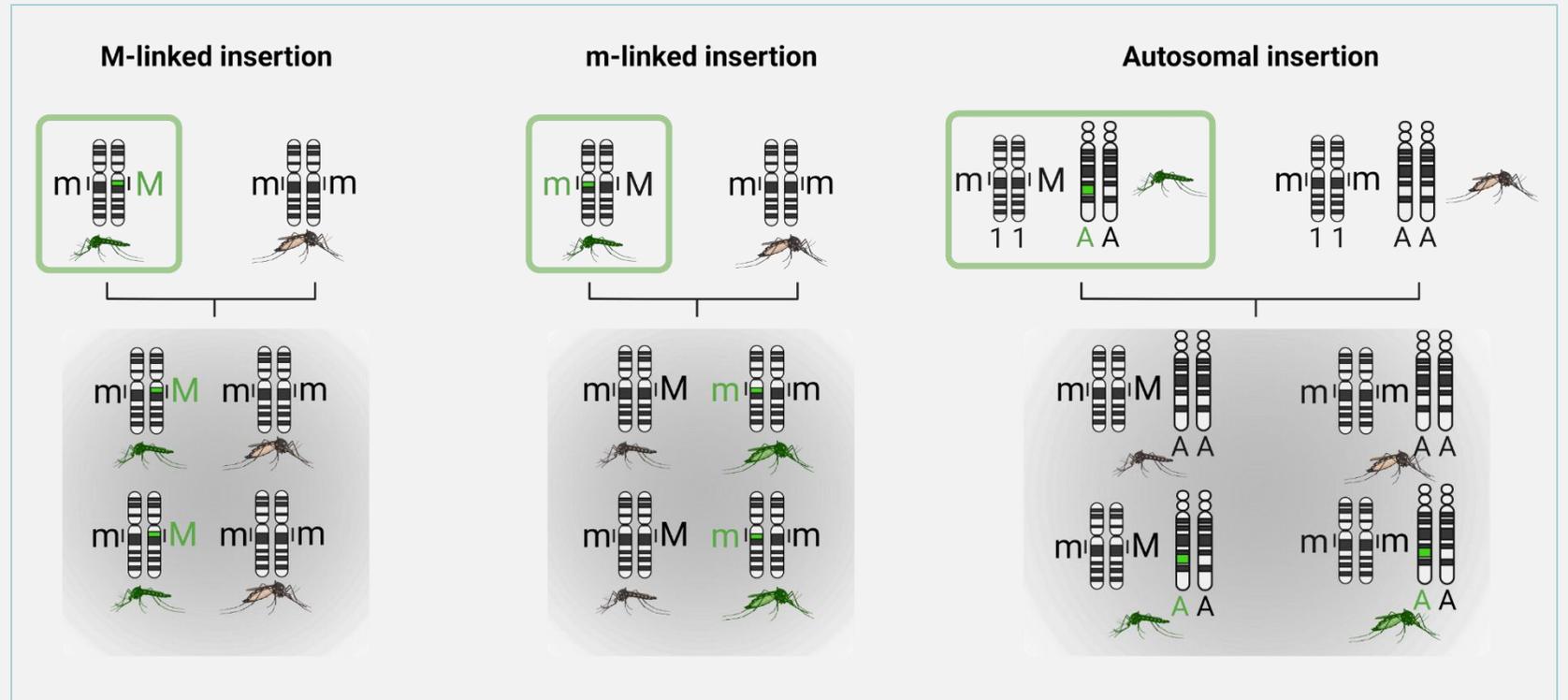
RESULTS – *Aedes Aegypti*

Interpretation

Father
genotype

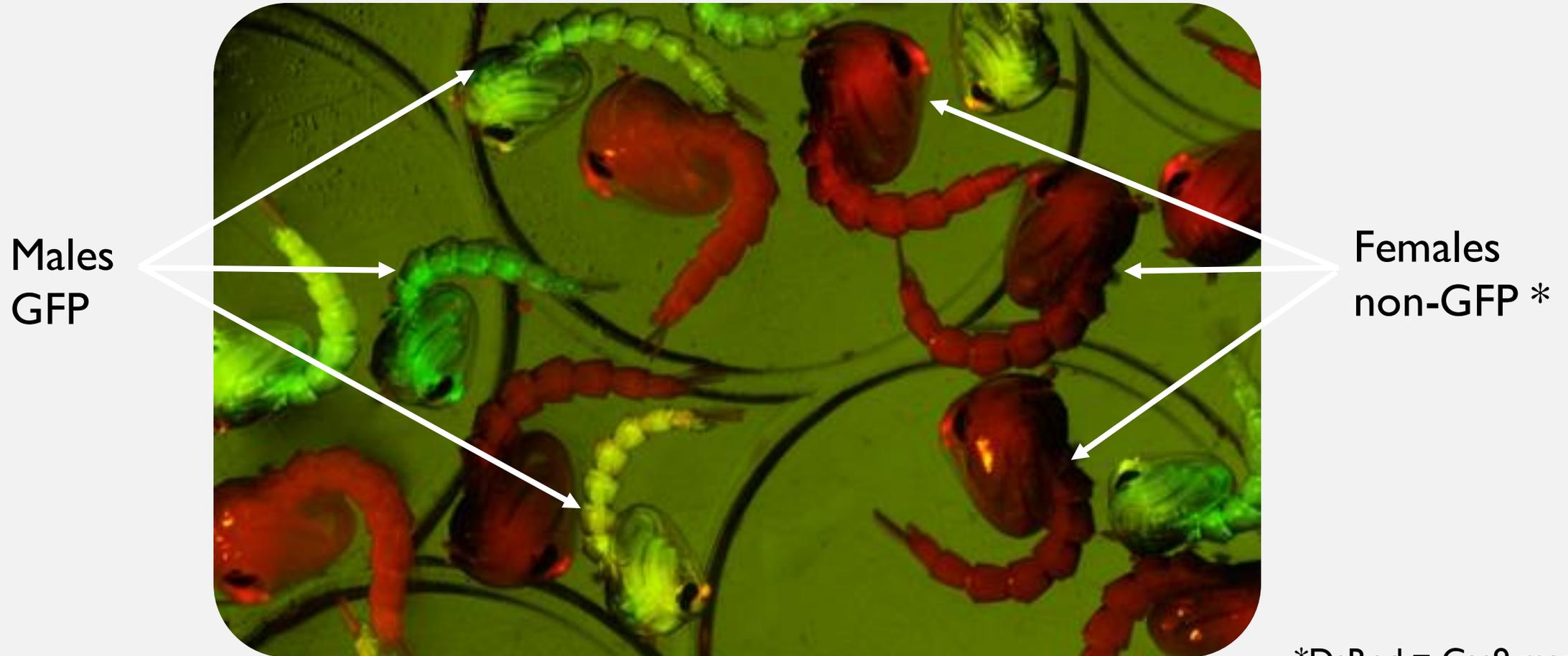


Offspring
phenotype



RESULTS – *Aedes Aegypti*

Obtaining of an M-linked line



*DsRed = Cas9 marker

All males express GFP → M-linkage

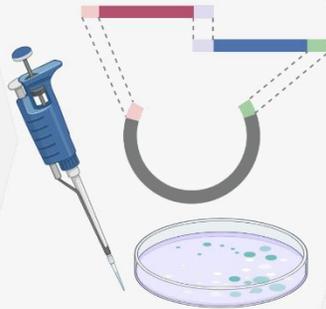
RESULTS – *Aedes Aegypti*

Obtaining of an M-linked line

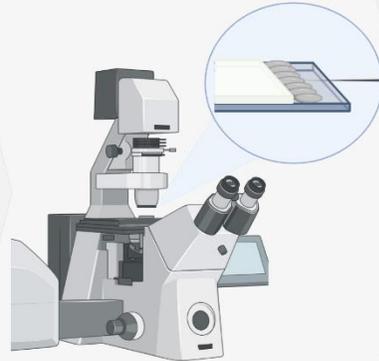
Design



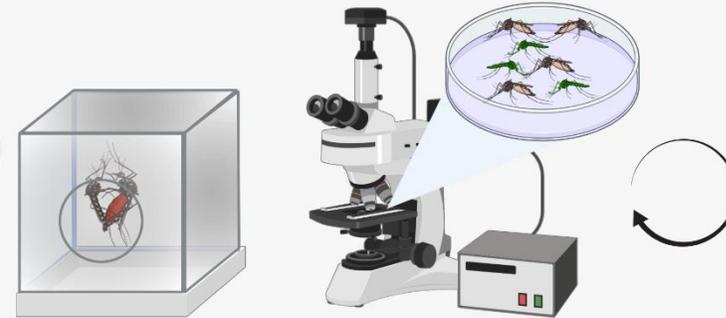
Cloning



Microinjection

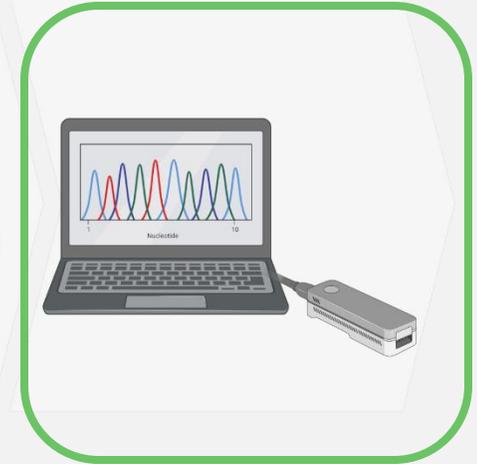


Crossing, Screening, Repeat



PCR failed
→ sequencing

Sequencing / PCR



RESULTS – *Aedes Aegypti*

Obtaining of an M-linked line

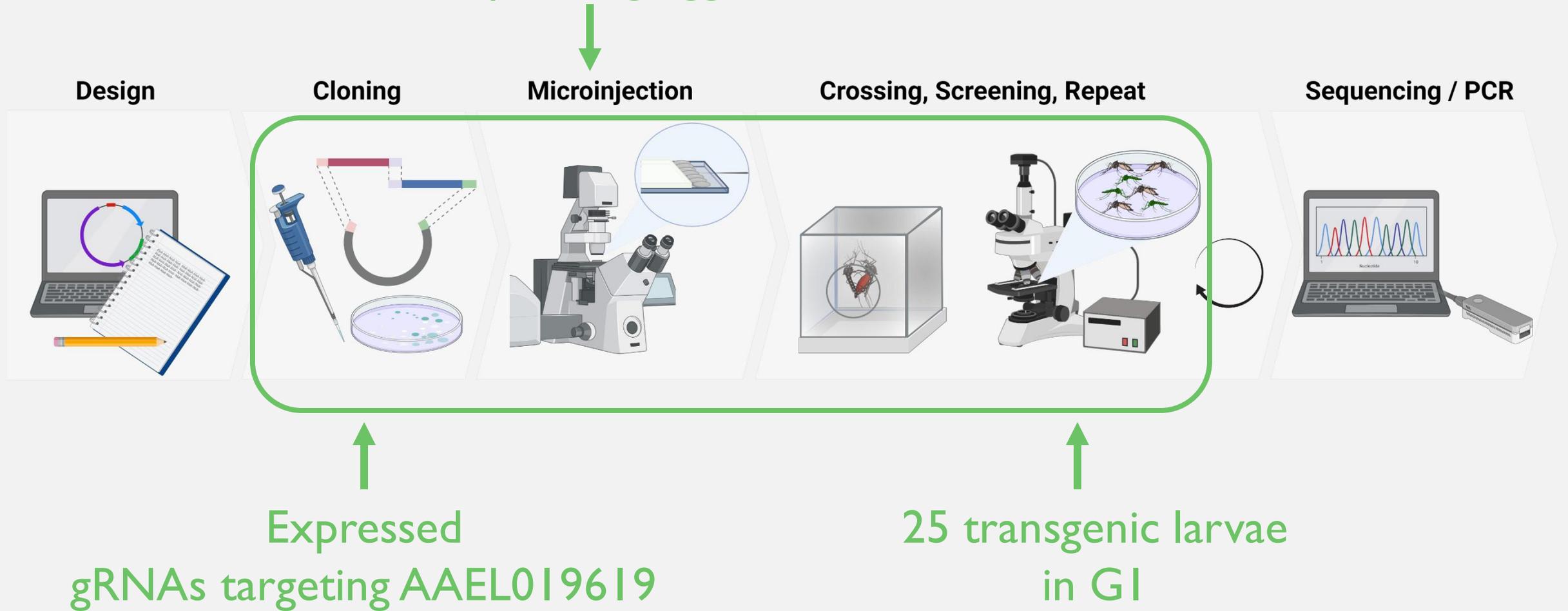


Controlled integration site:
it's AAEL019619 !

All males express GFP → M-linkage

RESULTS – *Aedes Aegypti*

2nd round AAEL019619
600 Cas9-
expressing eggs

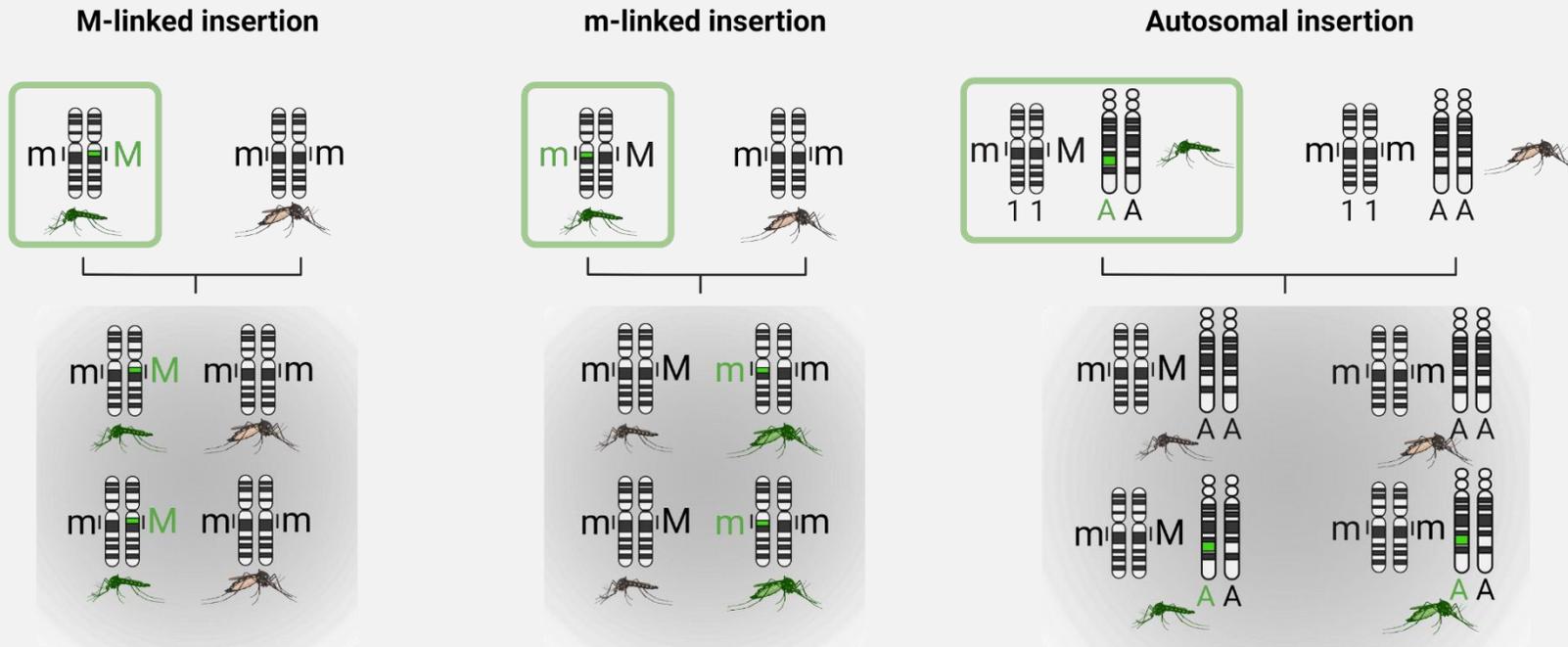


RESULTS – *Aedes Aegypti*

2nd round AAEL019619

14 transgenic fathers

11 transgenic mothers

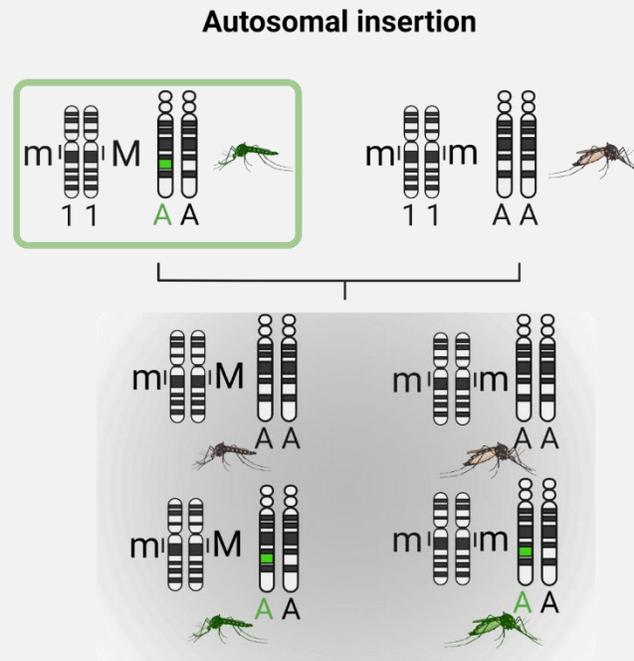
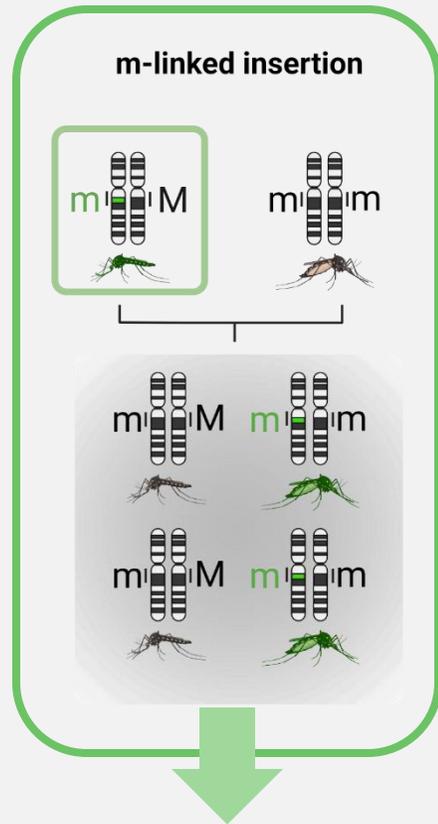
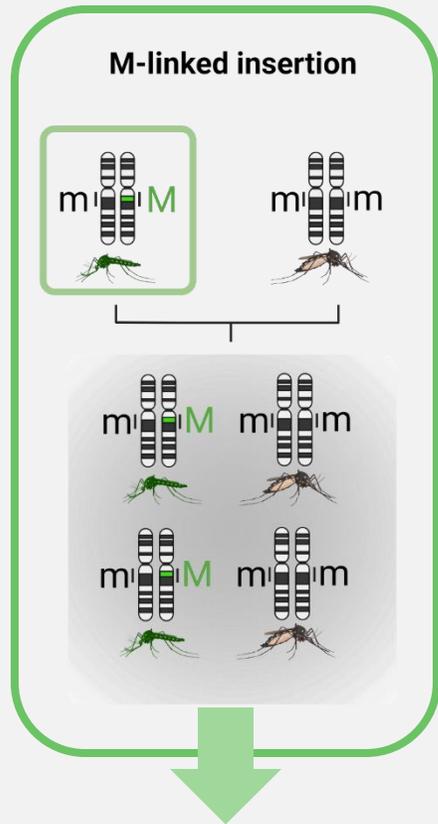


RESULTS - *Aedes Aegypti*

2nd round AAEL019619

14 transgenic fathers

11 transgenic mothers

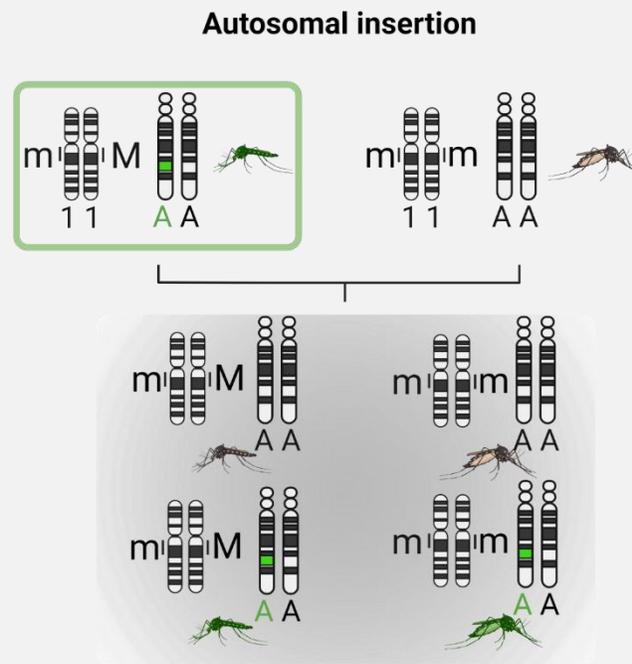
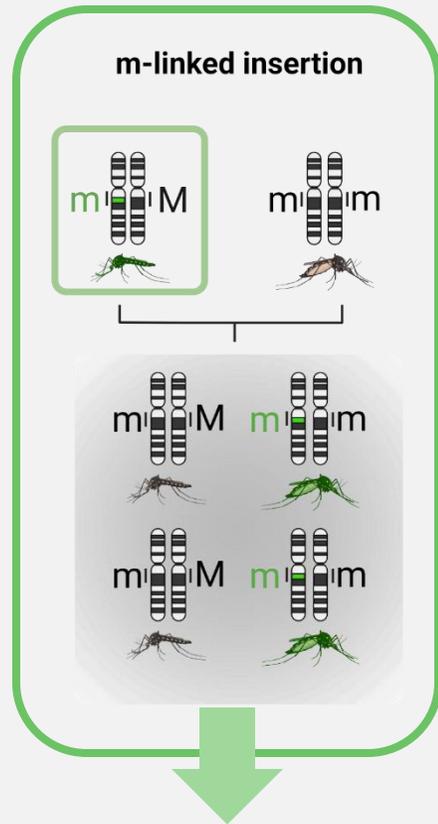
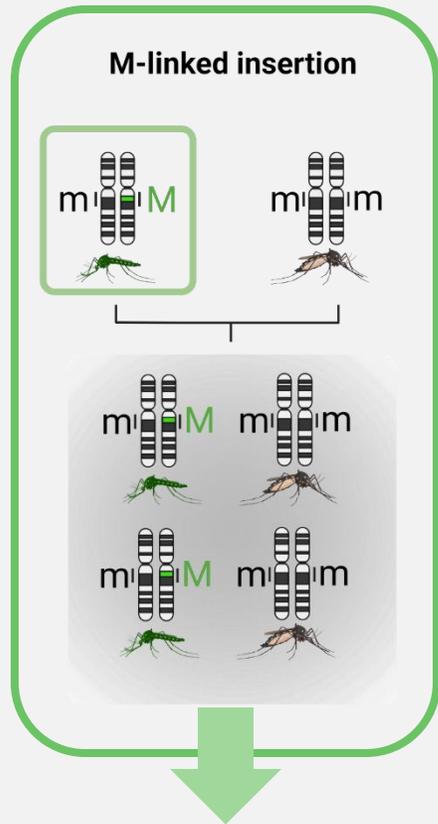


RESULTS – *Aedes Aegypti*

2nd round AAEL019619

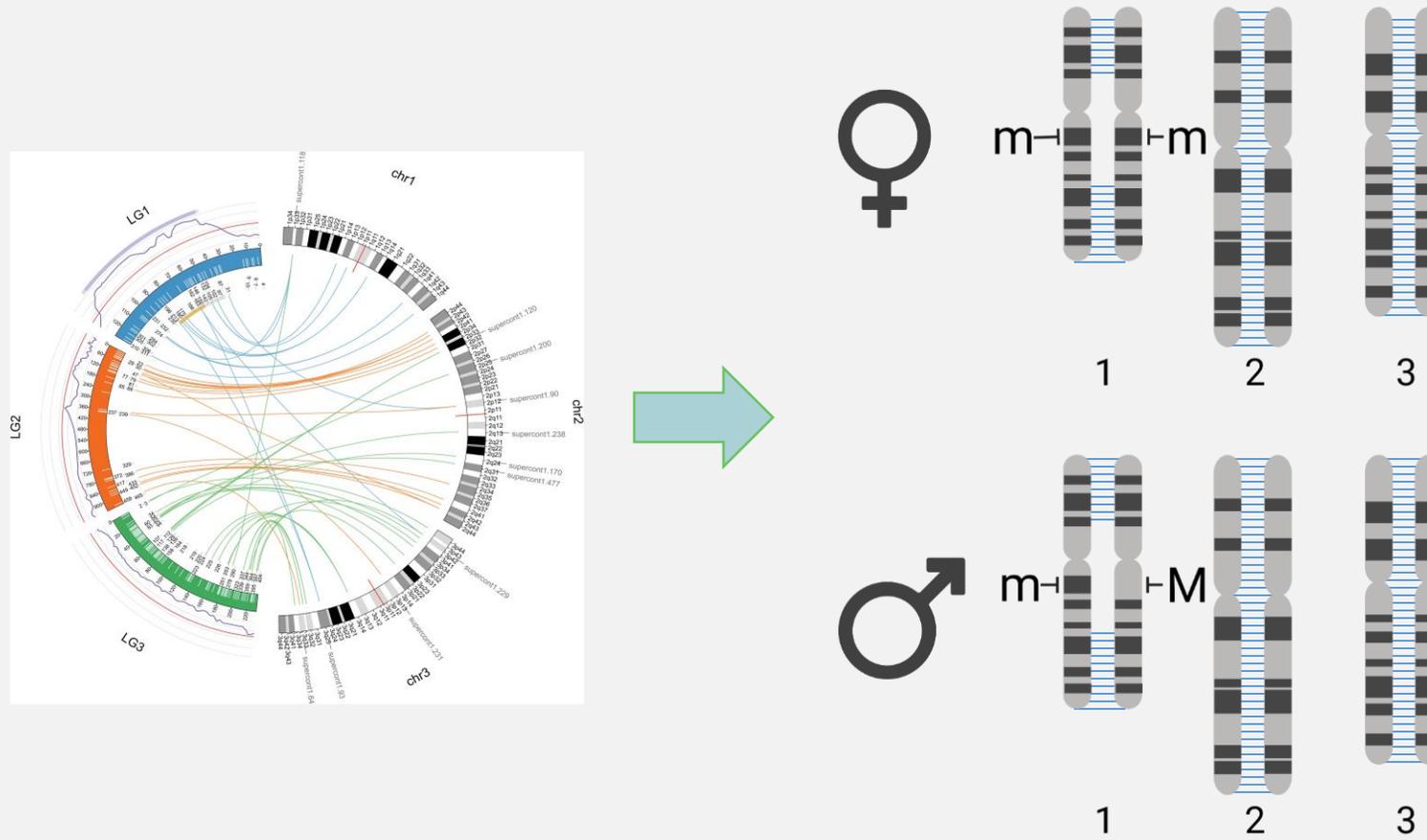
14 transgenic fathers

11 transgenic mothers



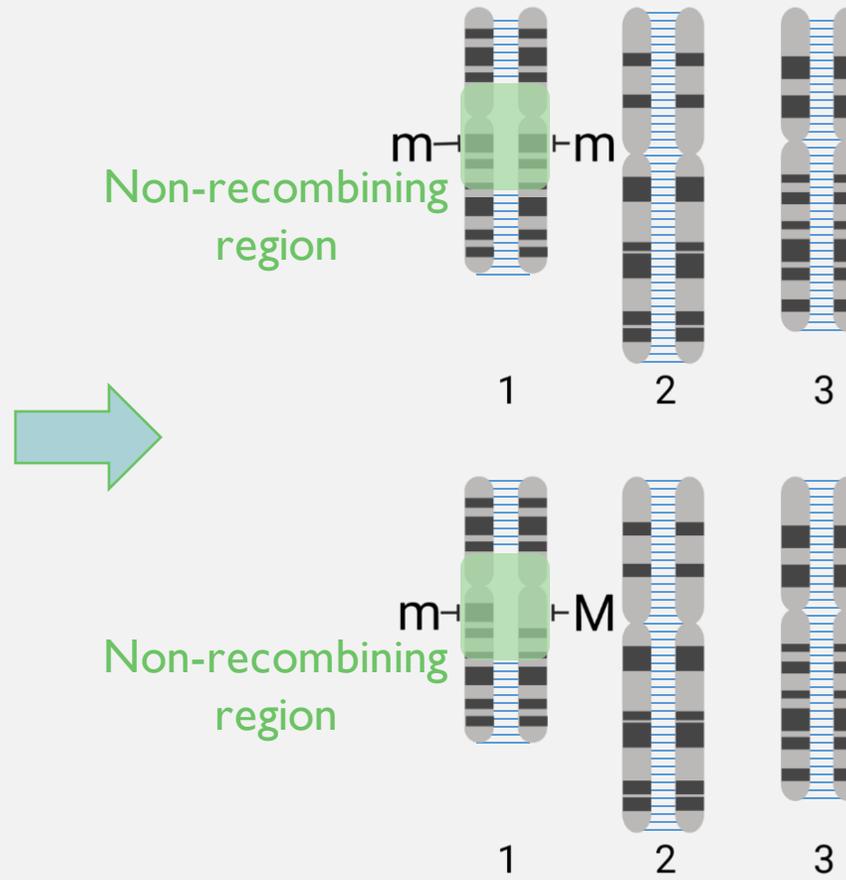
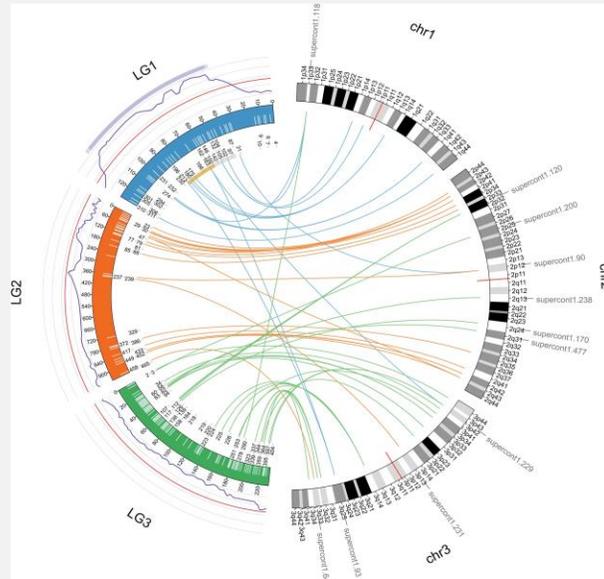
RESULTS – *Aedes Aegypti*

AAEL019619 is located at the neighborhood of both sex loci



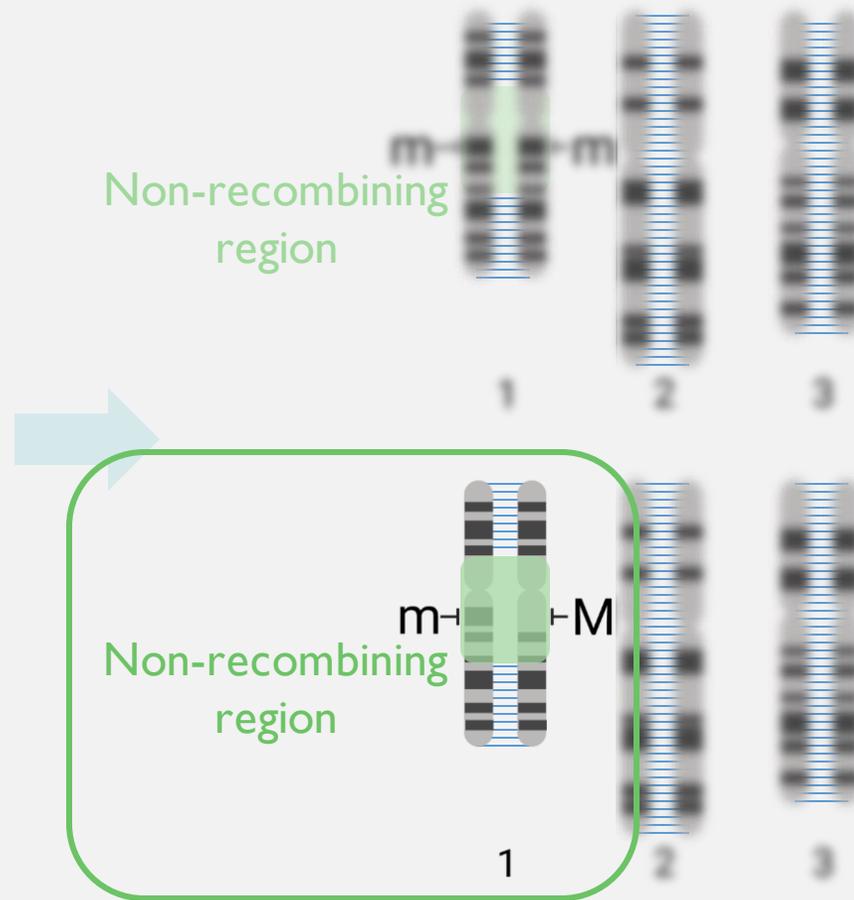
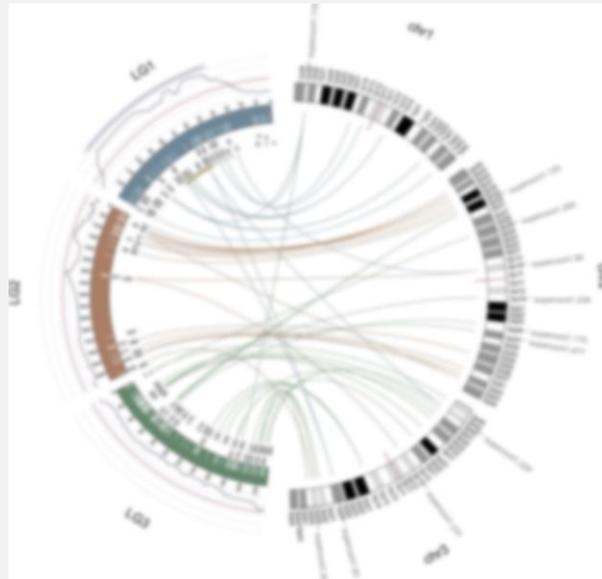
RESULTS – *Aedes Aegypti*

AAEL019619 is located at the neighborhood of both sex loci



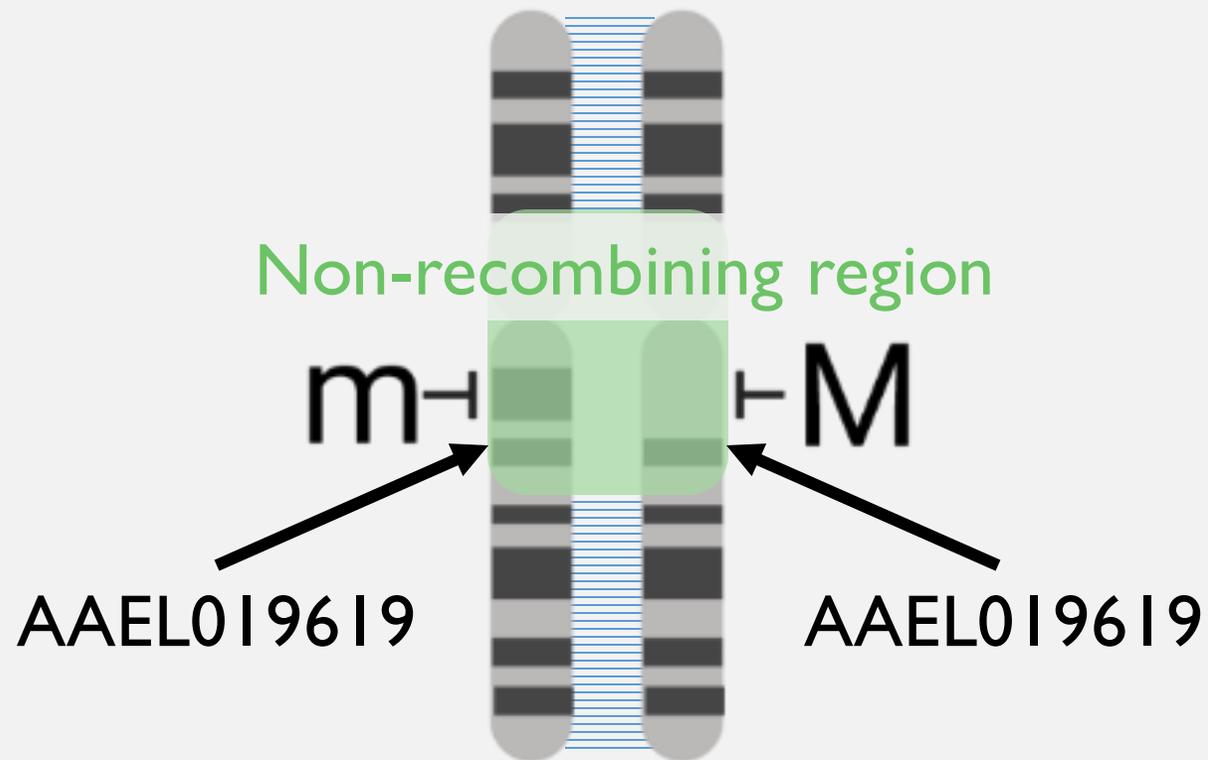
RESULTS – *Aedes Aegypti*

AAEL019619 is located at the neighborhood of both sex loci



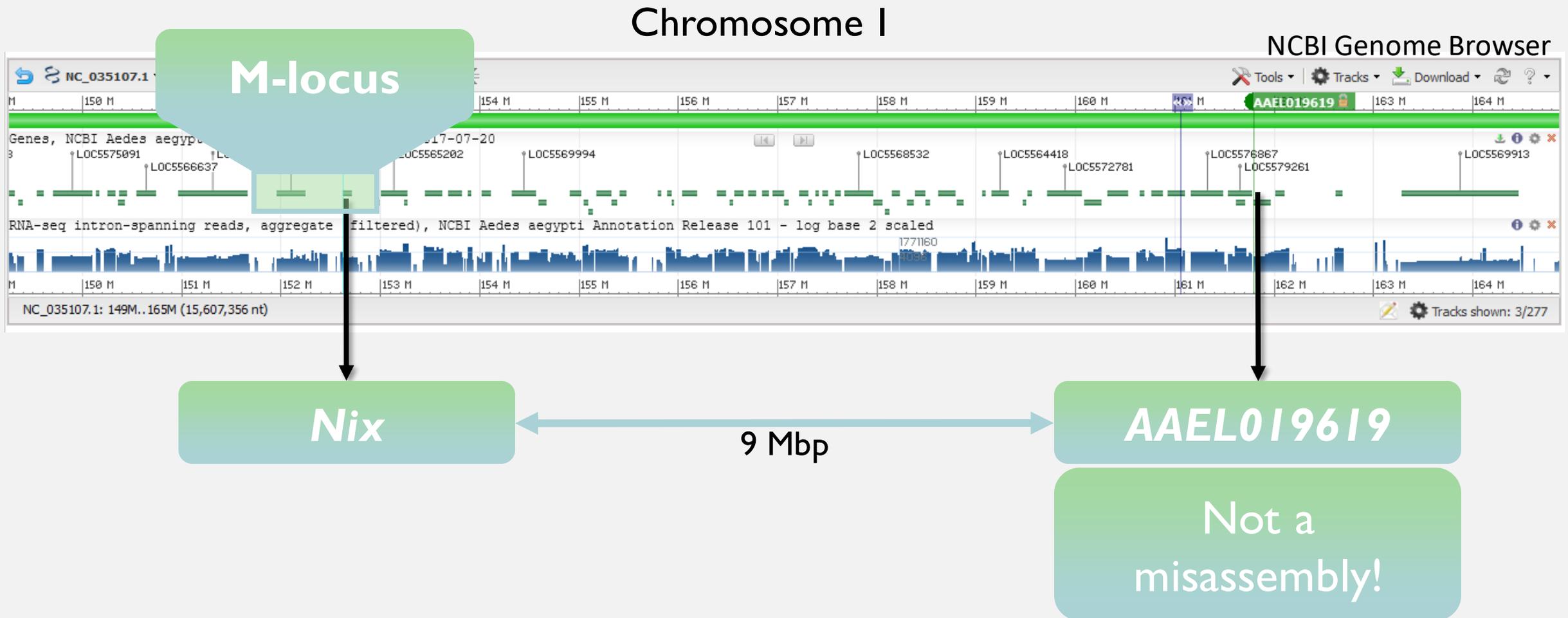
RESULTS – *Aedes Aegypti*

AAEL019619 is located in the non-recombining region near both sex loci



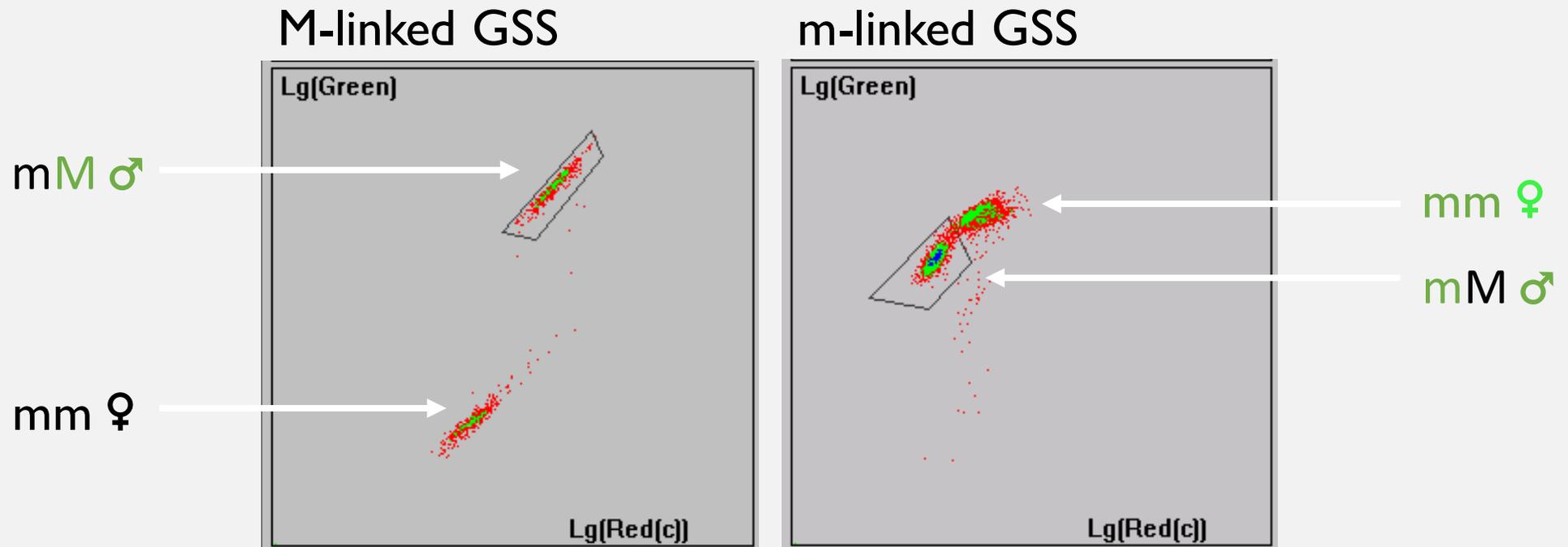
RESULTS – *Aedes Aegypti*

AAEL019619 is located at the neighborhood of both sex loci



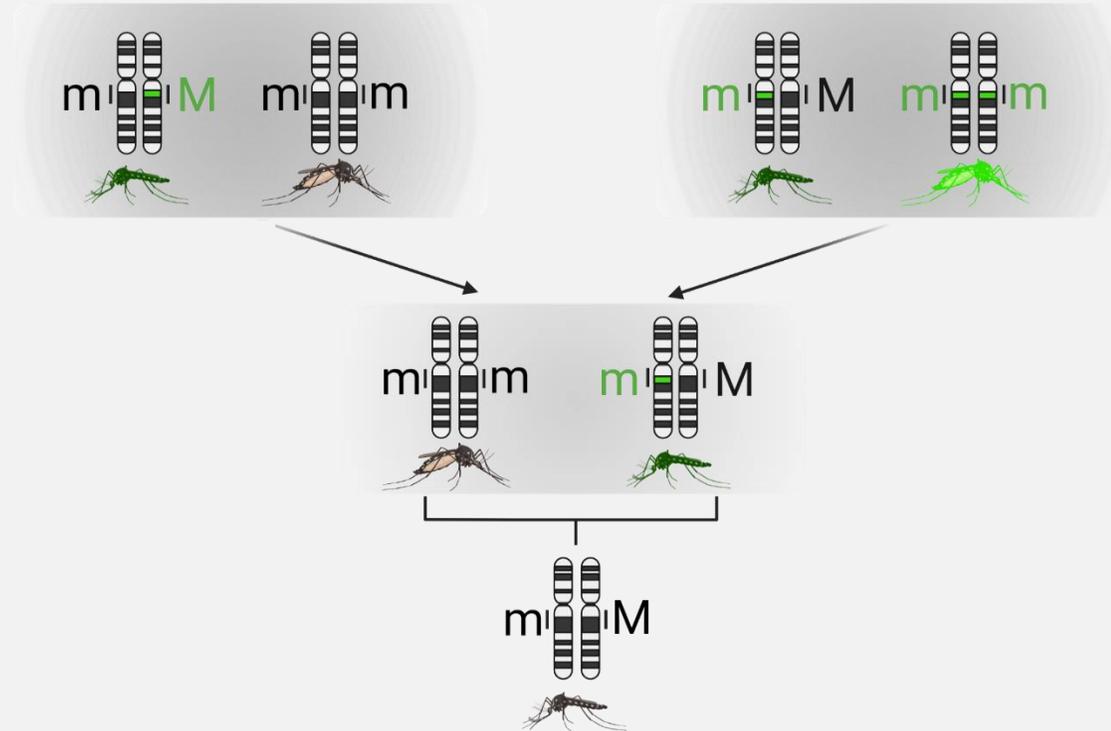
RESULTS – *Aedes Aegypti*

Both the M and m-linked lines can be sorted by COPAS



RESULTS – *Aedes Aegypti*

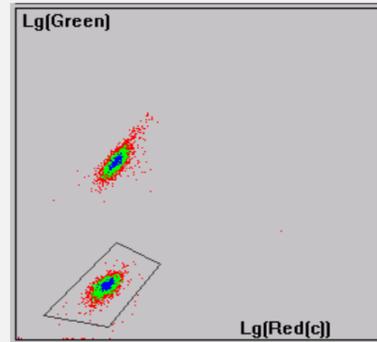
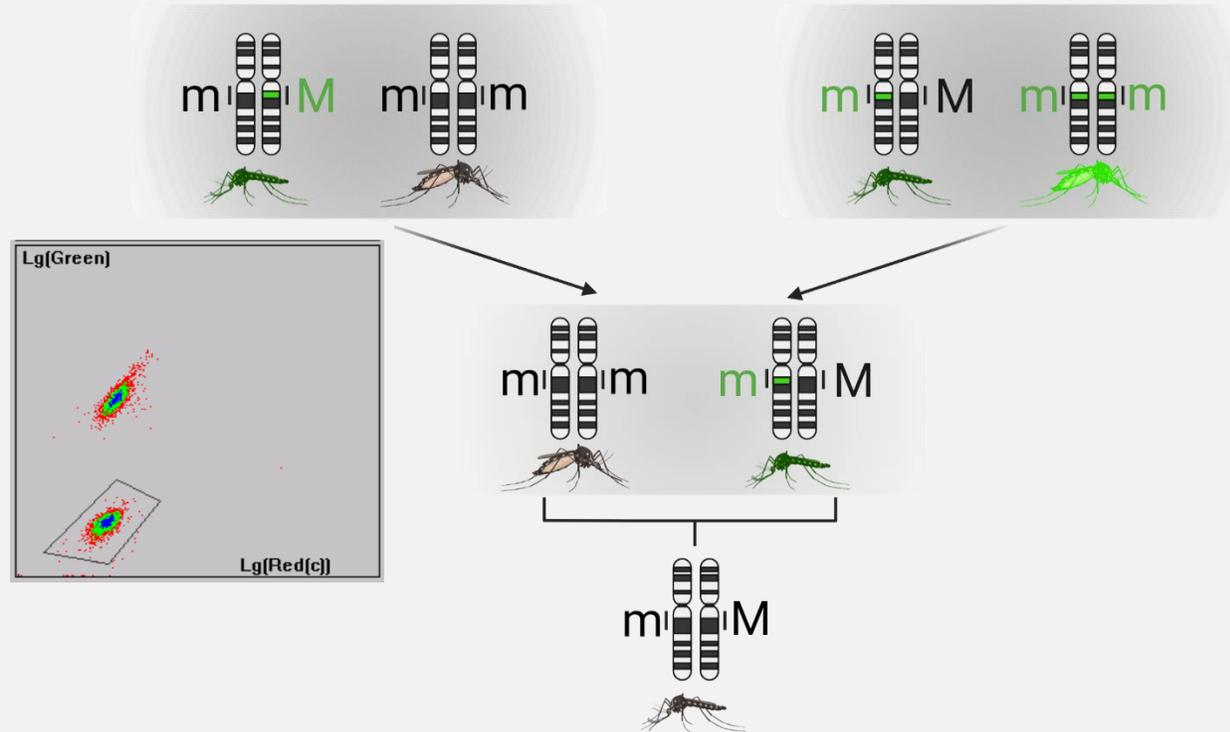
Obtaining non-transgenic males



RESULTS – *Aedes Aegypti*

Obtaining non-transgenic males

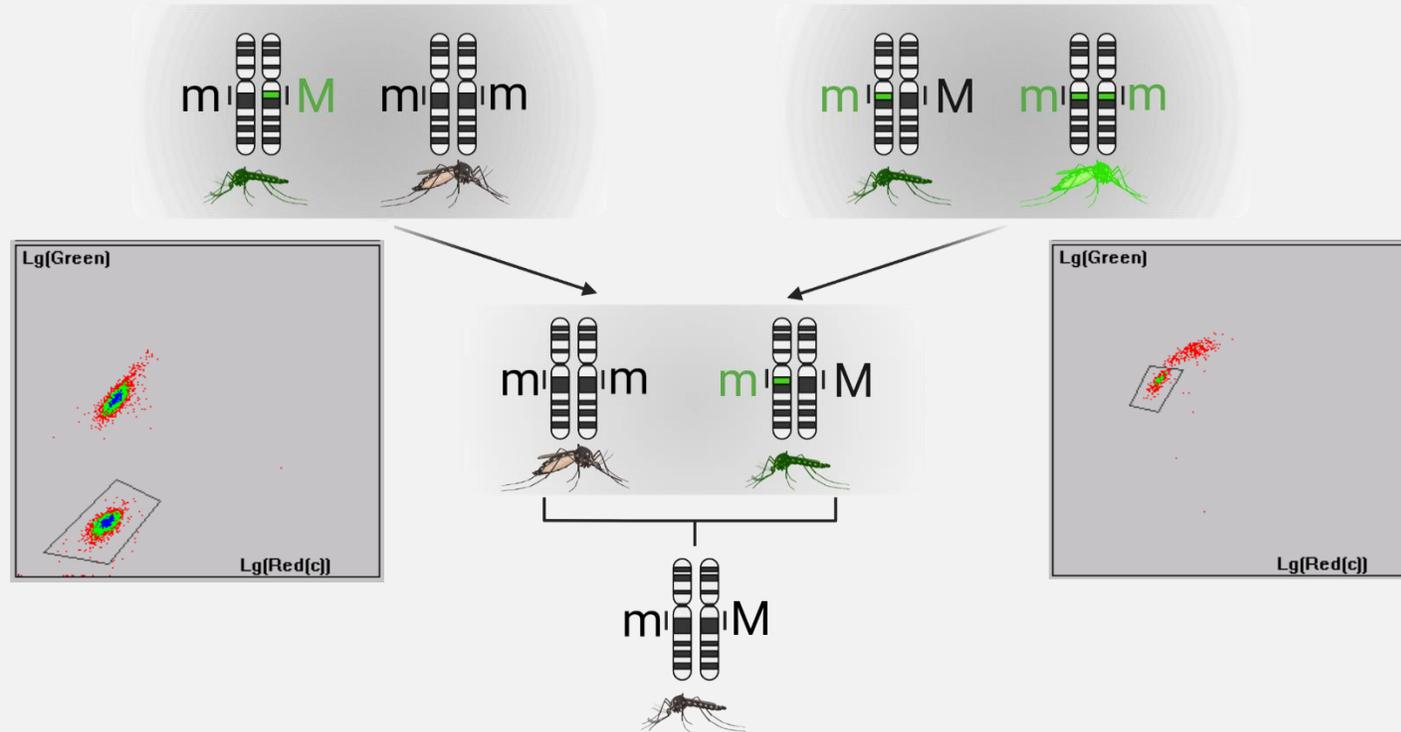
1,000 non-transgenic females from the M-linked GSS



RESULTS – *Aedes Aegypti*

Obtaining non-transgenic males

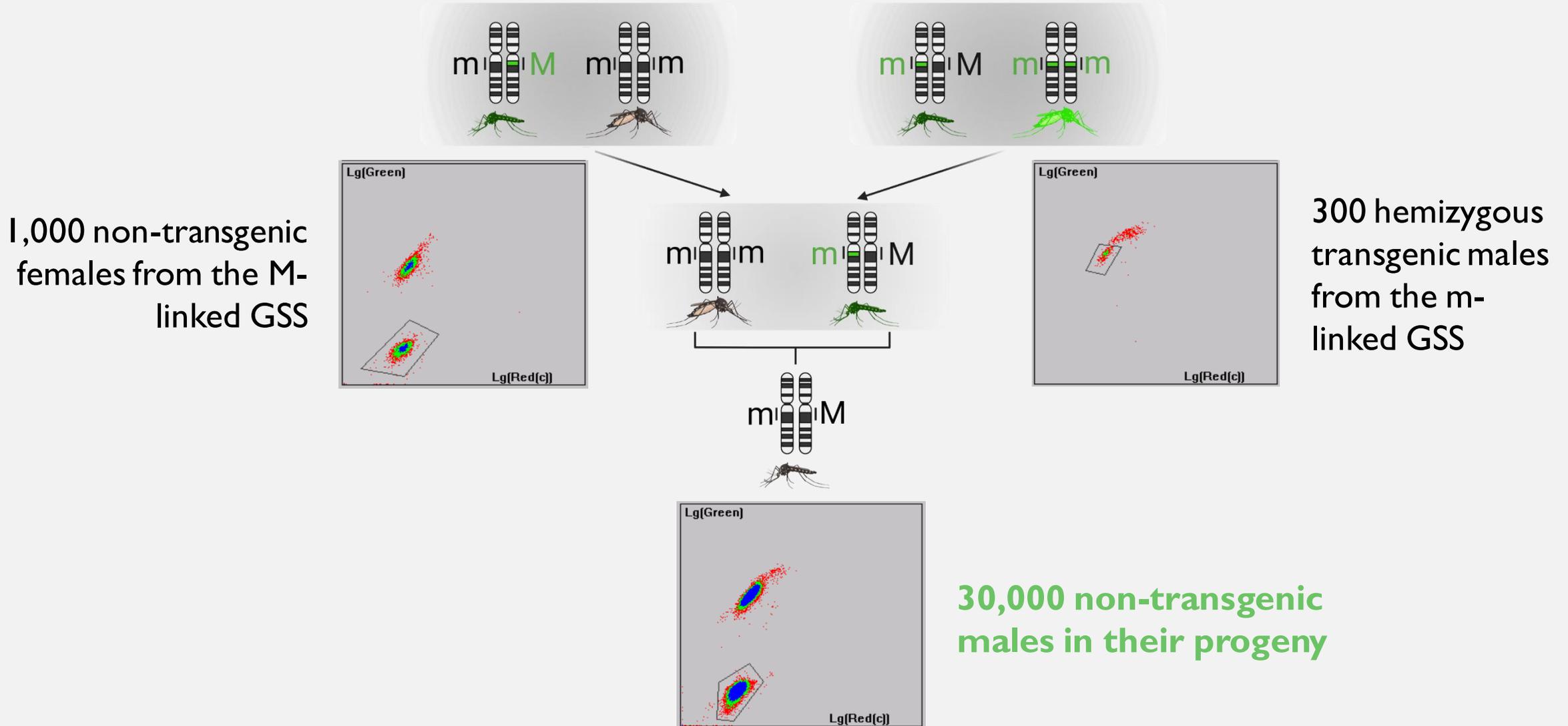
1,000 non-transgenic females from the M-linked GSS



300 hemizygous transgenic males from the m-linked GSS

RESULTS – *Aedes Aegypti*

Obtaining non-transgenic males



RESULTS

Developing GSS in *Aedes albopictus*



RESULTS – *Aedes albopictus*

In *Aedes albopictus*, there are no chromosome-long scaffolds.

Reference genome Aalbo_primary.1

None (2019). Isolate: FPA.

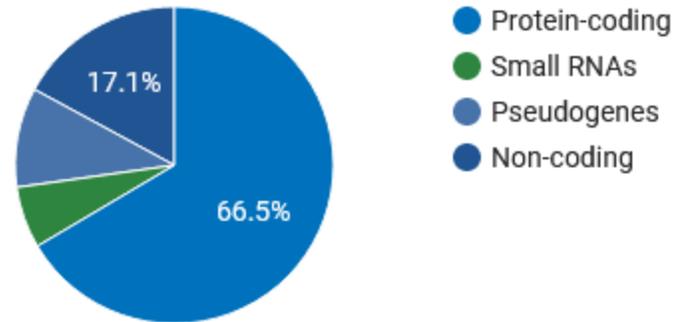
RefSeq GCF_006496715.1

Download

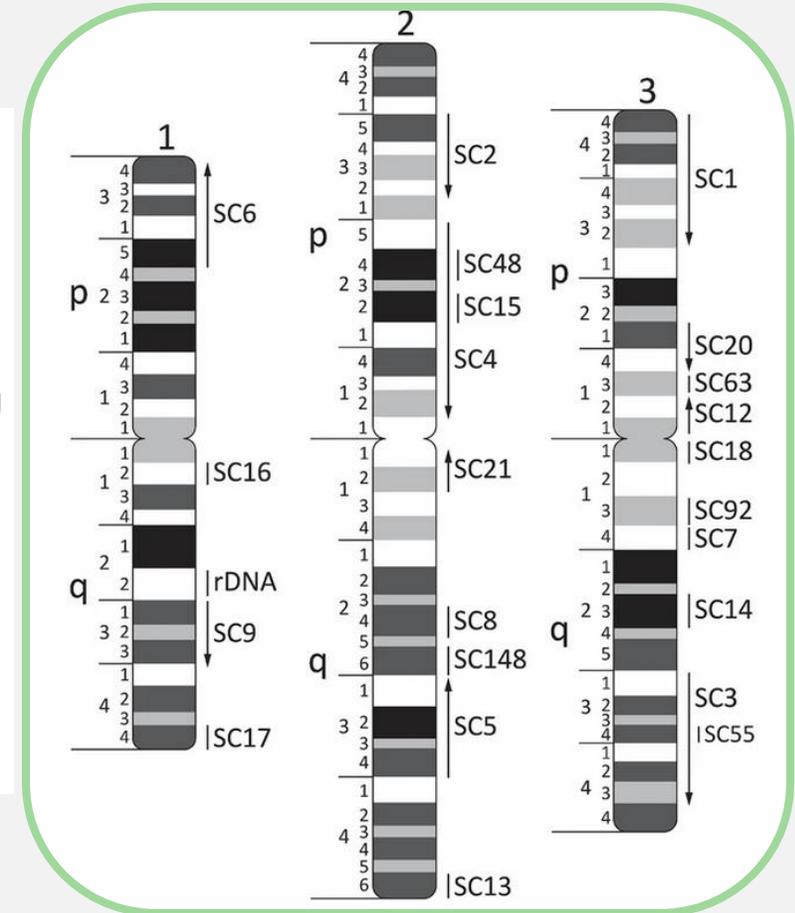
Genome size	2.5 Gb
Contig N50	1.2 Mb
Genes	40,494

NCBI Annotation Release 102 Jul 3, 2019

Current gene set

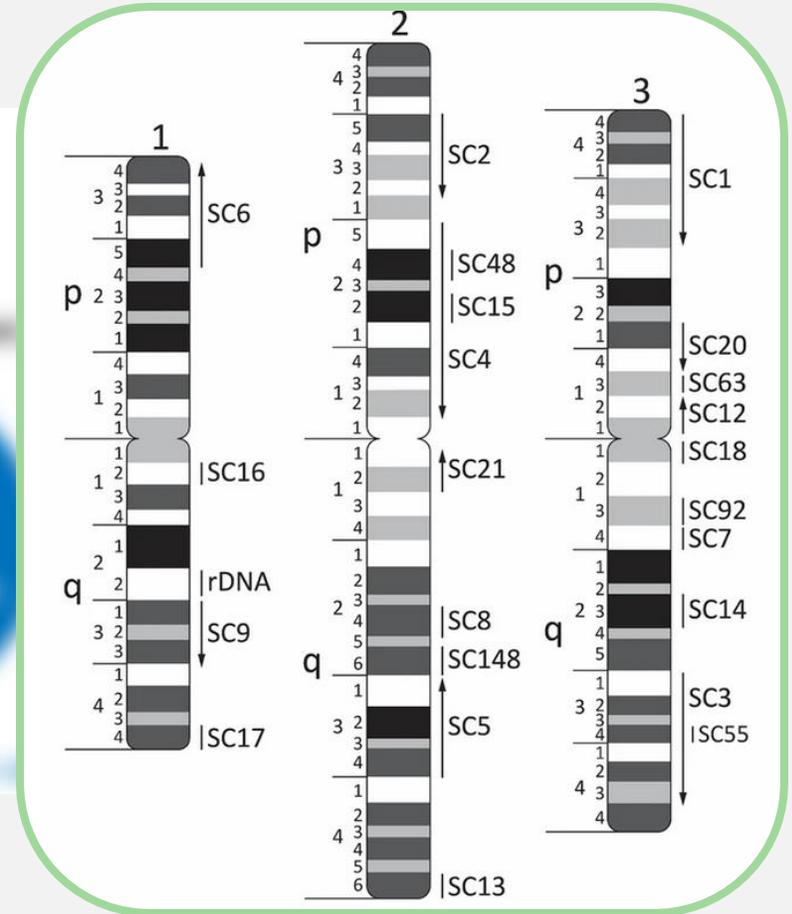
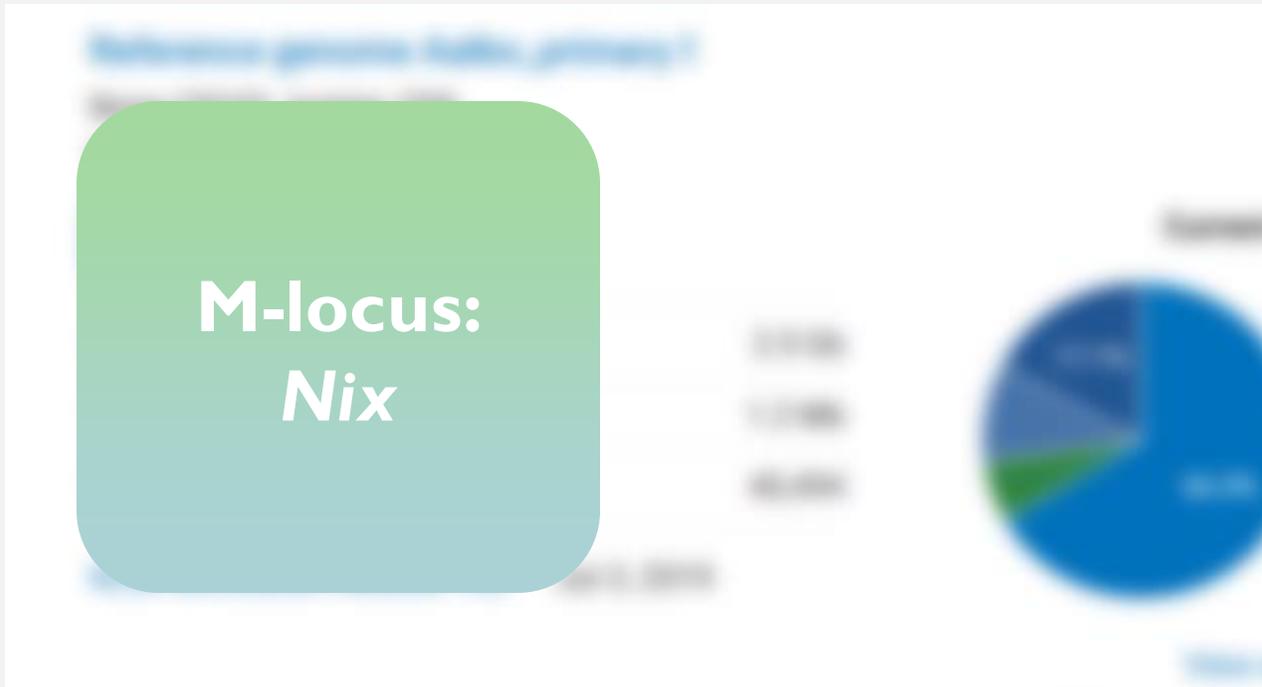


[View all genes](#)



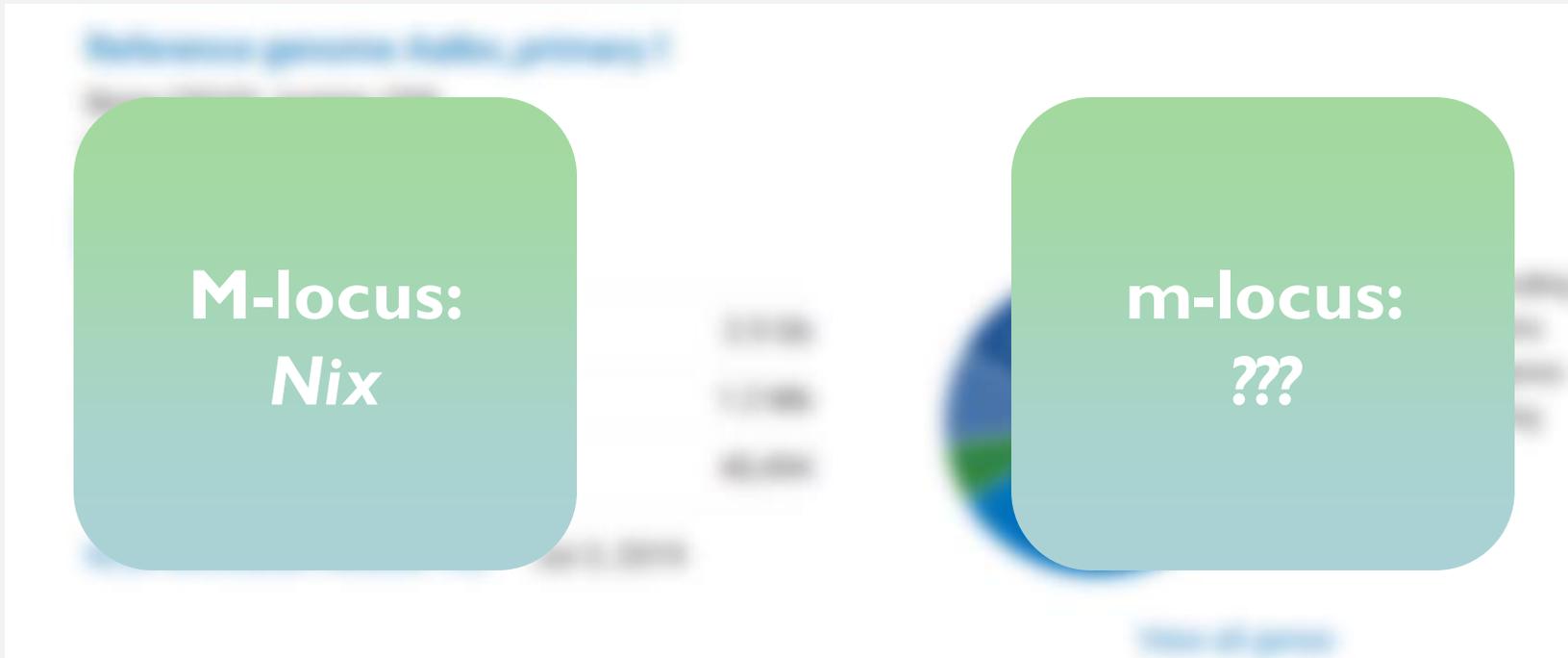
RESULTS – *Aedes albopictus*

In this genome, *Nix* belongs to scaffold 26I, which is <1 Mbp



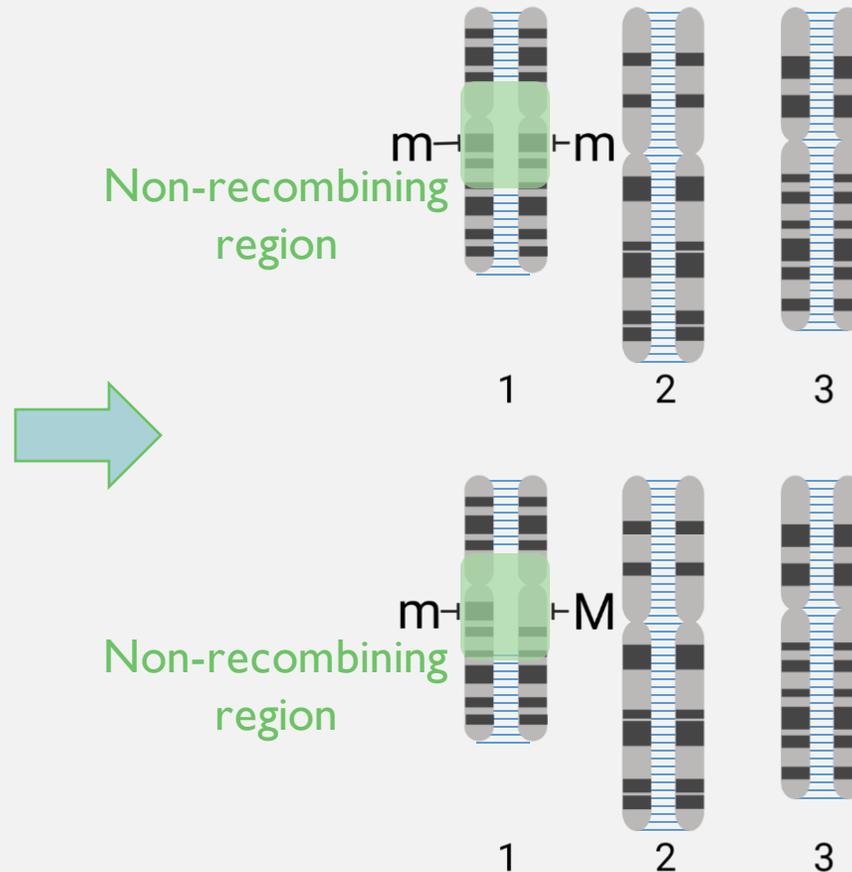
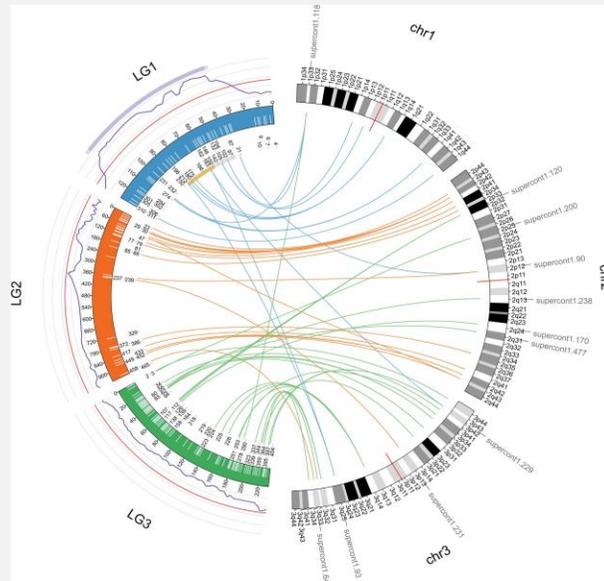
RESULTS – *Aedes albopictus*

There is no information about the m-locus



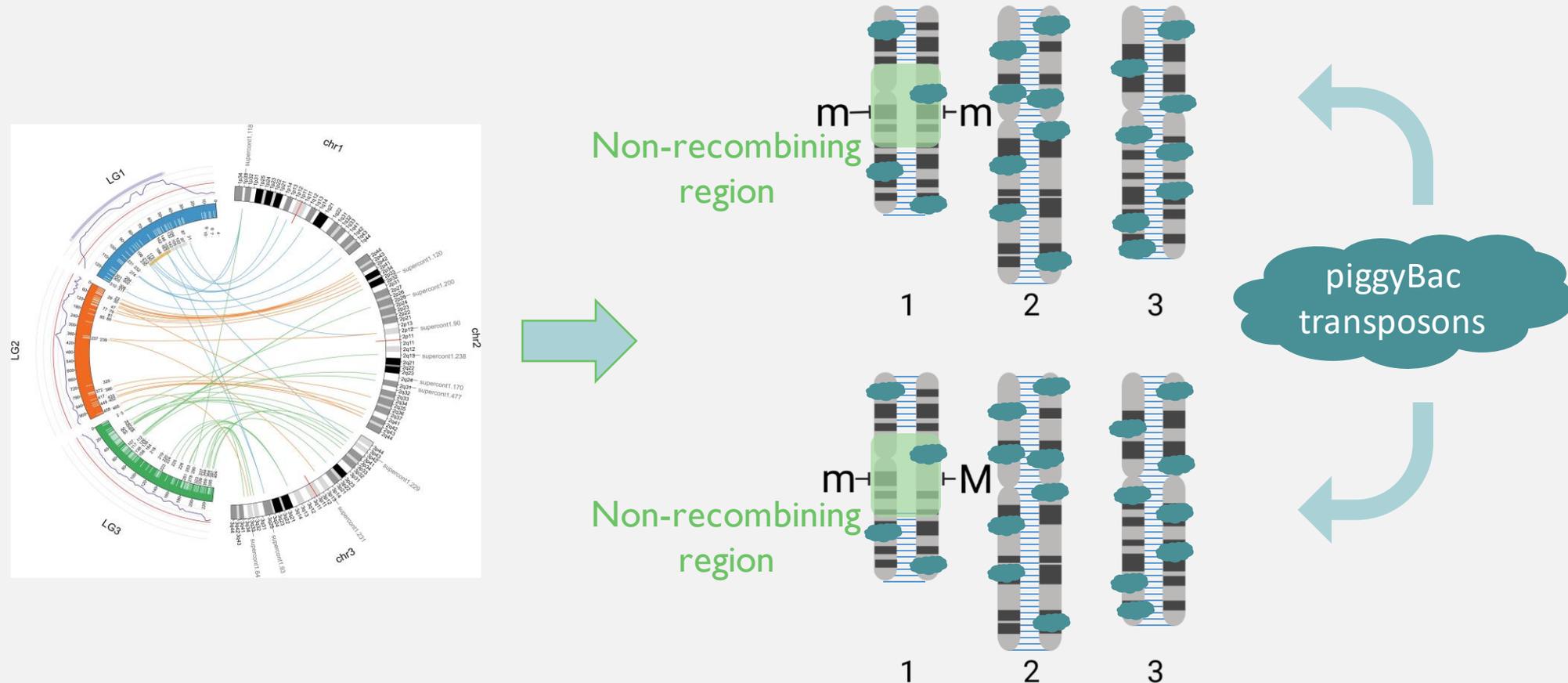
RESULTS – *Aedes albopictus*

If the non-recombining region is the same in *Ae. albopictus*, there is a ~5% chance to randomly land in a sex locus by non-targeted approaches



RESULTS – *Aedes albopictus*

If the non-recombining region is the same in *Ae. albopictus*, there is a ~5% chance to randomly land in a sex locus by non-targeted approaches



RESULTS – *Aedes albopictus*

piggyBac
approach



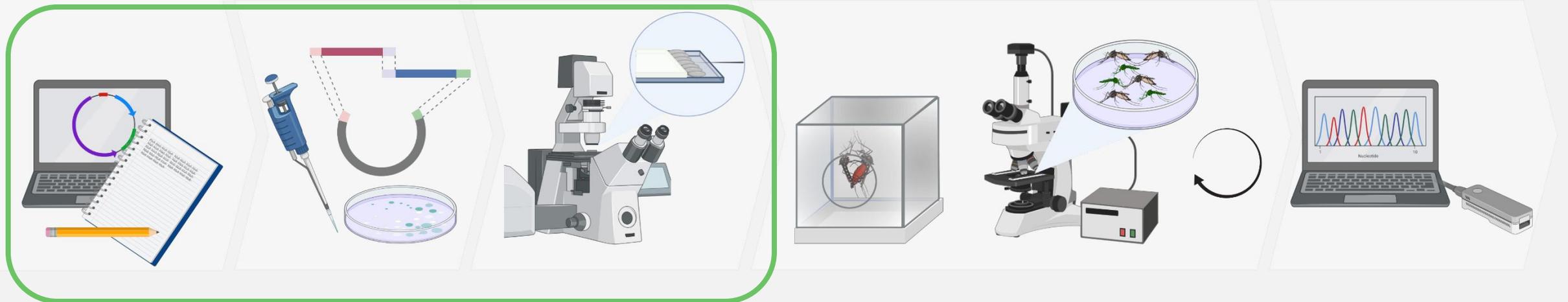
Design

Cloning

Microinjection

Crossing, Screening, Repeat

Sequencing / PCR



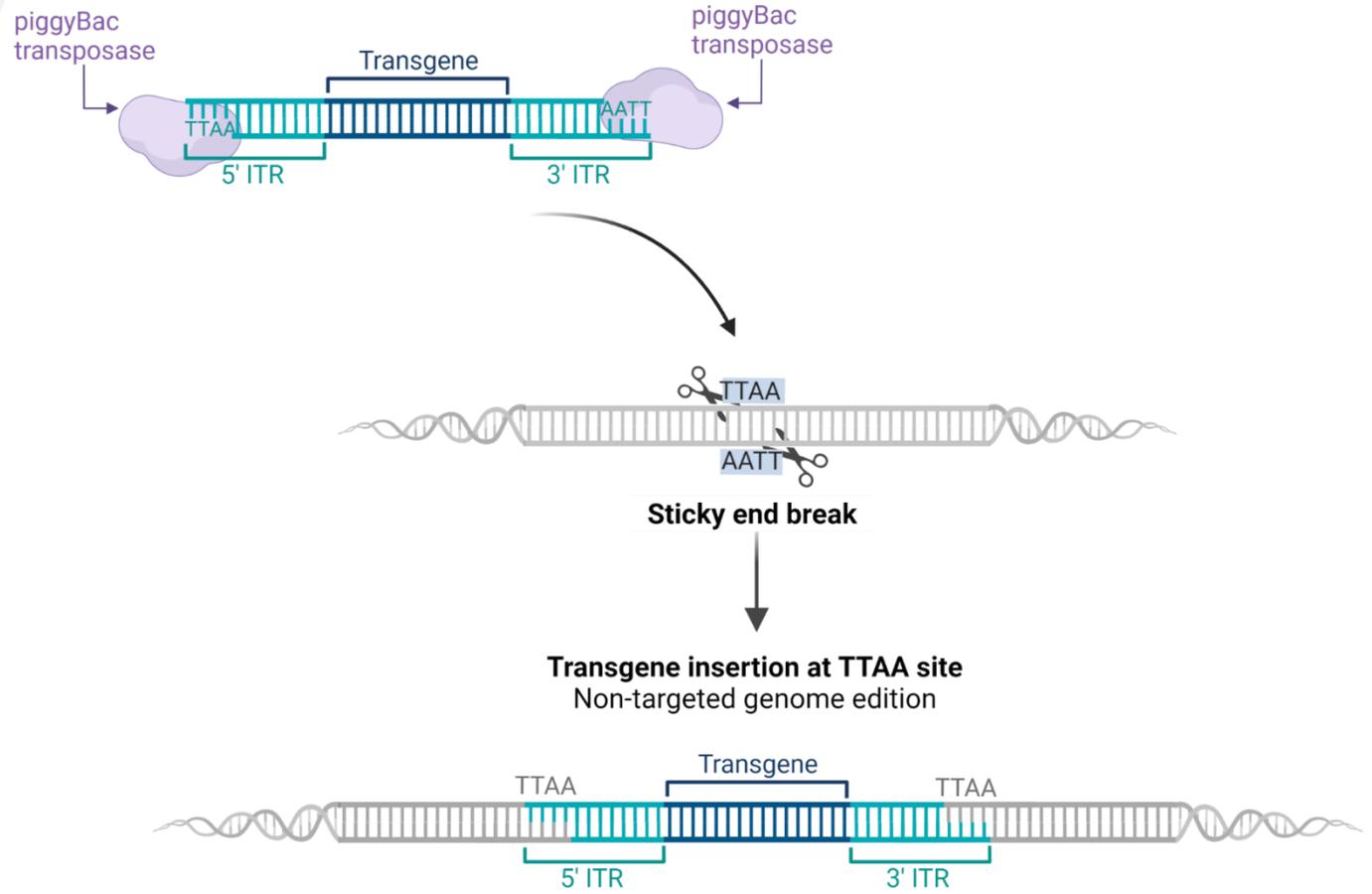
RESULTS – *Aedes albopictus*

piggyBac approach

Design

Cloning

Microi



RESULTS – *Aedes albopictus*

piggyBac approach



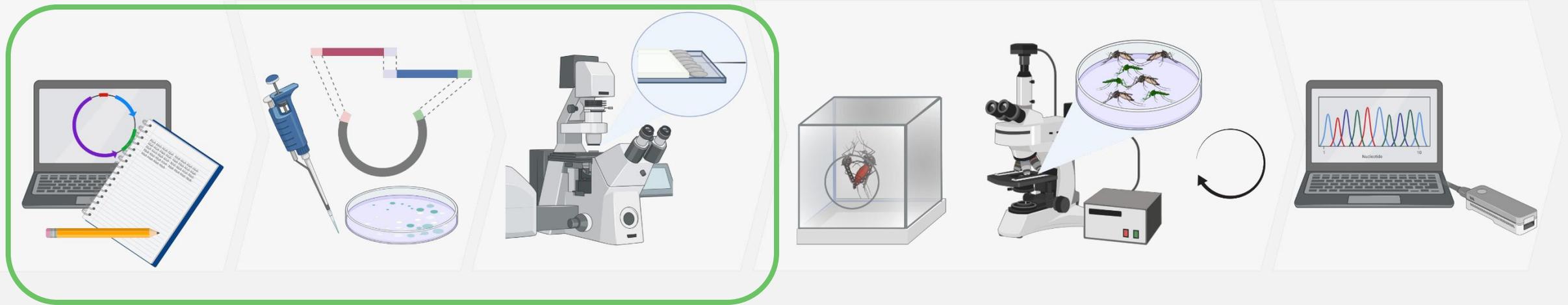
Design

Cloning

Microinjection

Crossing, Screening, Repeat

Sequencing / PCR



6 piggyBac plasmids
encoding different fluorochromes

RESULTS – *Aedes albopictus*

piggyBac approach



Design

Injection of 100s of WT eggs

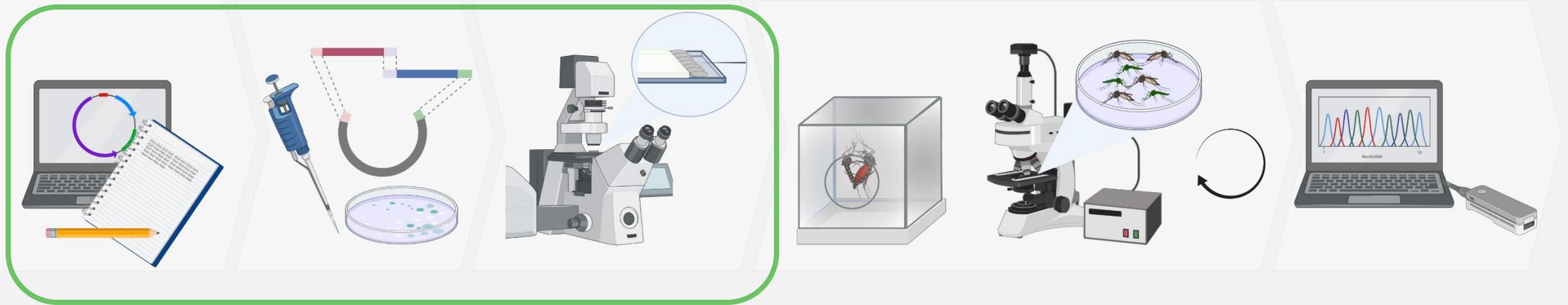


Microinjection

Cloning

Crossing, Screening, Repeat

Sequencing / PCR



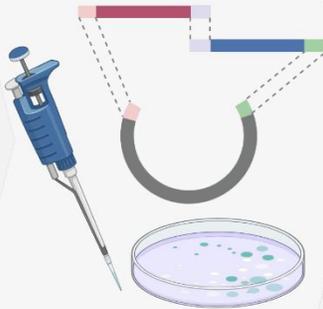
6 piggyBac plasmids encoding different fluorochromes

RESULTS – *Aedes albopictus*

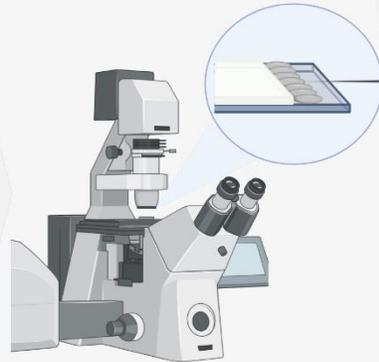
Design



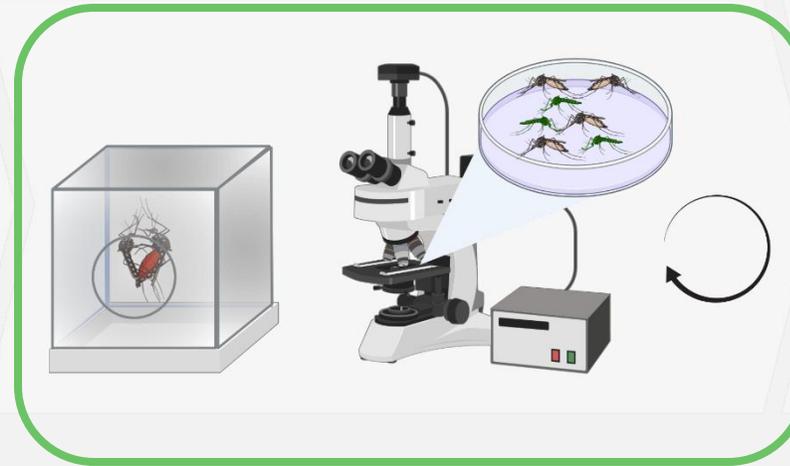
Cloning



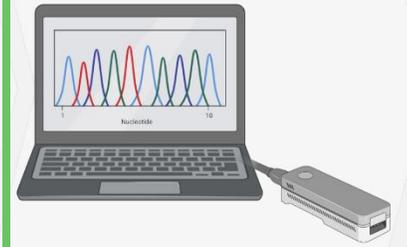
Microinjection



Crossing, Screening, Repeat



Sequencing / PCR



RESULTS – *Aedes albopictus*

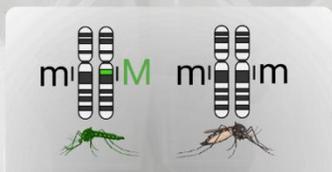
> 120 individual insertions



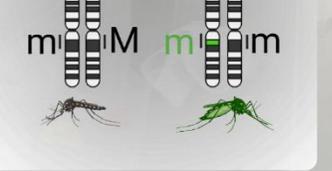
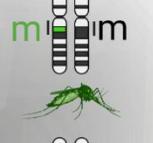
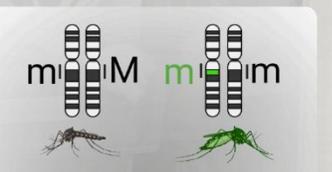
RESULTS – *Aedes albopictus*

> 120 individual insertions

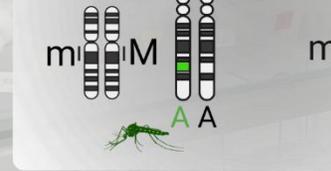
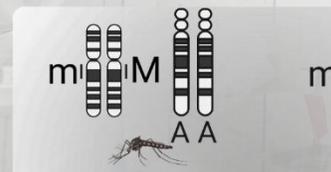
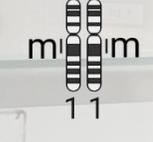
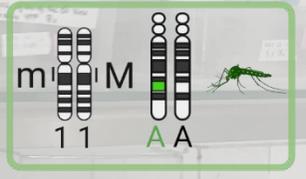
M-linked insertion



m-linked insertion



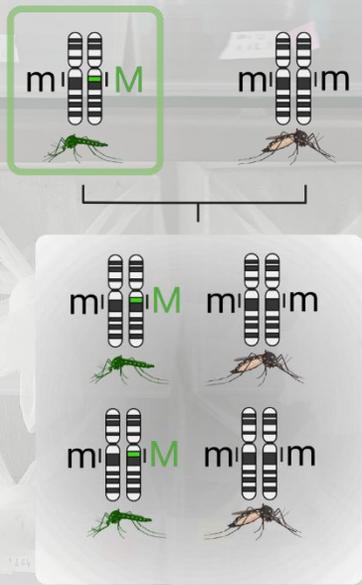
Autosomal insertion



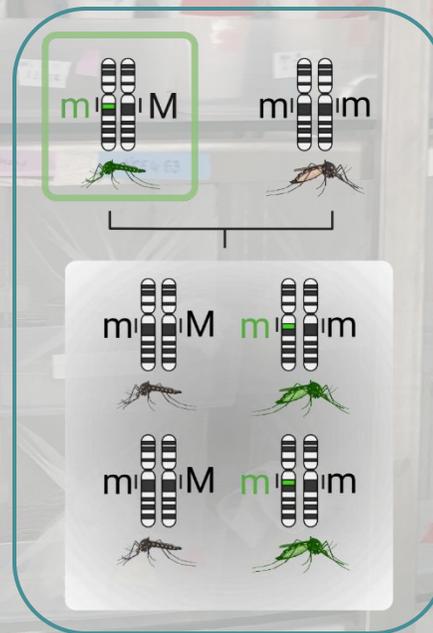
RESULTS – *Aedes albopictus*

> 120 individual insertions + older lines

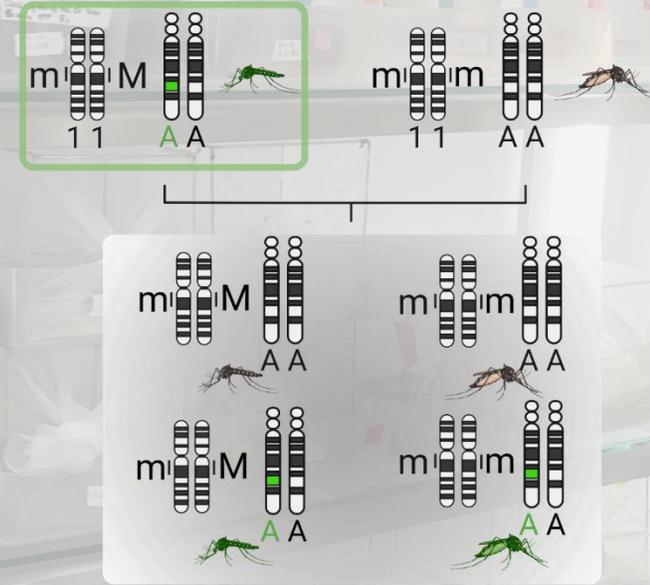
M-linked insertion



m-linked insertion



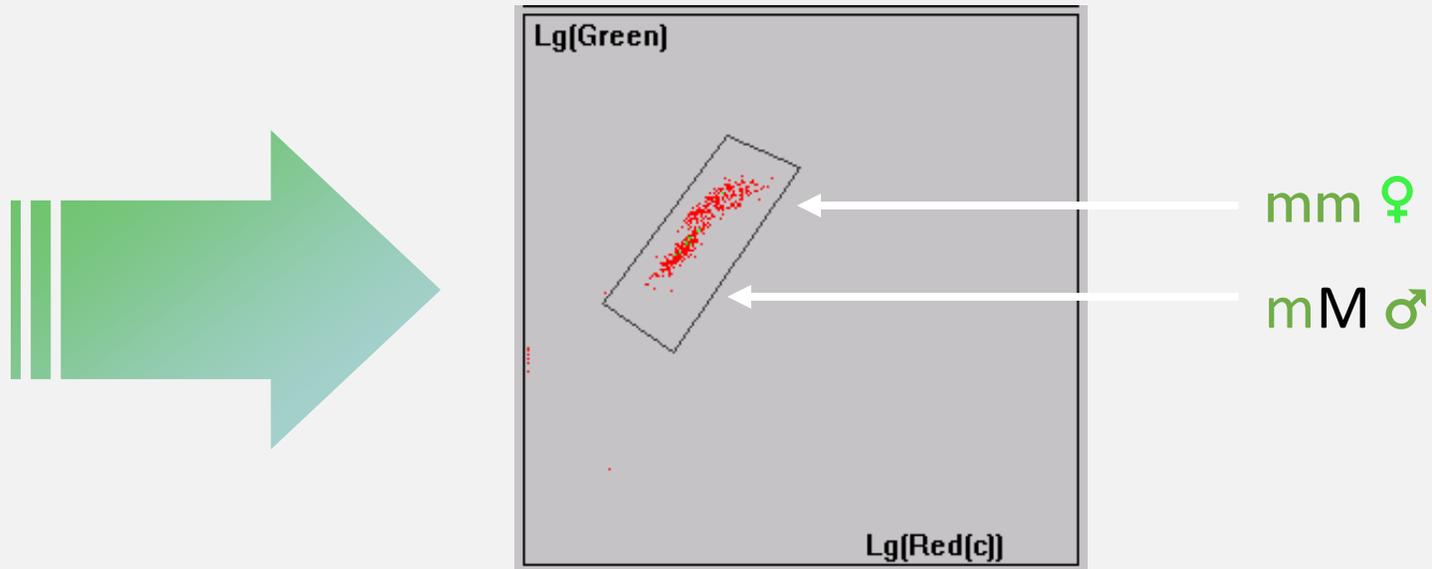
Autosomal insertion



1 m-linkage with 0.1% recombination

RESULTS – *Aedes albopictus*

This m-linked line allows sex separation of neonate larvae



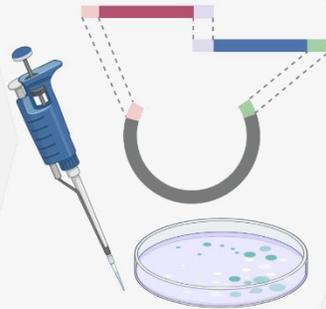
RESULTS – *Aedes albopictus*

Targeted sequencing recovered the sequence of the insertion locus

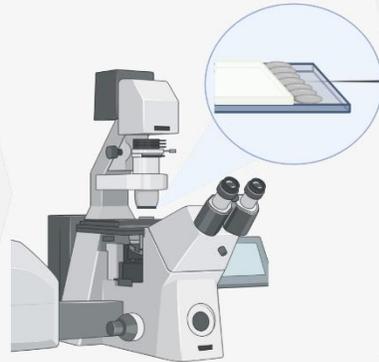
Design



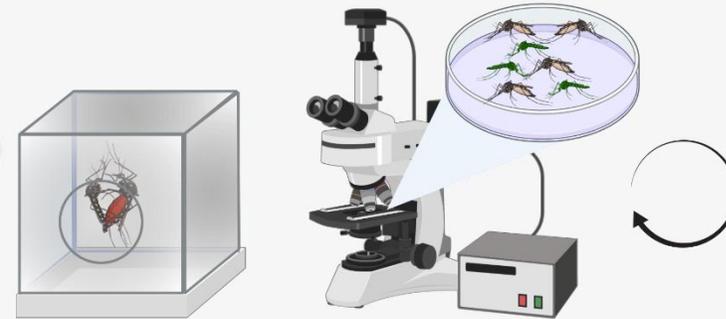
Cloning



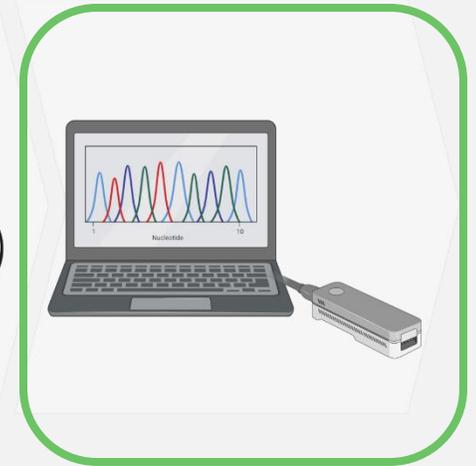
Microinjection



Crossing, Screening, Repeat



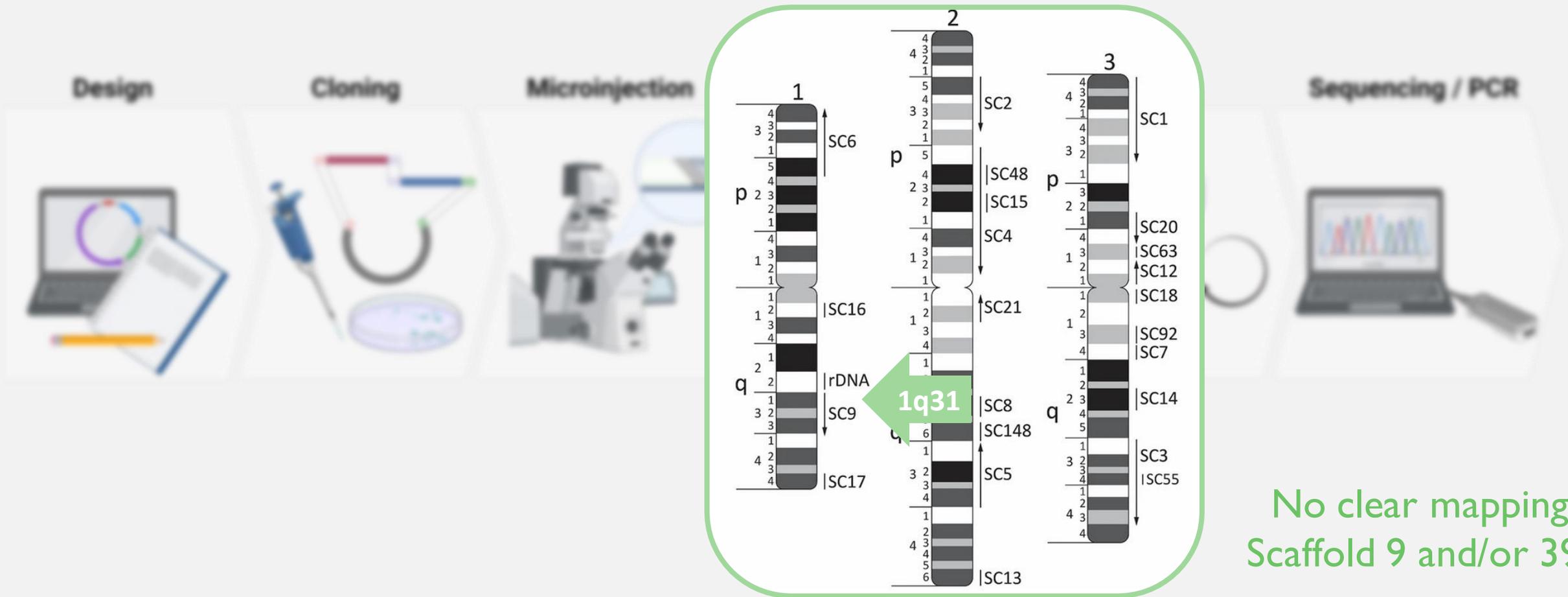
Sequencing / PCR



↑
No clear mapping
Scaffold 9 and/or 39?

RESULTS – *Aedes albopictus*

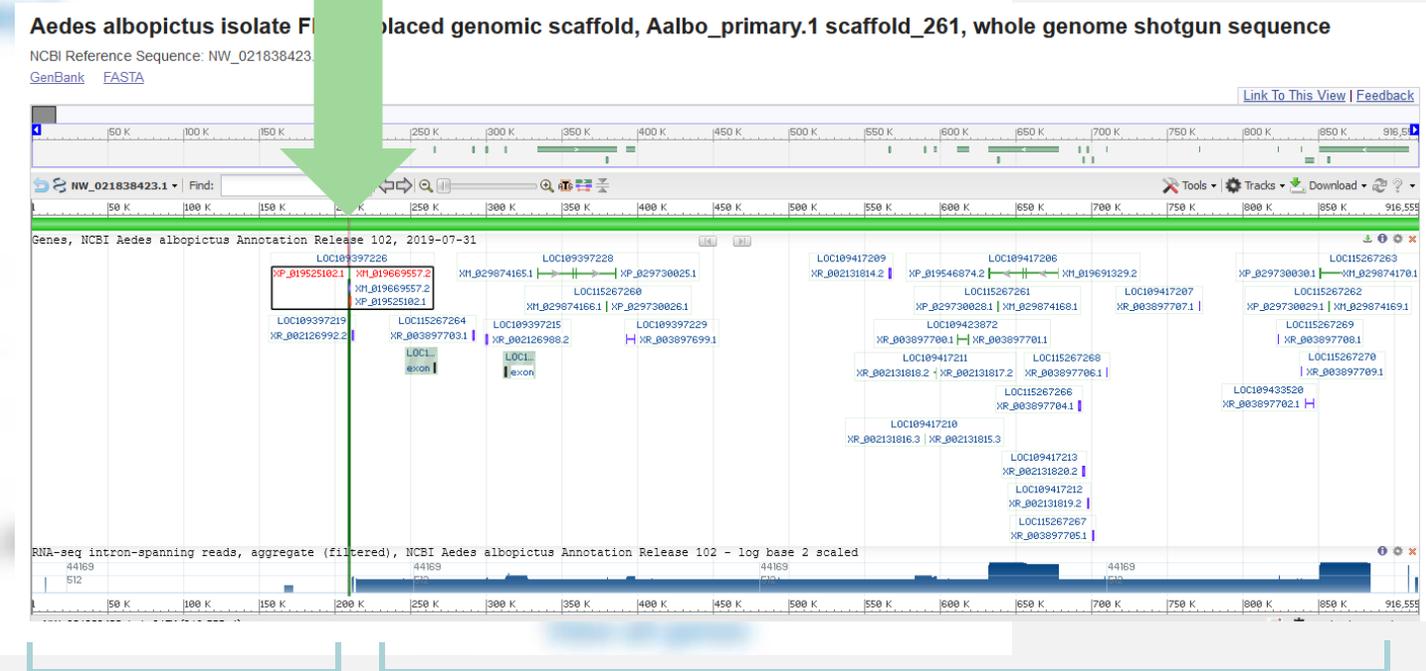
Targeted sequencing recovered the sequence of the insertion locus



RESULTS – *Aedes albopictus*

Finding targets for the M-locus

M-locus:
Nix is the only
gene we know

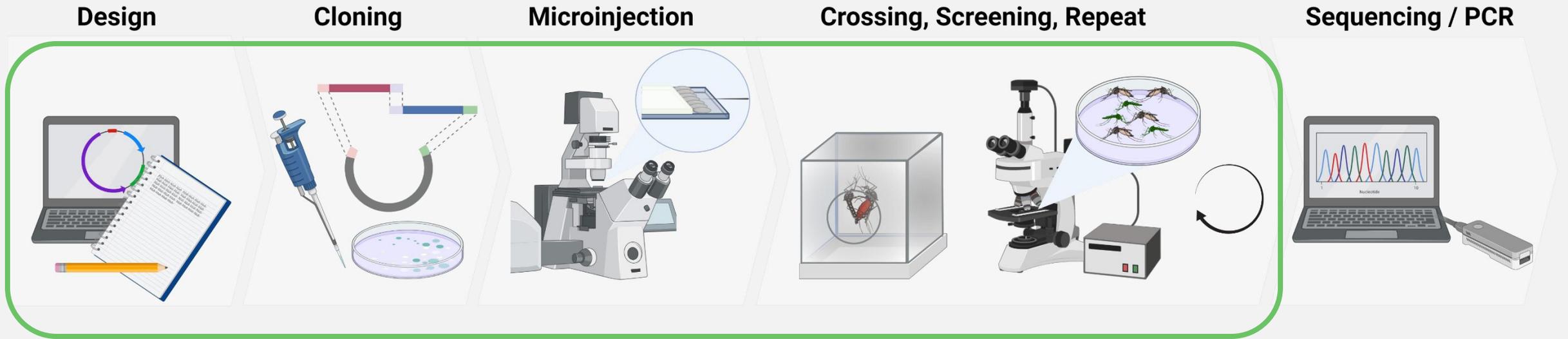


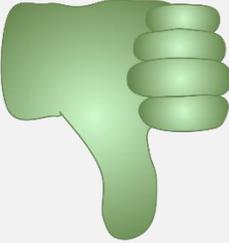
?

No male-specific gene

RESULTS – *Aedes albopictus*

Targeting *Nix* by CRISPR-Cas9 failed



No transgenic 

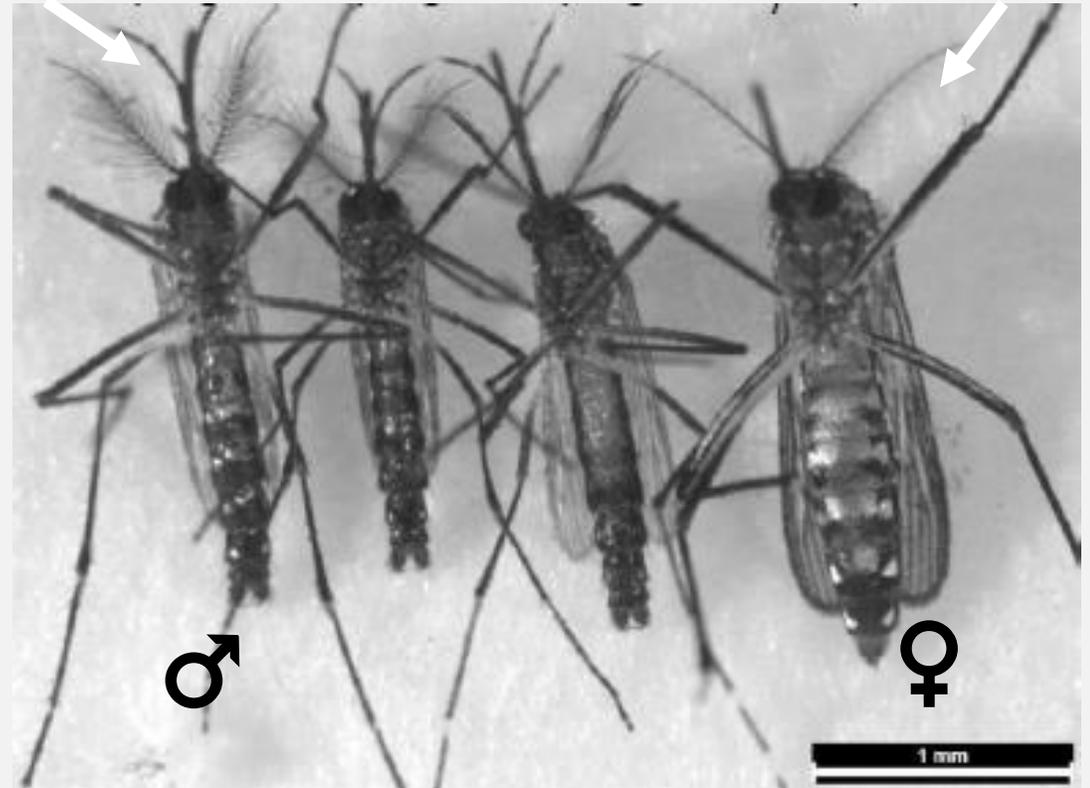
RESULTS – *Aedes albopictus*

If you can't reach the M-locus: create your M-locus!



Genetic
male

Genetic
female

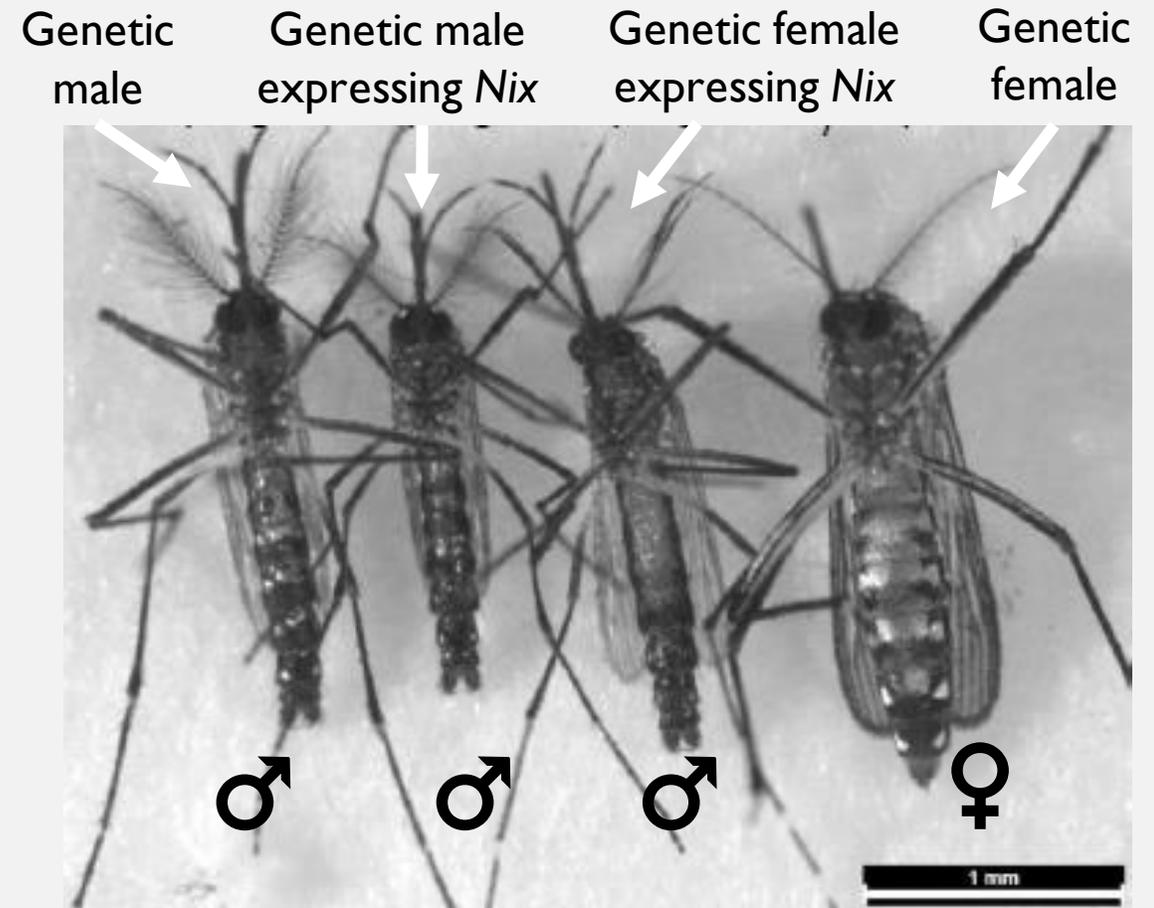


Aryan et al. 2020

RESULTS – *Aedes albopictus*

If you can't reach the M-locus: create your M-locus!

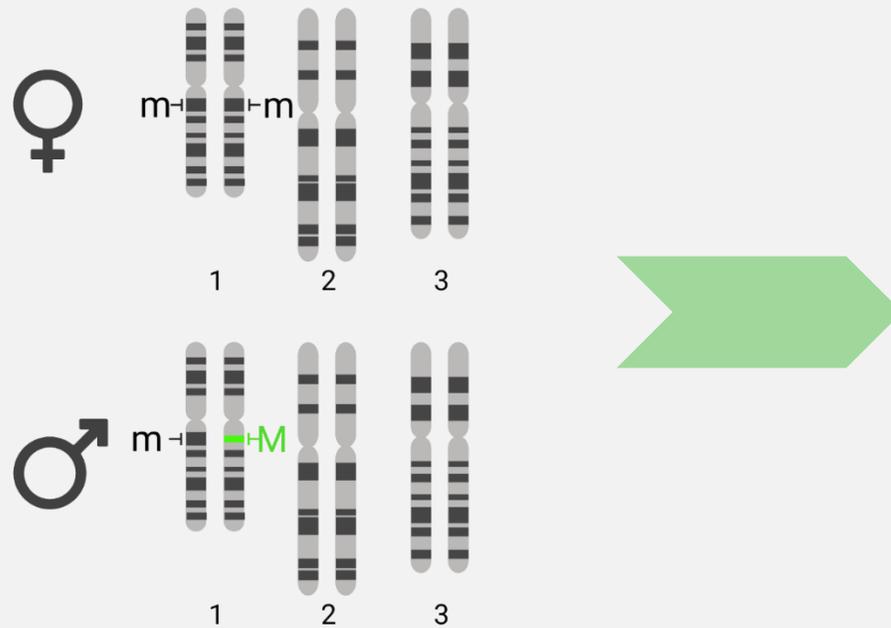
Aryan *et al.* (2020) showed in *Ae. aegypti* that *Nix* alone could turn genetic females into fertile phenotypic males



Aryan *et al.* 2020

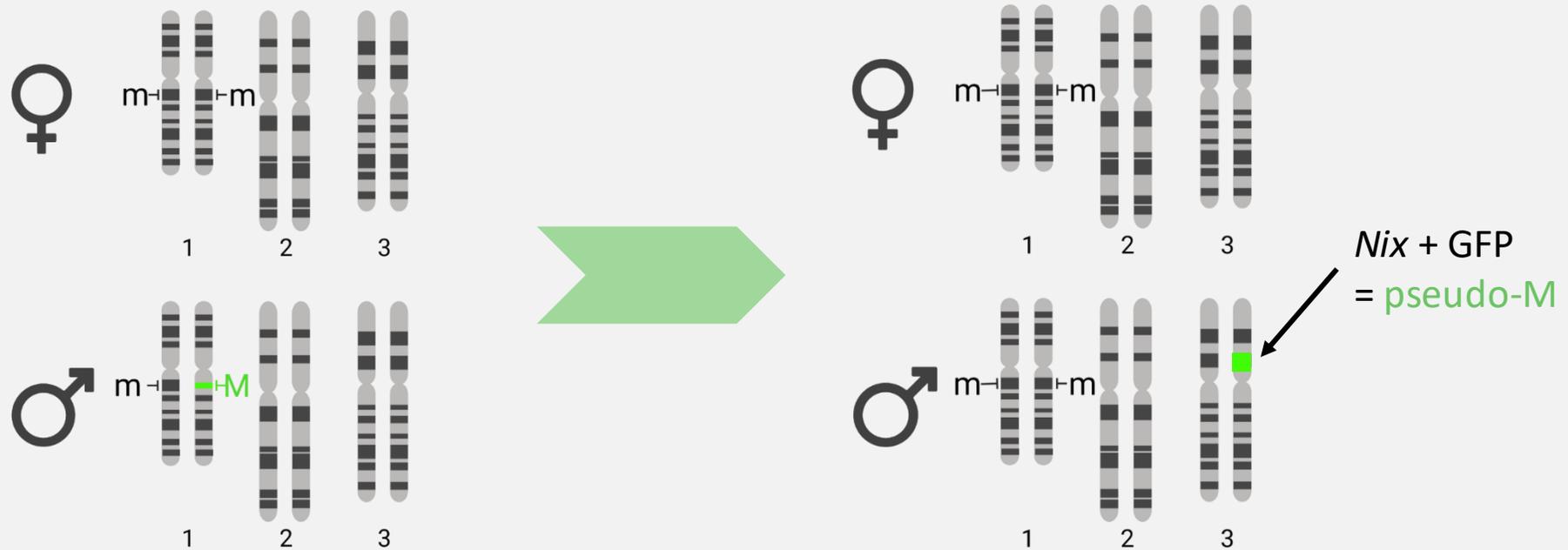
RESULTS – *Aedes albopictus*

If you can't reach the M-locus: create your M-locus!



RESULTS – *Aedes albopictus*

If you can't reach the M-locus: create your M-locus!



RESULTS – *Aedes albopictus*

piggyBac approach

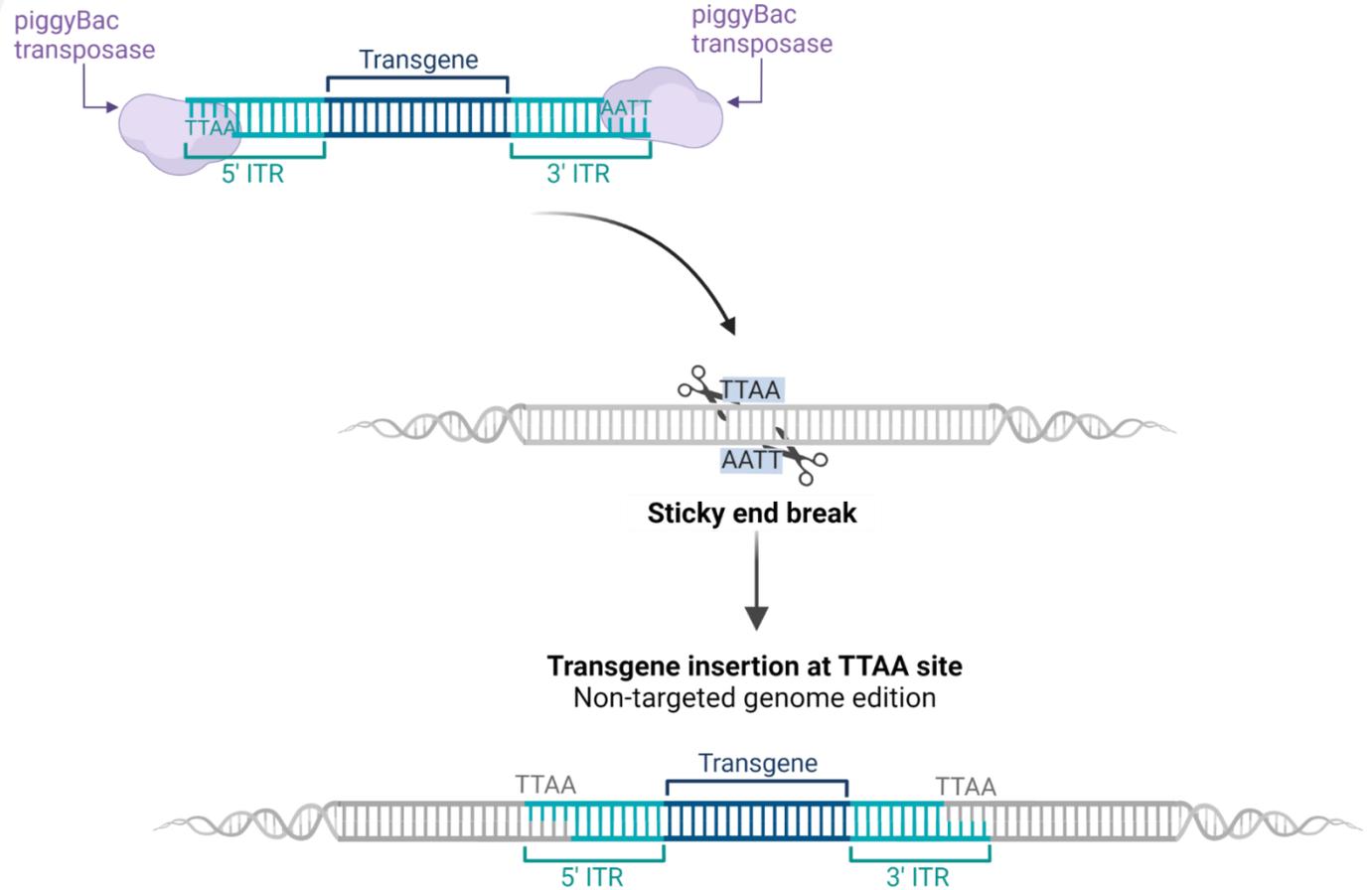
Design



Cloning



Microi



RESULTS – *Aedes albopictus*

piggyBac
approach



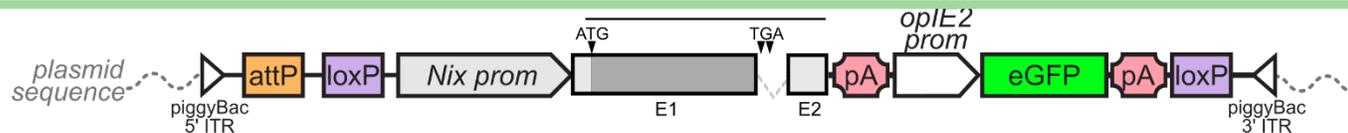
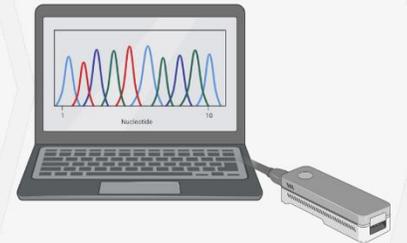
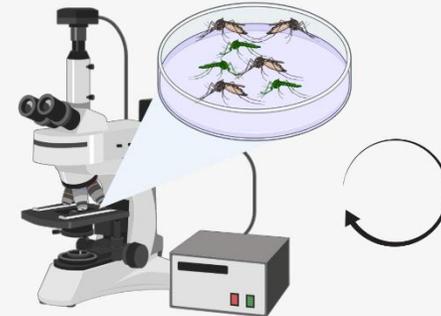
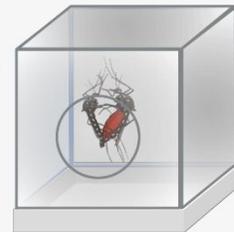
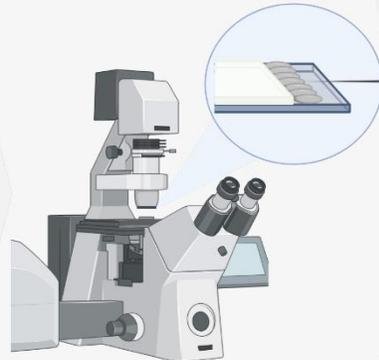
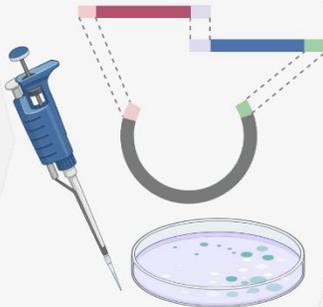
Design

Cloning

Microinjection

Crossing, Screening, Repeat

Sequencing / PCR



RESULTS – *Aedes albopictus*

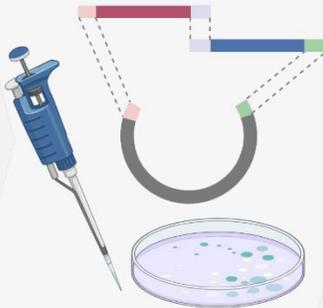
22 transgenic males
9 transgenic females



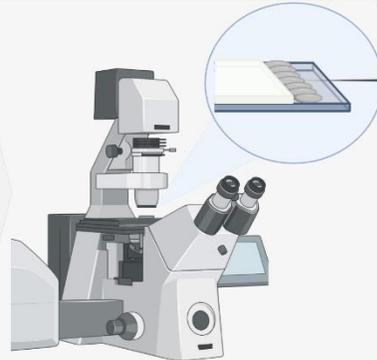
Design



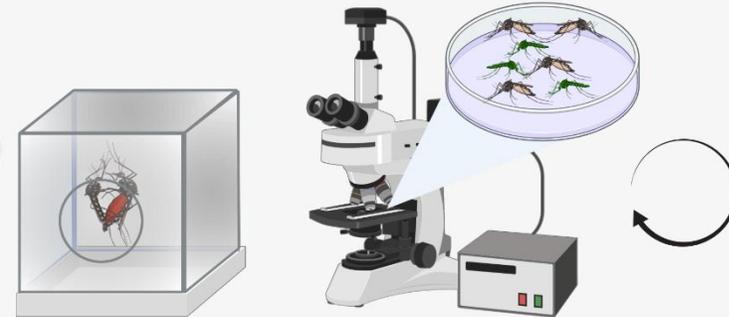
Cloning



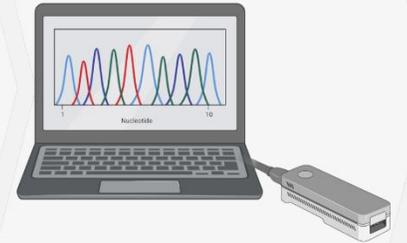
Microinjection



Crossing, Screening, Repeat

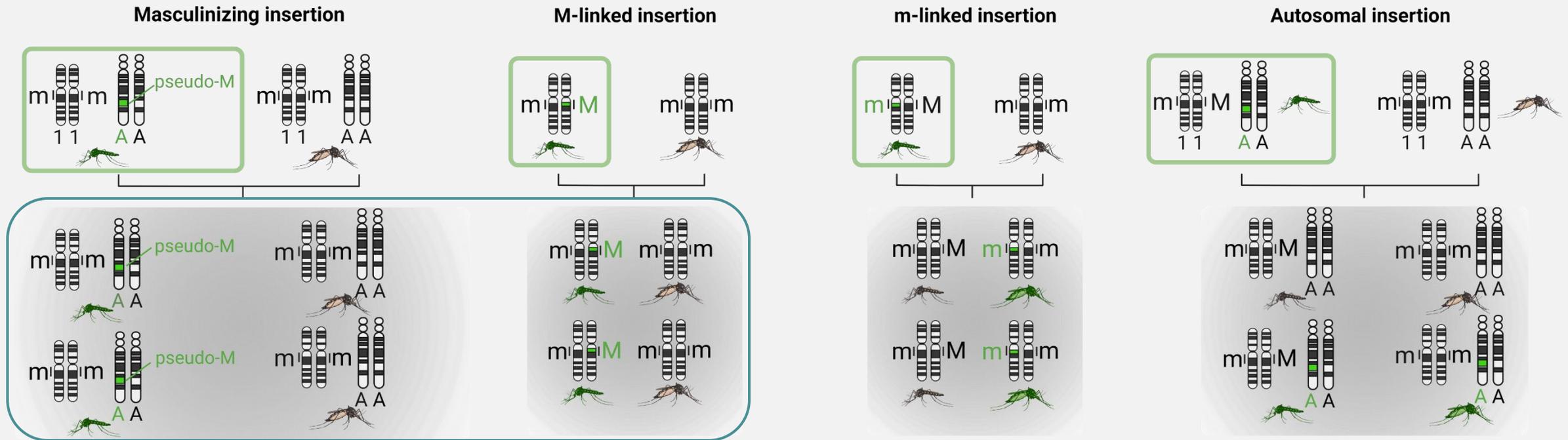


Sequencing / PCR



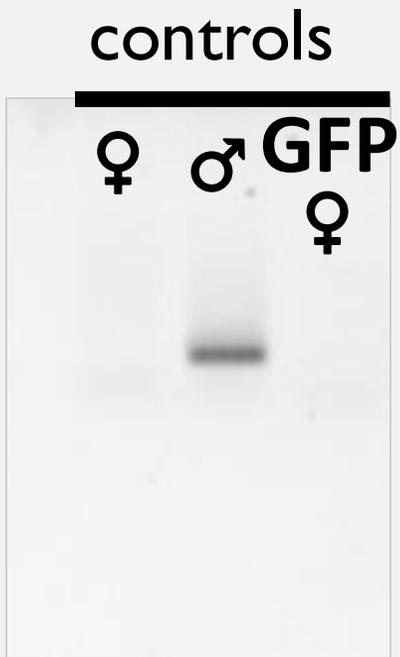
RESULTS – *Aedes albopictus*

12 individual males



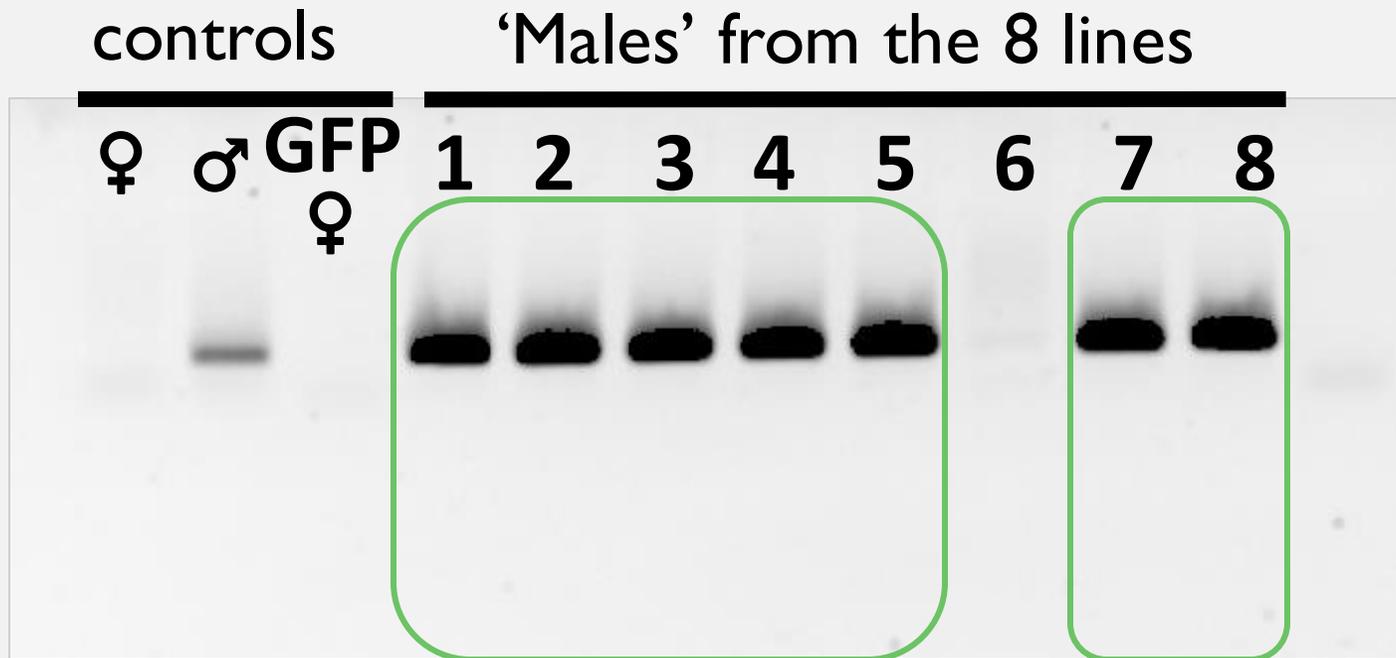
8 lines with transgenic sons and non-transgenic daughters

RESULTS – *Aedes albopictus*



Amplification of the
endogenous *Nix* gene

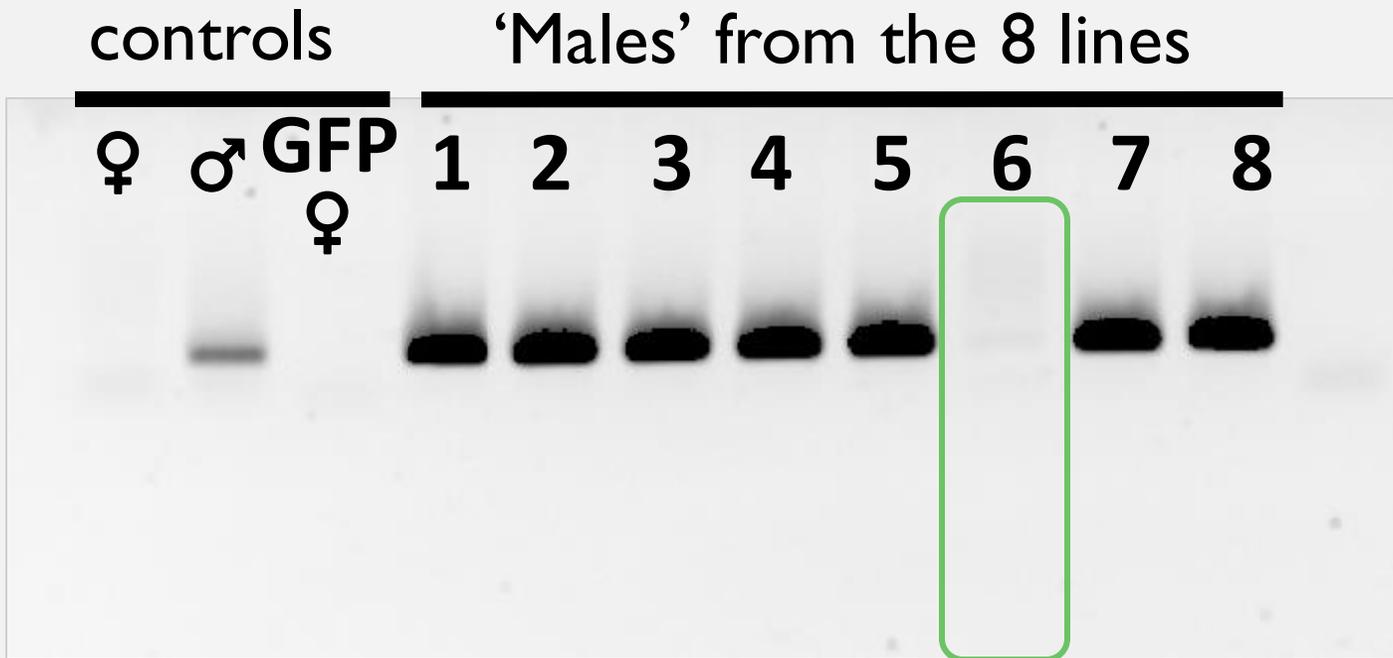
RESULTS – *Aedes albopictus*



Amplification of the endogenous *Nix* gene

7 out of 8 carry it:
M linkage

RESULTS – *Aedes albopictus*



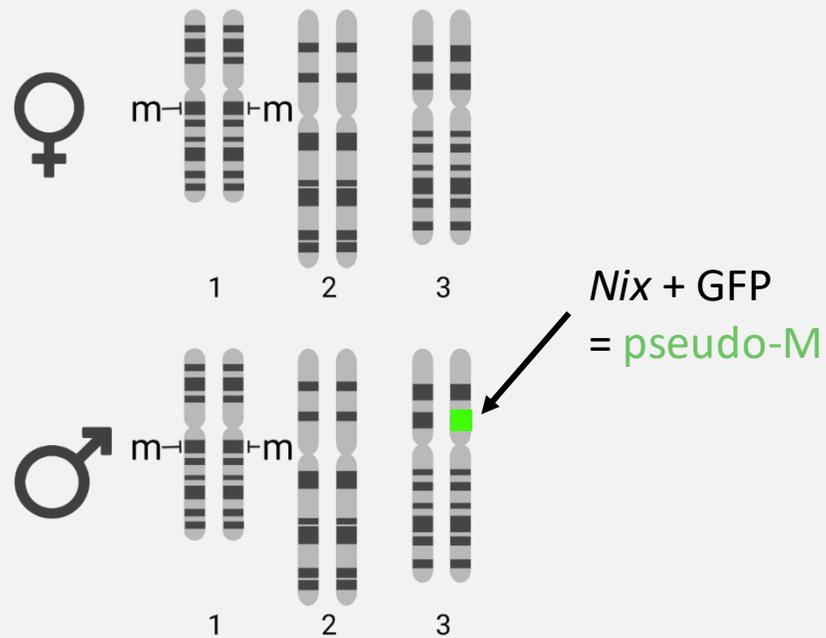
Amplification of the endogenous *Nix* gene

7 out of 8 carry it:
M linkage

1 does not:
pseudo-M linkage

RESULTS – *Aedes albopictus*

Hypotheses



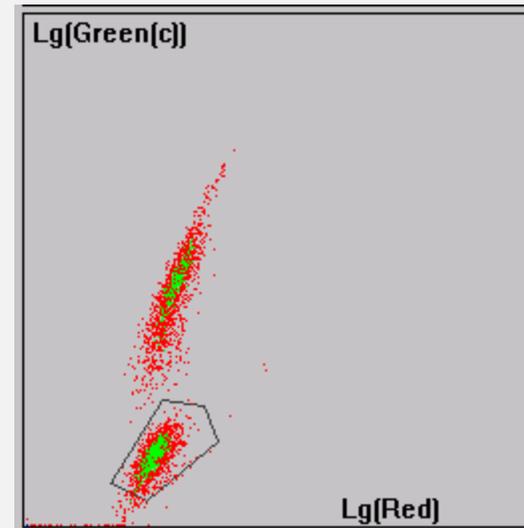
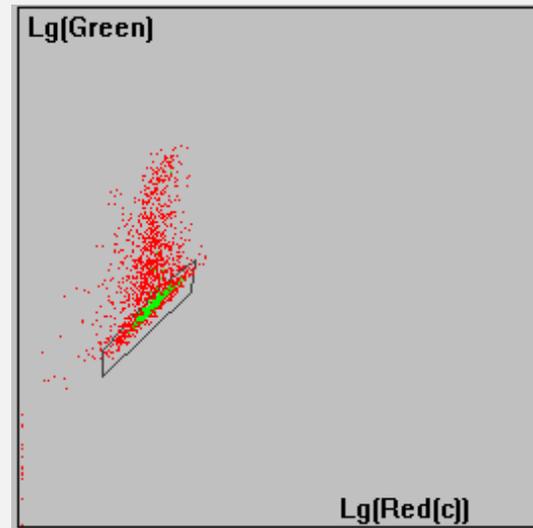
Amplification of the
endogenous *Nix* gene

7 out of 8 carry it:
M linkage

I does not:
pseudo-M linkage

RESULTS – *Aedes albopictus*

Issue: GFP is weak and unevenly expressed under the OpIE2 promoter



RESULTS – *Aedes albopictus*

Solution: replacing OpIE2 with PUb promoter

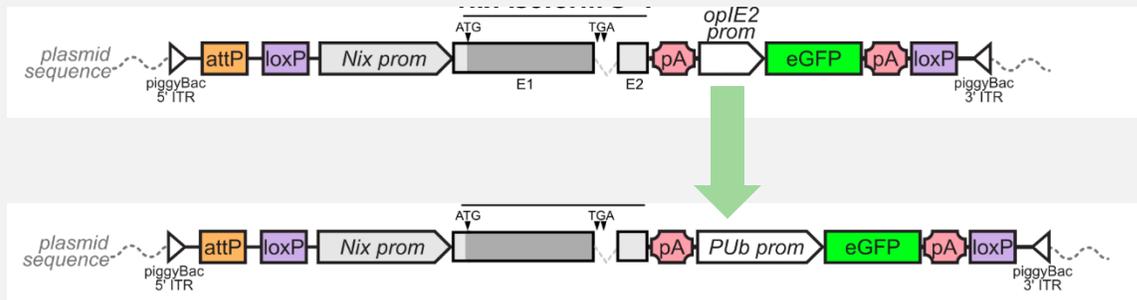
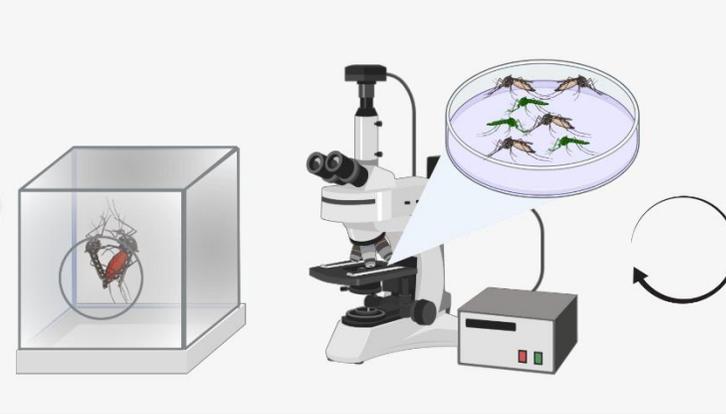
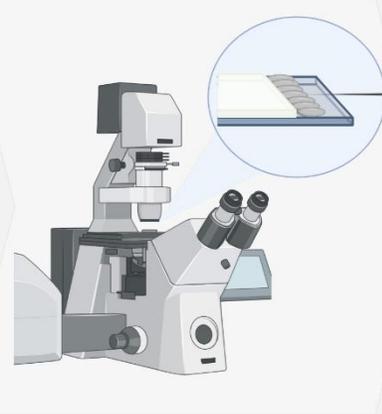
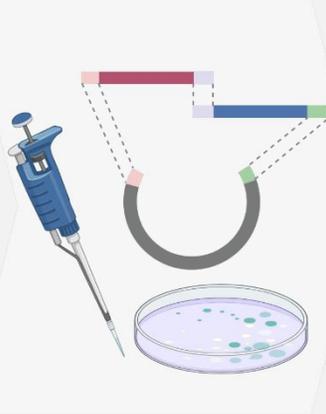
Design

Cloning

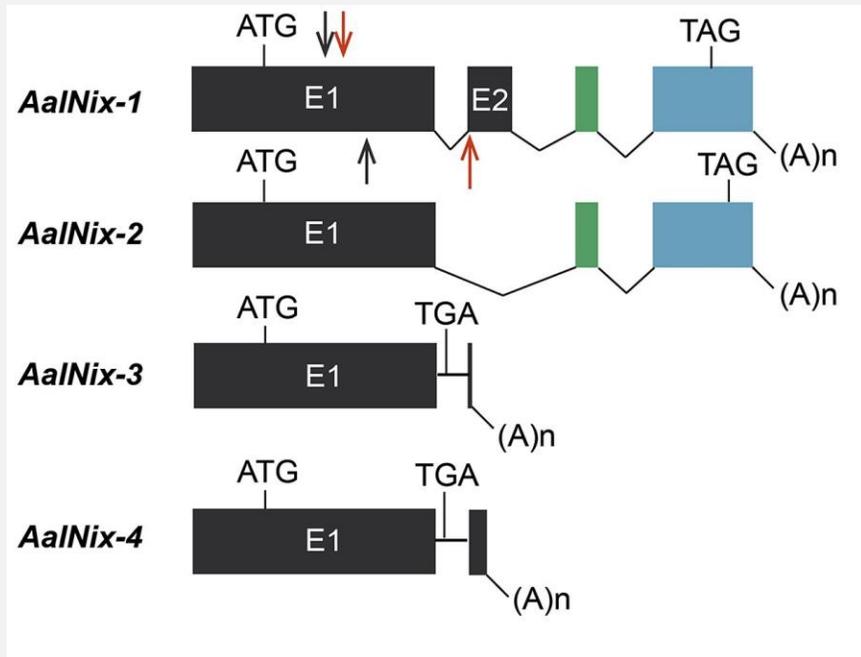
Microinjection

Crossing, Screening, Repeat

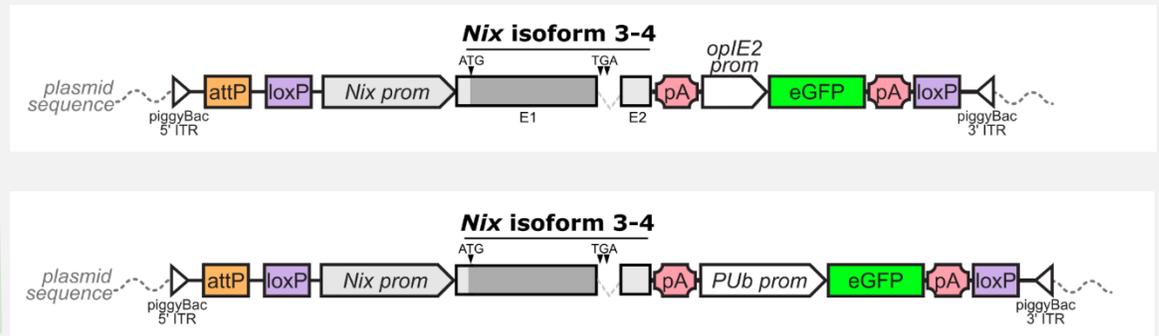
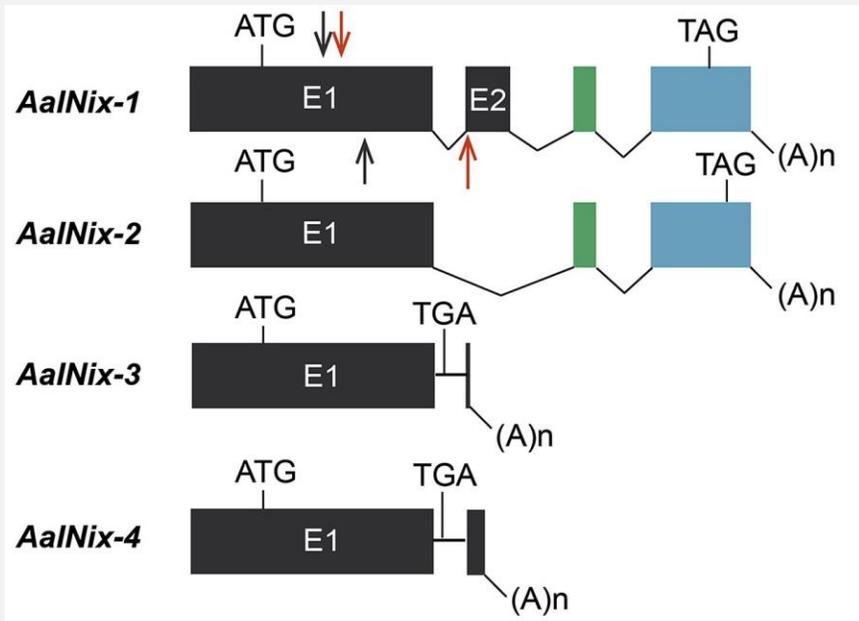
Sequencing / PCR



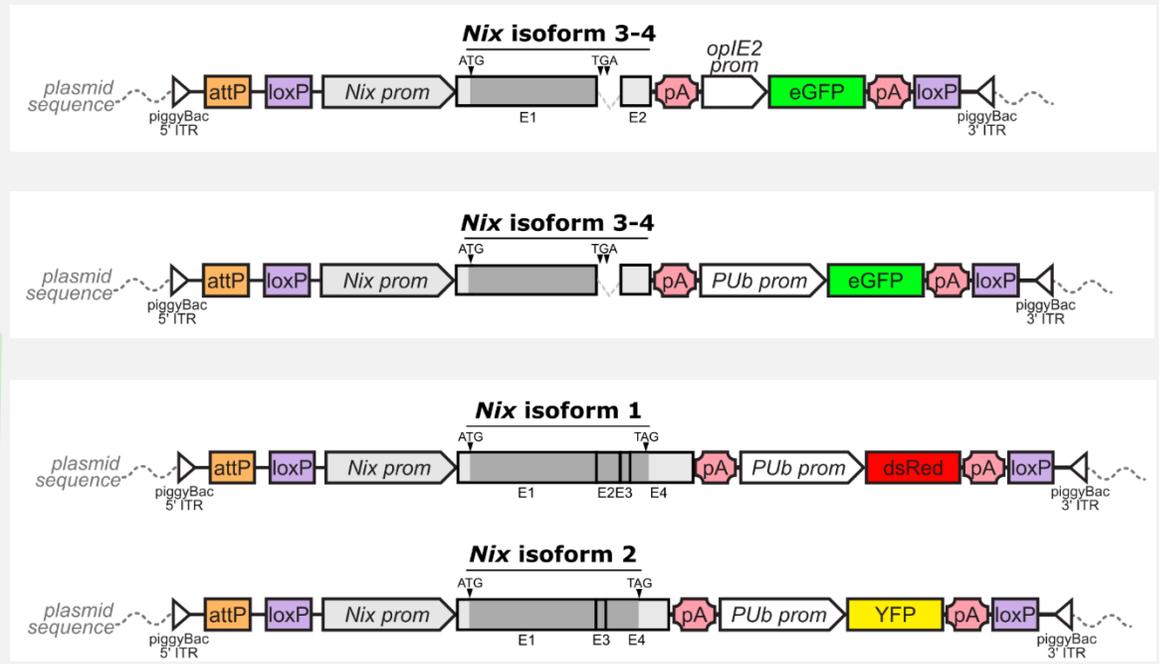
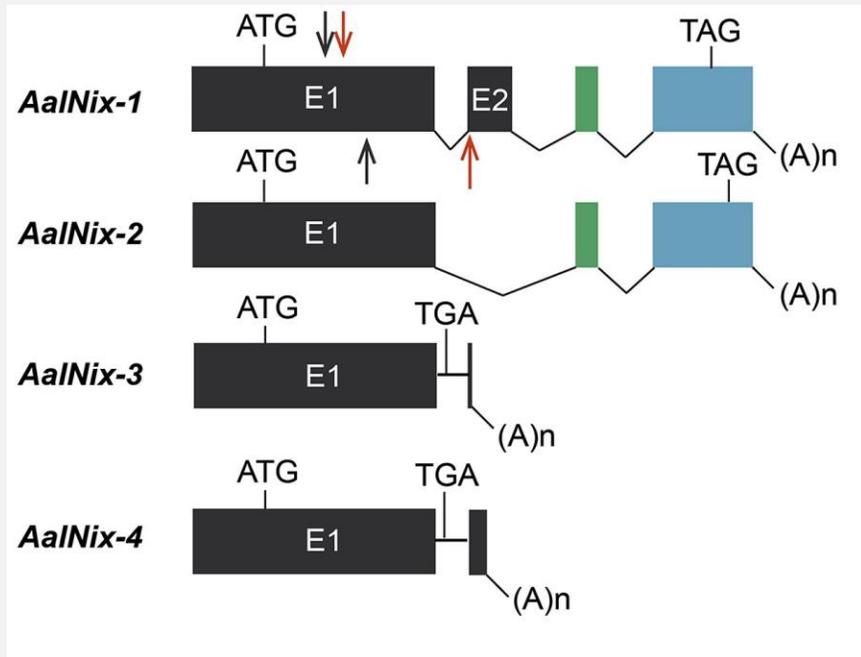
RESULTS – *Aedes albopictus*



RESULTS – *Aedes albopictus*



RESULTS – *Aedes albopictus*



RESULTS – *Aedes albopictus*

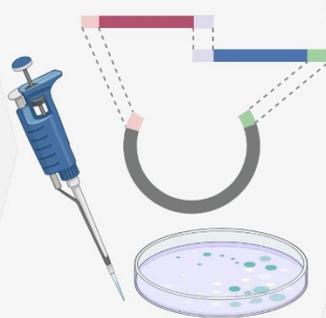
> 100 transgenic individuals
~85% of males



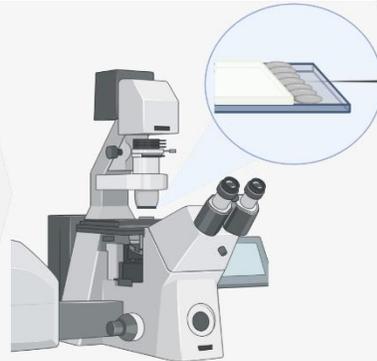
Design



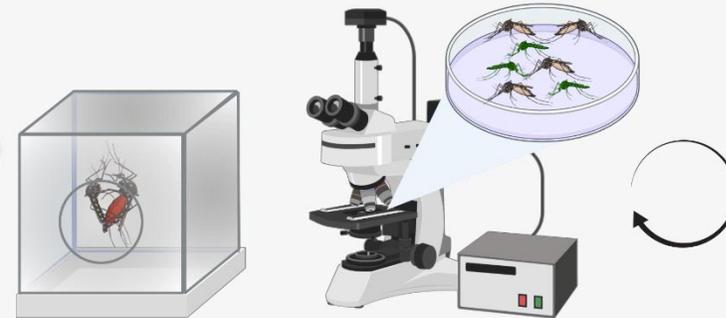
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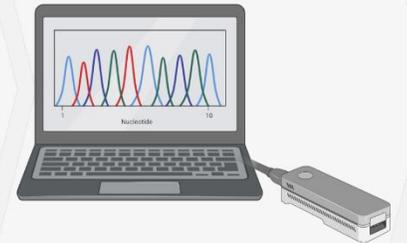
Microinjection



Crossing, Screening, Repeat

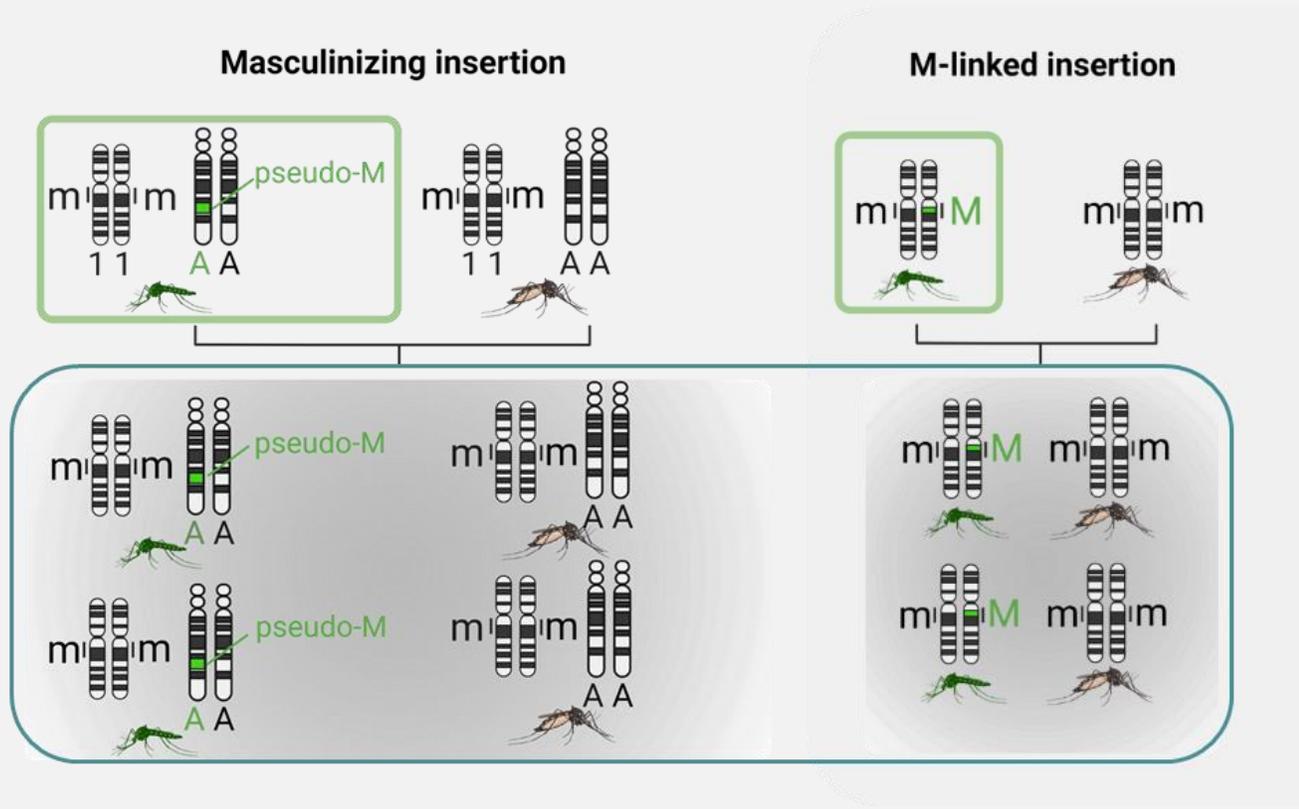


Sequencing / PCR



RESULTS – *Aedes albopictus*

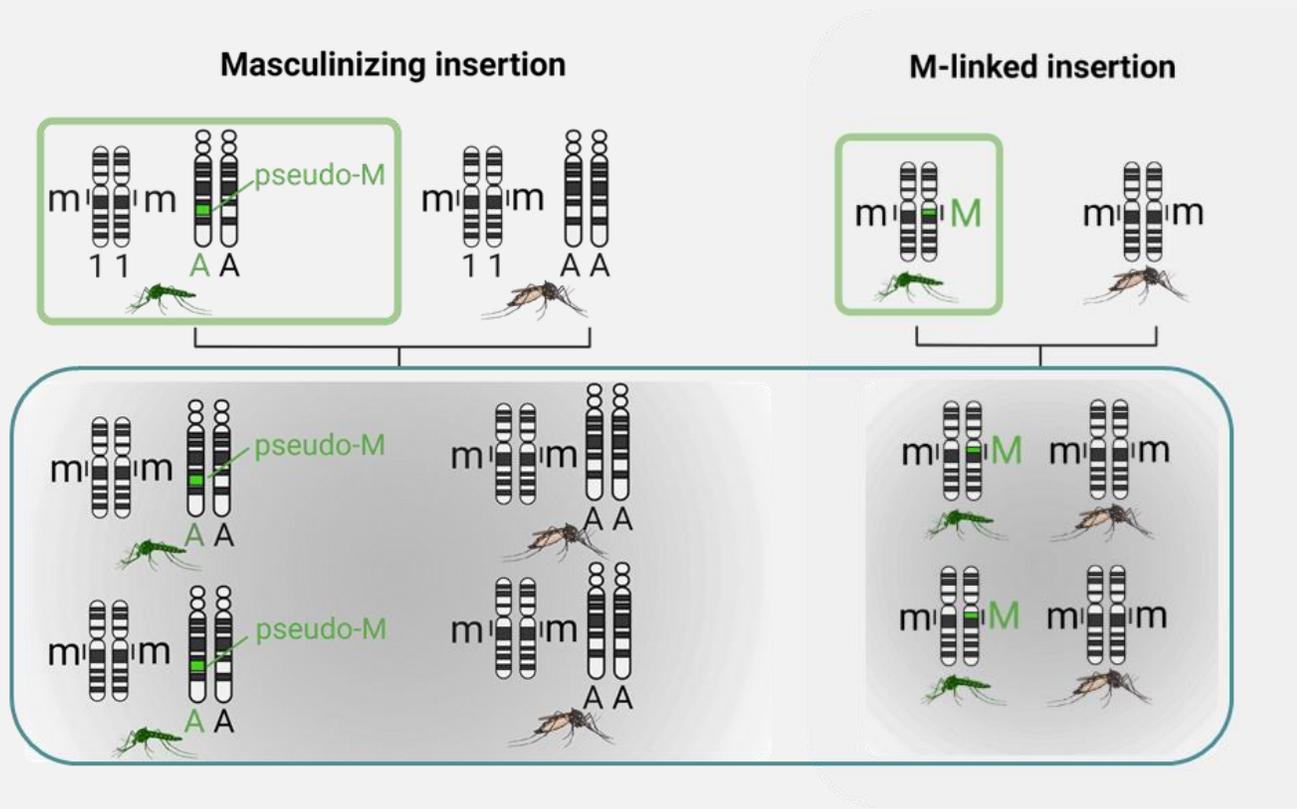
13 single-male lines
with 100% male fluorescence



RESULTS – *Aedes albopictus*

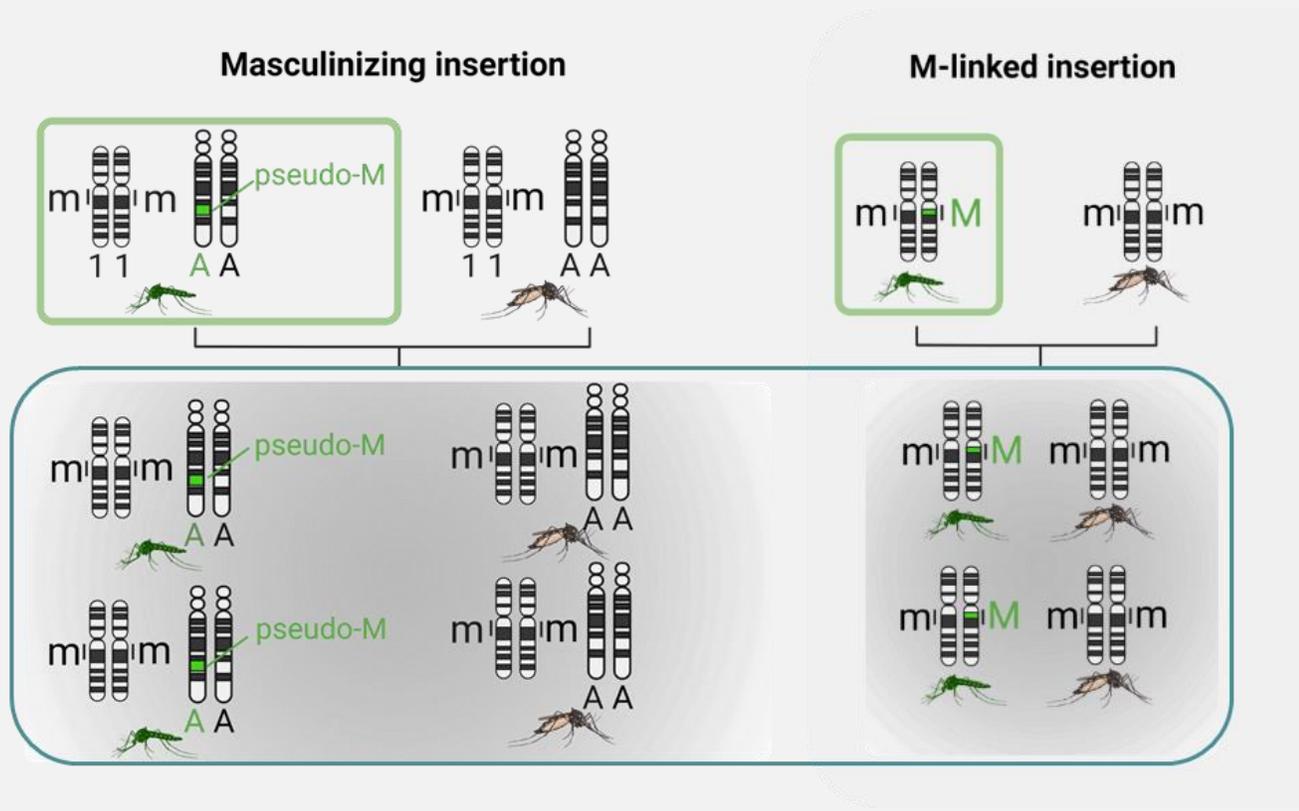
13 single-male lines
with 100% male fluorescence

Amplification of the
endogenous *Nix* gene



RESULTS – *Aedes albopictus*

13 single-male lines
with 100% male fluorescence

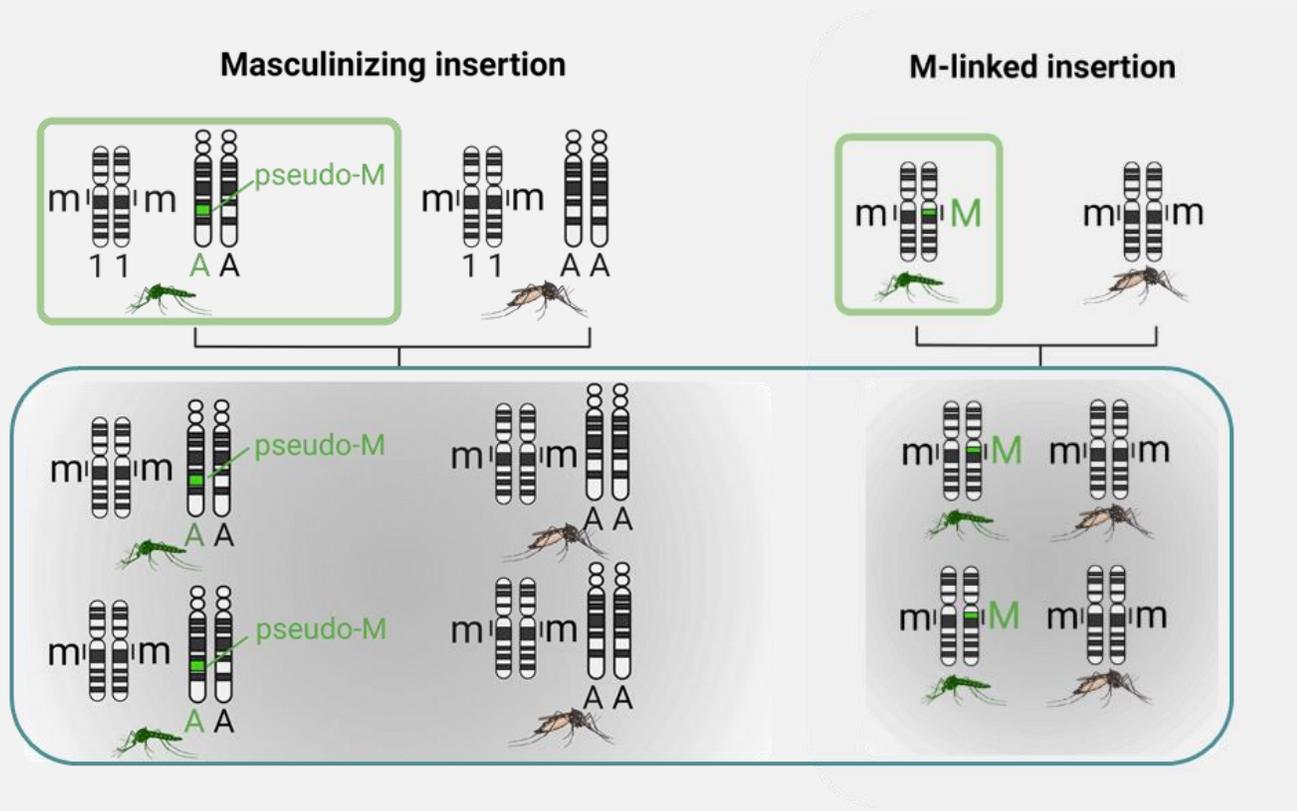


Amplification of the
endogenous *Nix* gene

4 out of 13 were
pseudo-M linked (GFP)

RESULTS – *Aedes albopictus*

13 single-male lines
with 100% male fluorescence



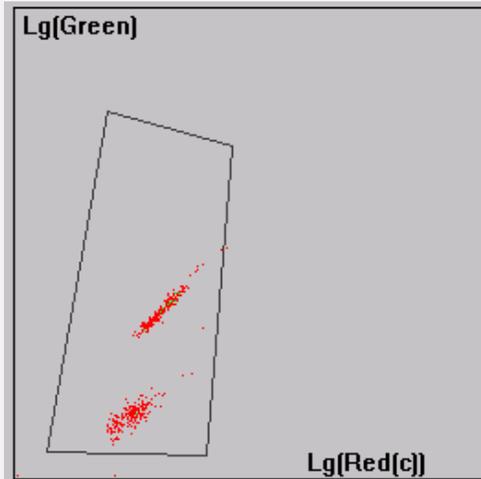
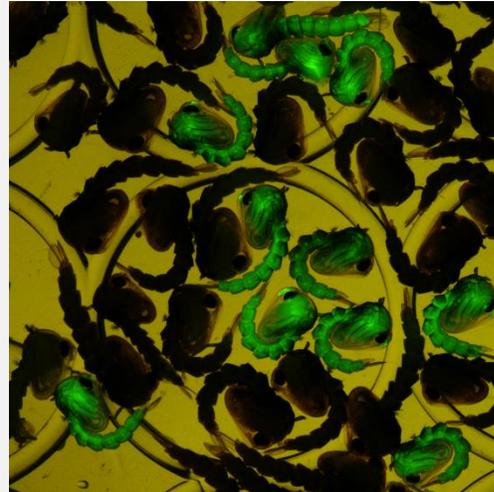
Amplification of the
endogenous *Nix* gene

4 out of 13 were
pseudo-M linked (GFP)

7 out of 13 were
M linked
(YFP and/or DsRed)

RESULTS – *Aedes albopictus*

Selection of an M-linked GSS



Amplification of the endogenous *Nix* gene

4 out of 13 were pseudo-M linked (GFP)

7 out of 13 were M linked (YFP and/or DsRed)

RESULTS – *Aedes albopictus*

Selection of an M-linked GSS

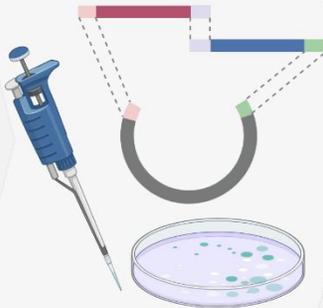
Mapped to scaffold 16



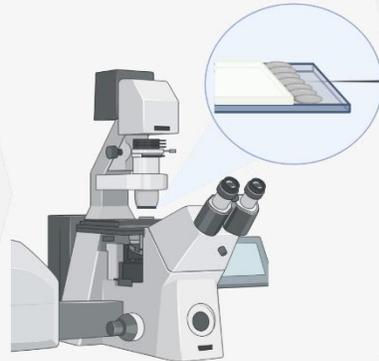
Design



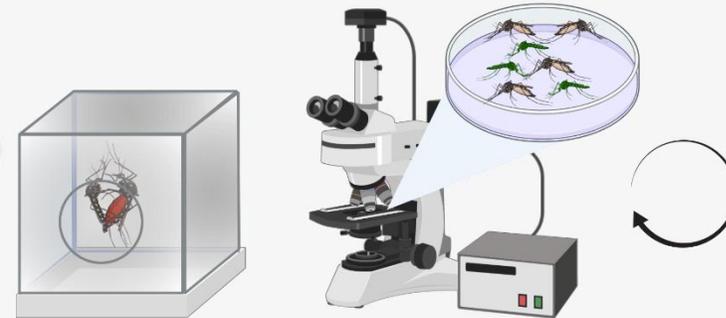
Cloning



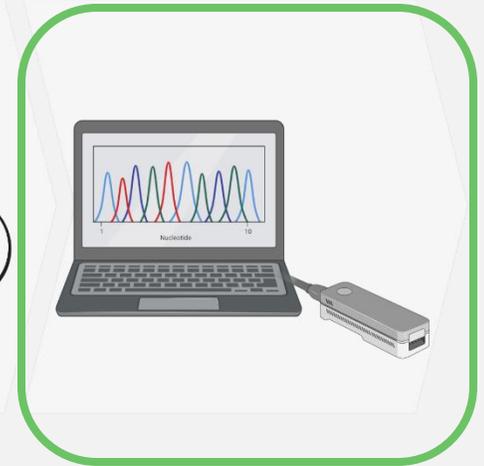
Microinjection



Crossing, Screening, Repeat



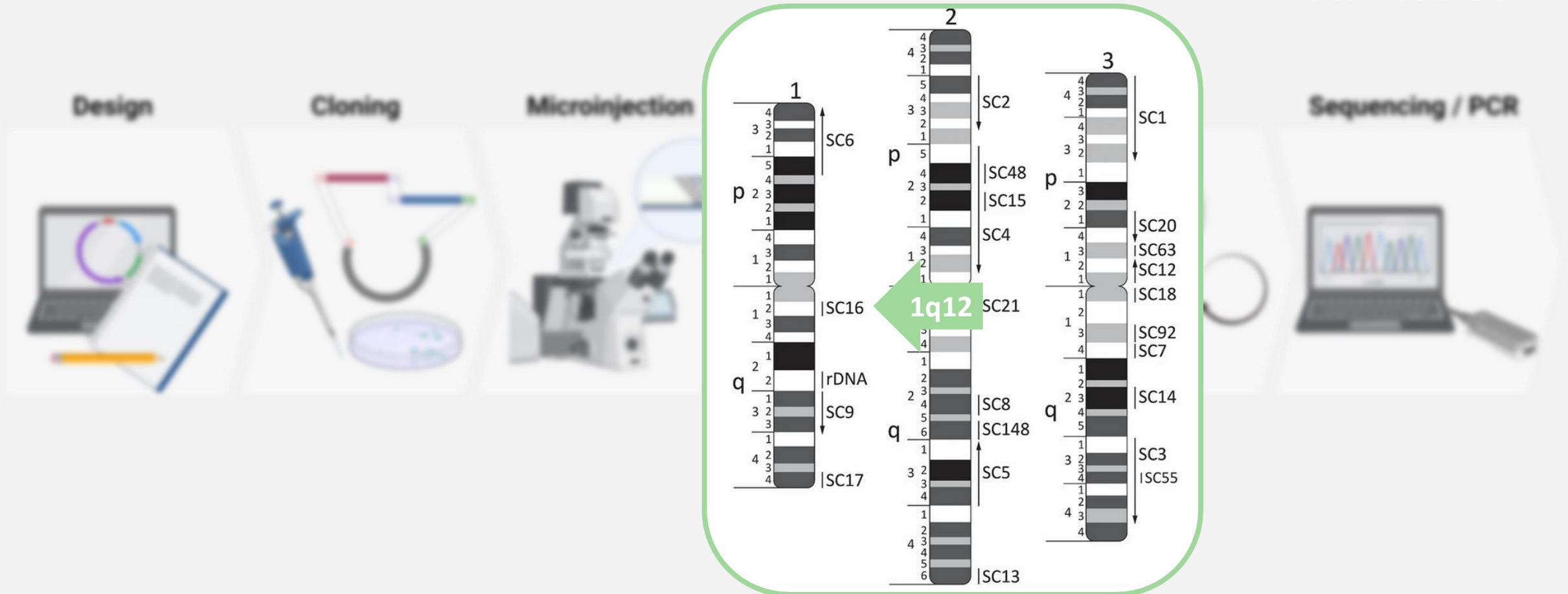
Sequencing / PCR



RESULTS – *Aedes albopictus*

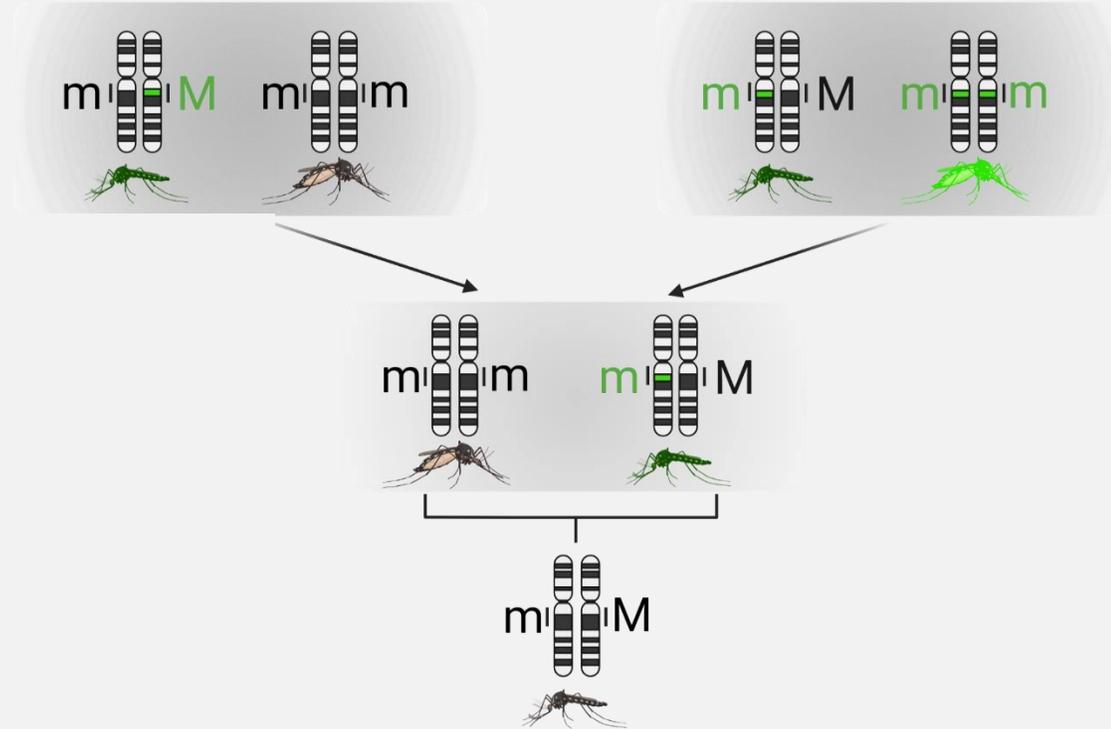
Selection of an M-linked GSS

Mapped to scaffold I6



RESULTS – *Aedes albopictus*

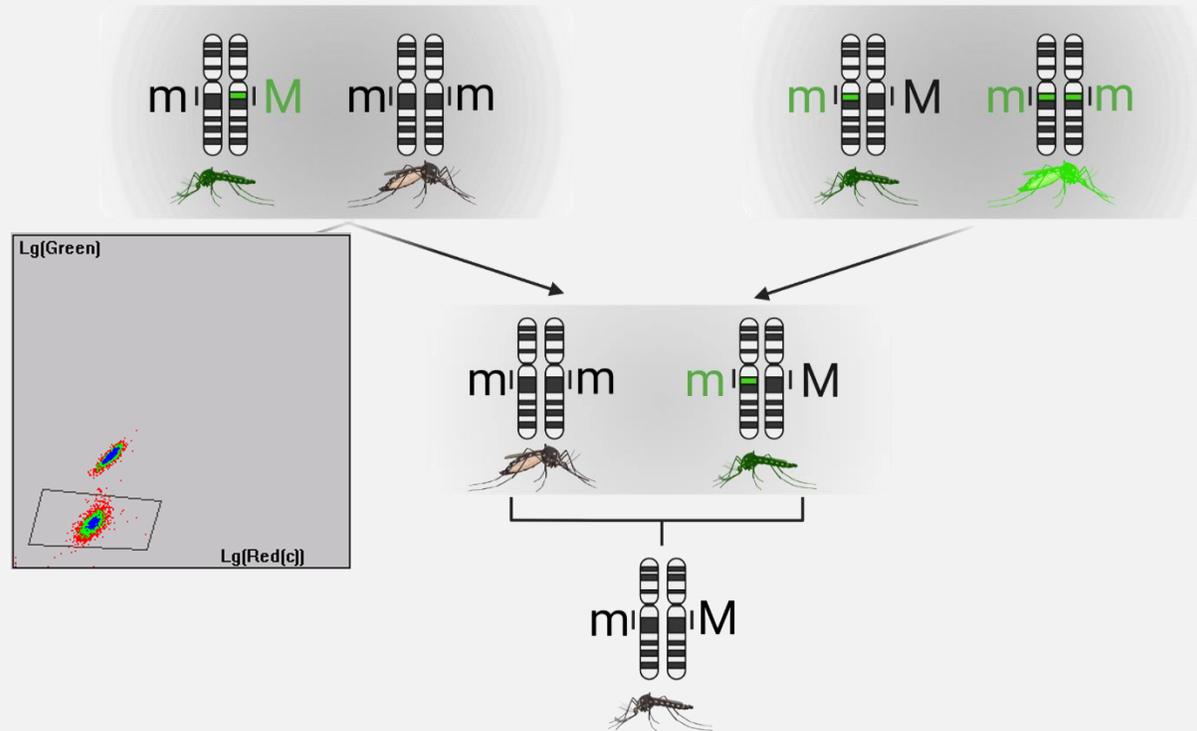
Obtaining non-transgenic males



RESULTS – *Aedes albopictus*

Obtaining non-transgenic males

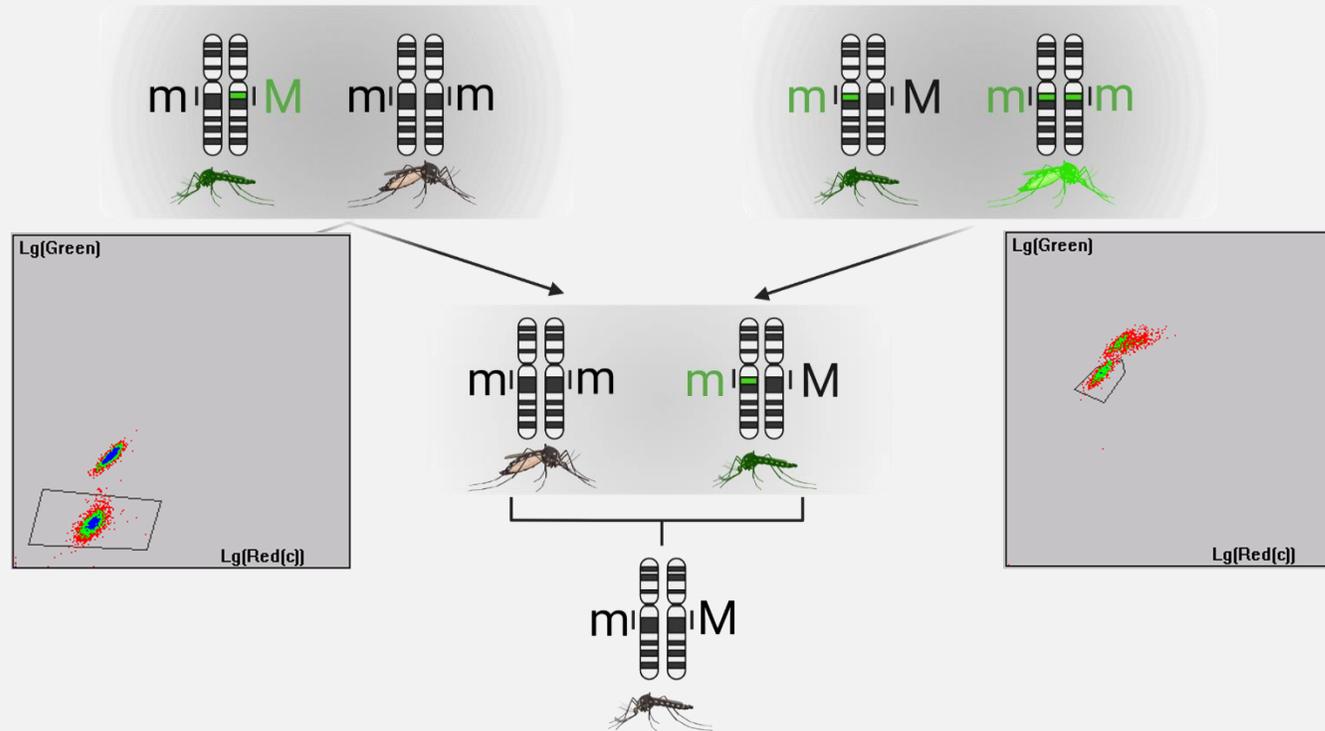
850 non-transgenic females from the M-linked GSS



RESULTS – *Aedes albopictus*

Obtaining non-transgenic males

850 non-transgenic females from the M-linked GSS



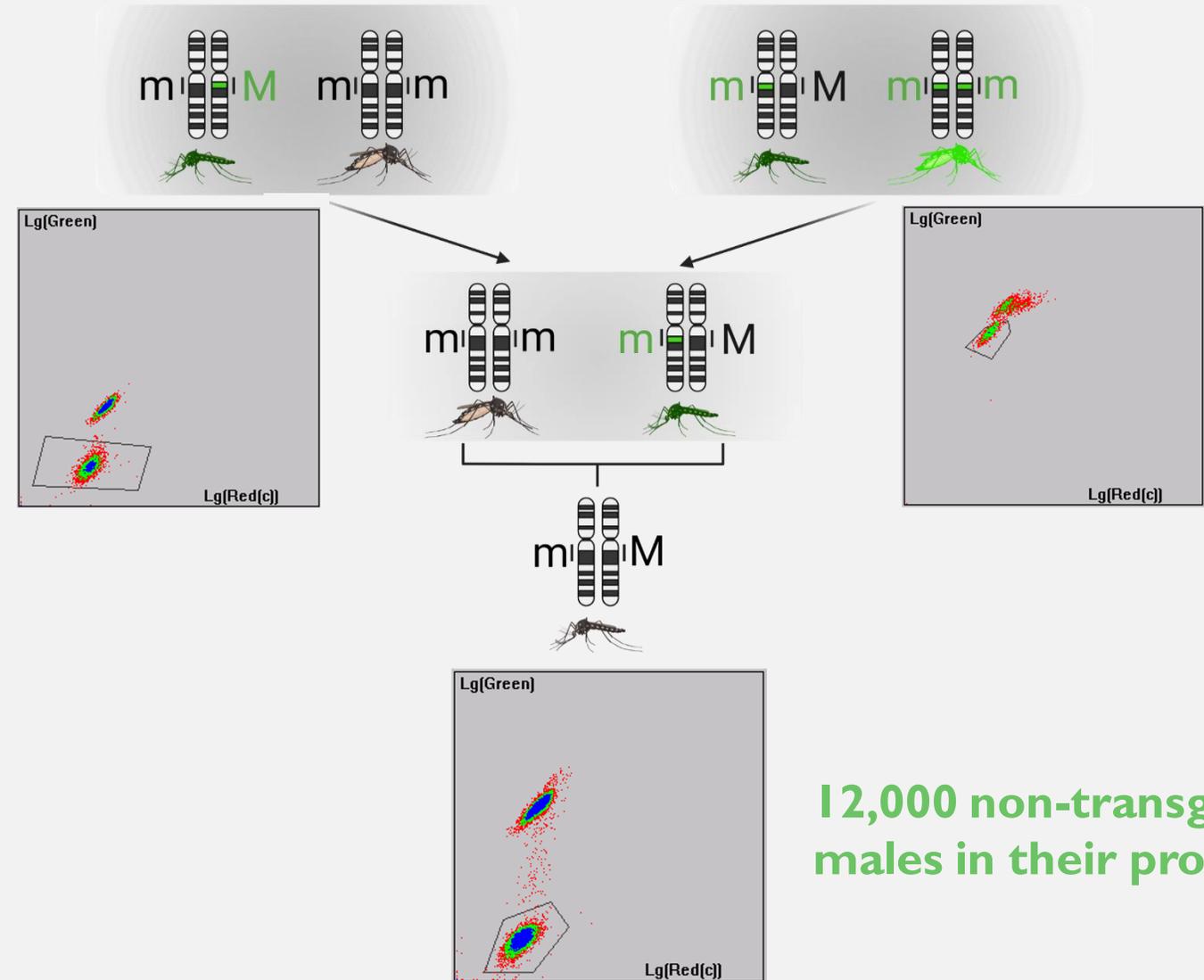
250 hemizygous transgenic males from the m-linked GSS

RESULTS – *Aedes albopictus*

Obtaining non-transgenic males

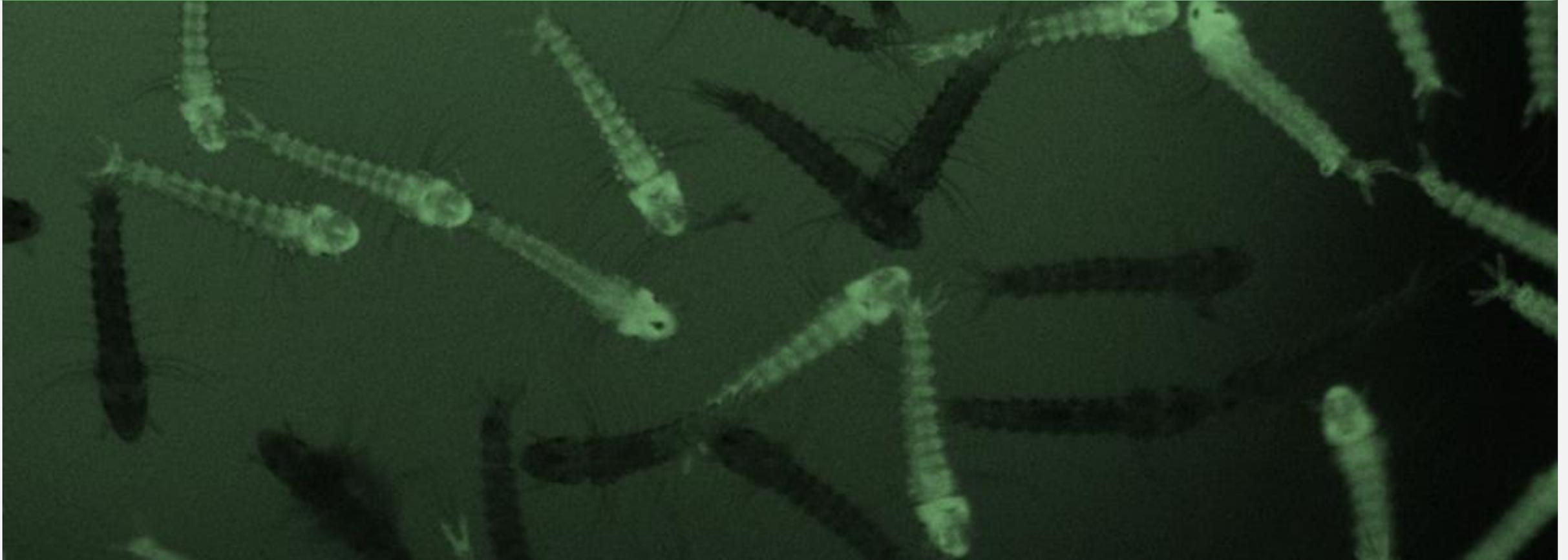
850 non-transgenic females from the M-linked GSS

250 hemizygous transgenic males from the m-linked GSS



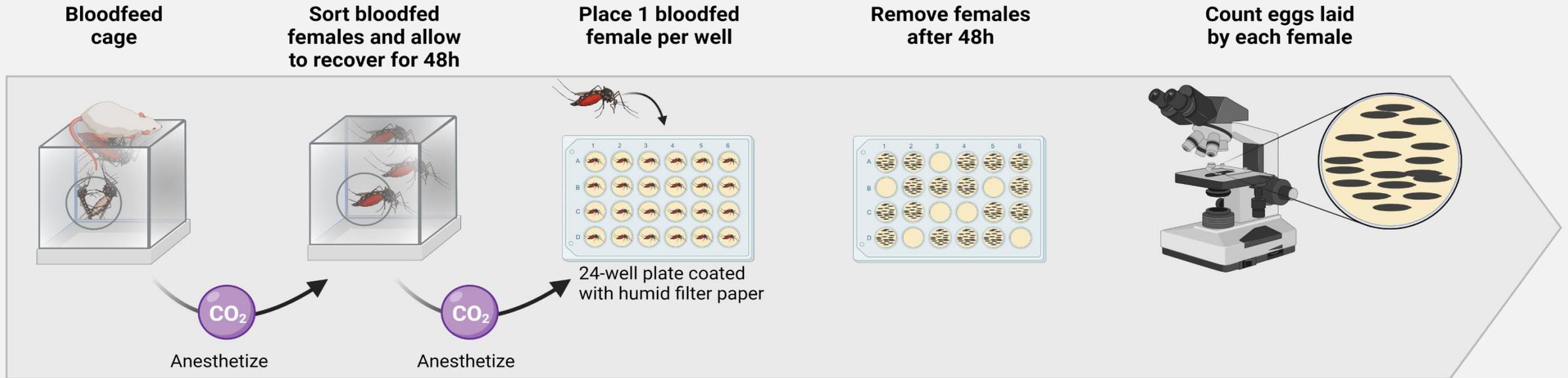
12,000 non-transgenic males in their progeny

RESULTS – PERFORMANCE OF THE GSS



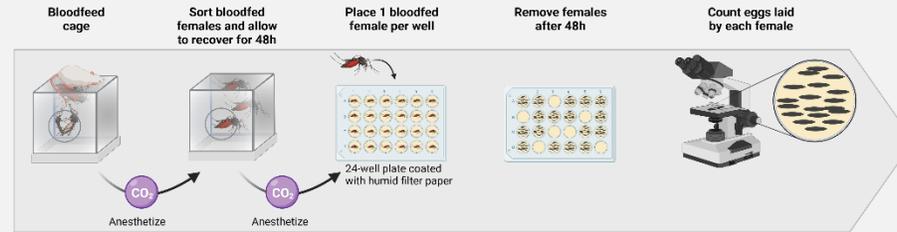
RESULTS – PERFORMANCE OF THE GSS

Fecundity

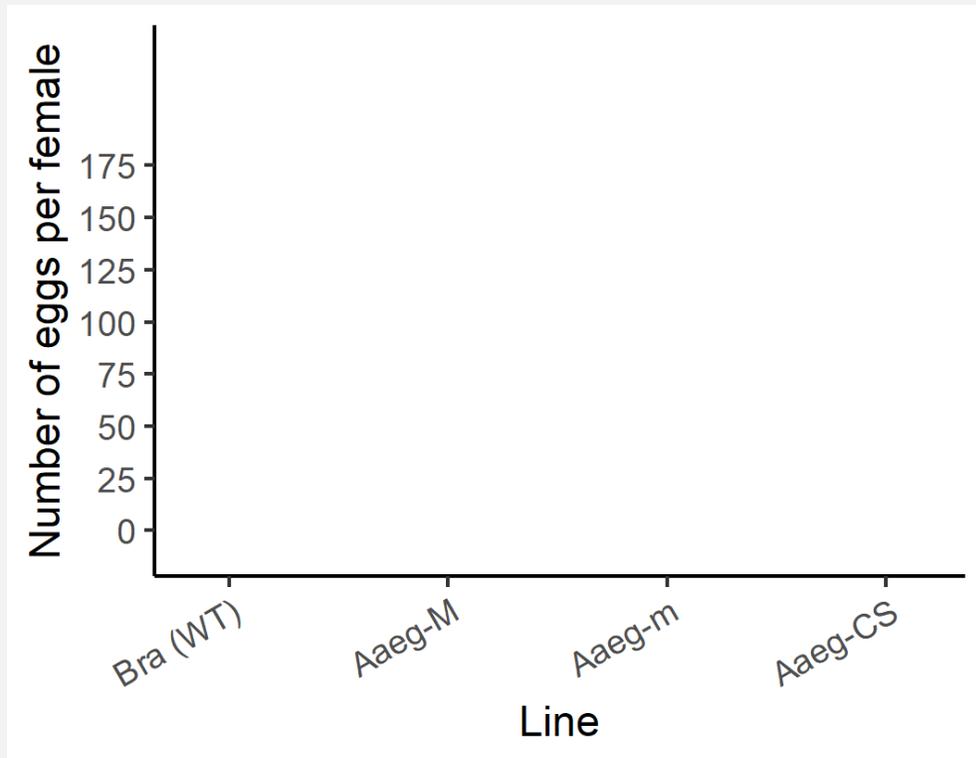


RESULTS – PERFORMANCE OF THE GSS

Fecundity

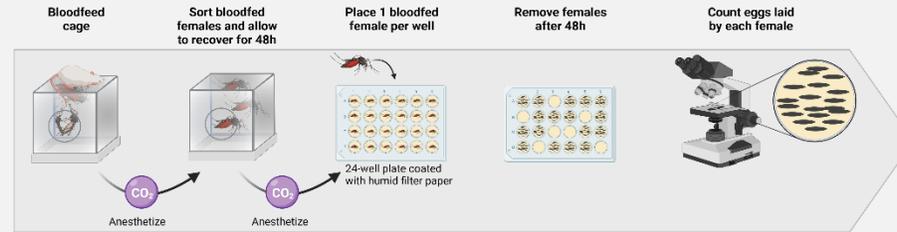


Aedes aegypti

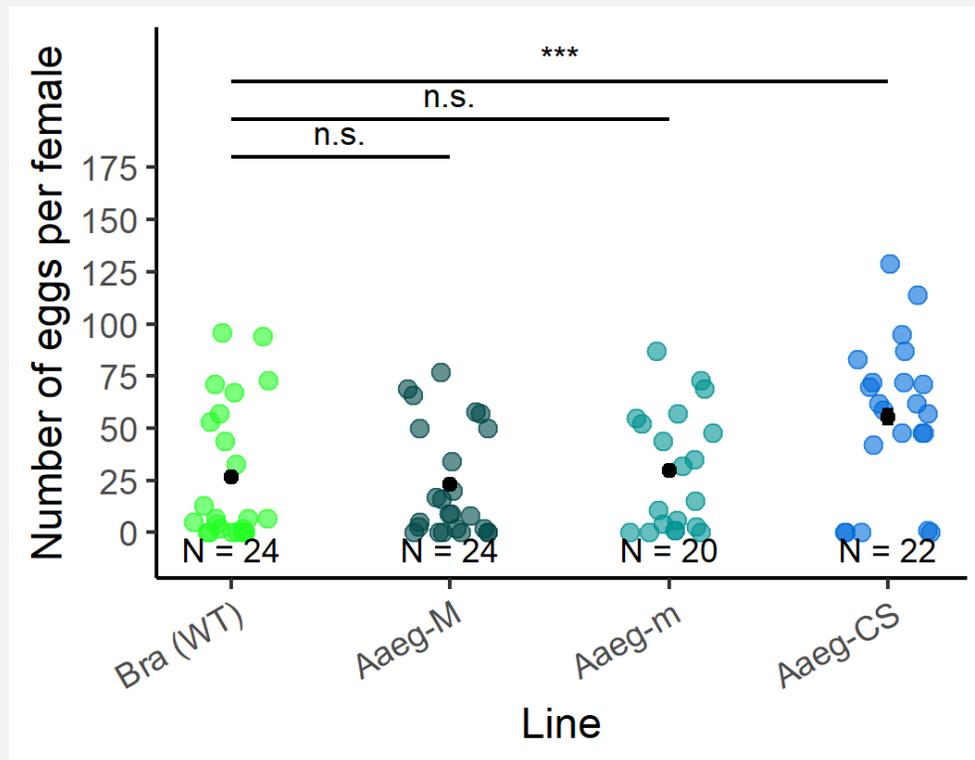


RESULTS – PERFORMANCE OF THE GSS

Fecundity

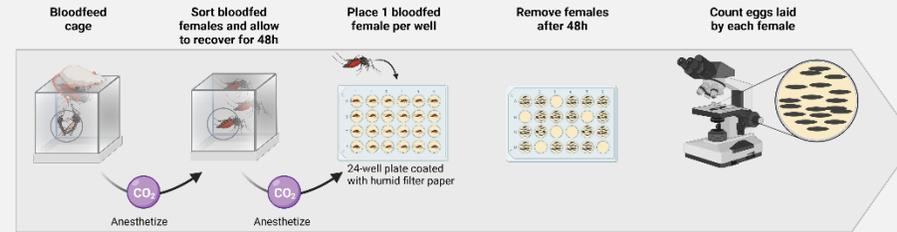


Aedes aegypti

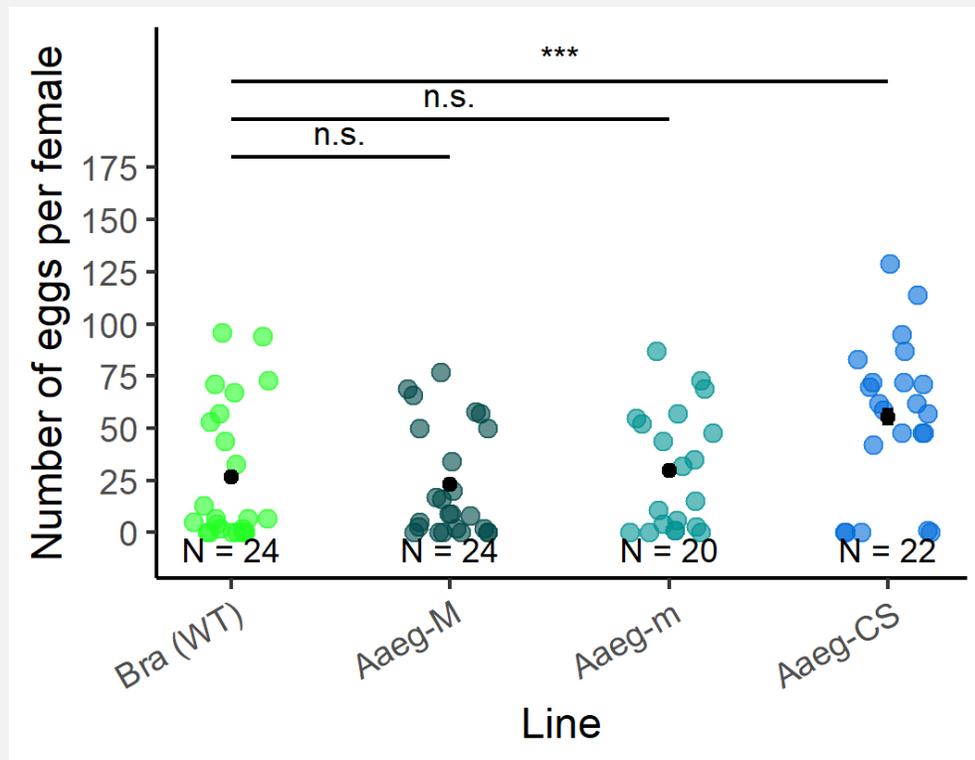


RESULTS – PERFORMANCE OF THE GSS

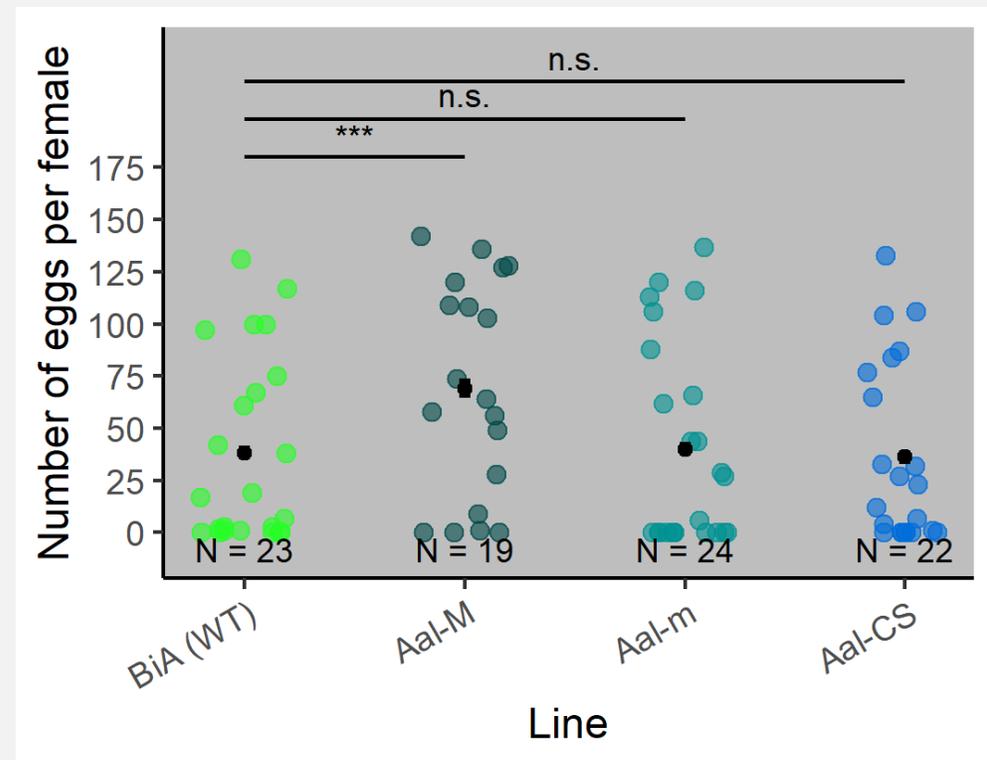
Fecundity



Aedes aegypti



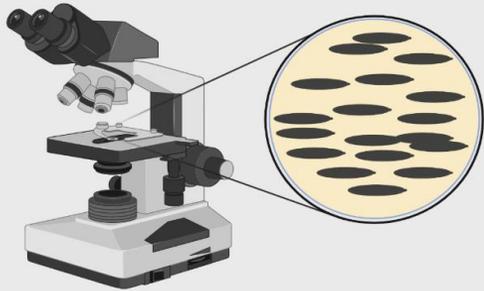
Aedes albopictus



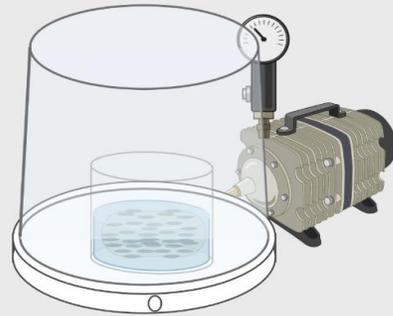
RESULTS – PERFORMANCE OF THE GSS

Egg hatching rate

Count eggs

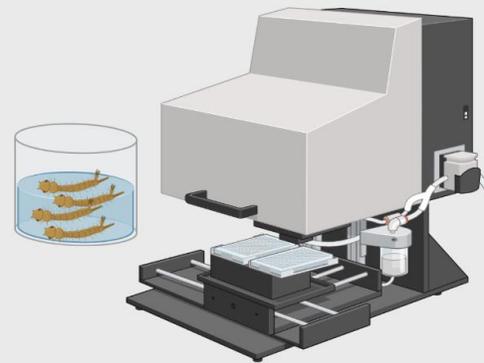


Submerge eggs
and apply a vacuum

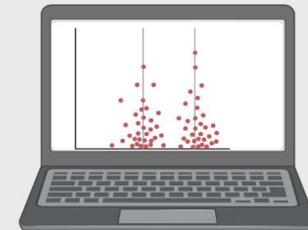


30' vacuum

Count larvae using
COPAS

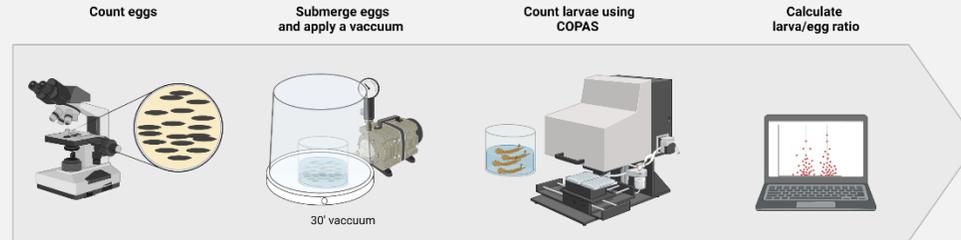


Calculate
larva/egg ratio

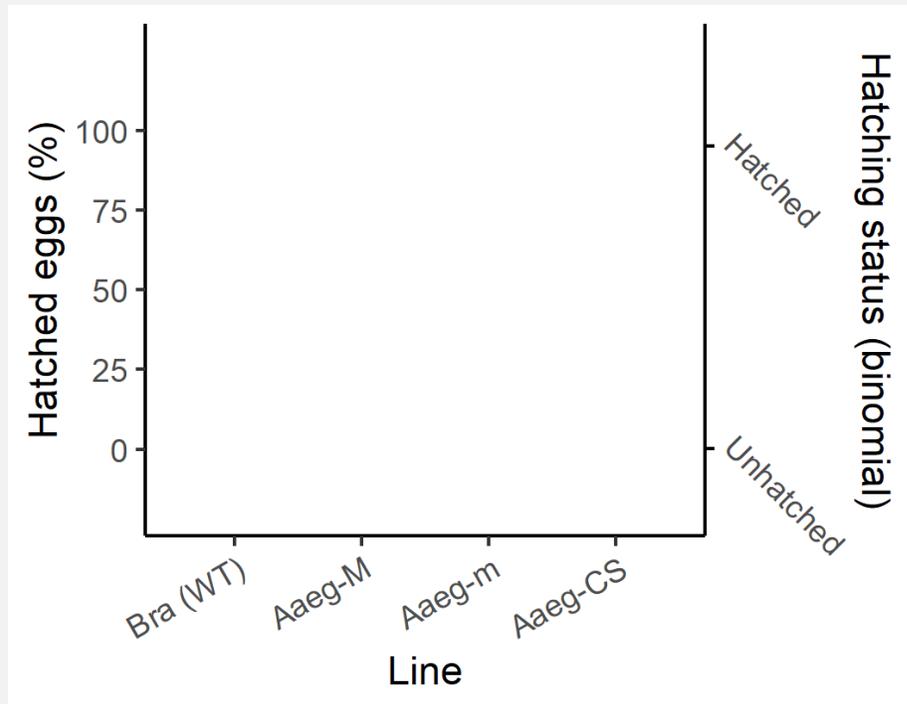


RESULTS – PERFORMANCE OF THE GSS

Egg hatching rate

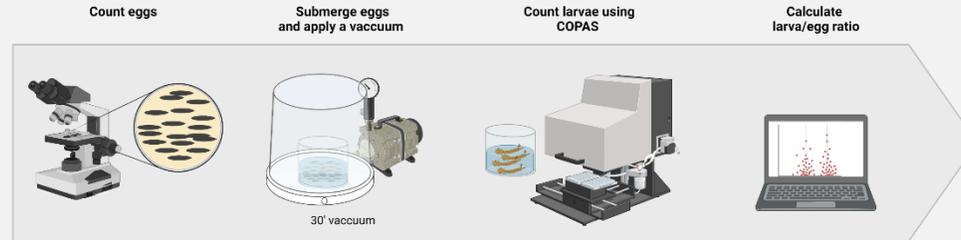


Aedes aegypti

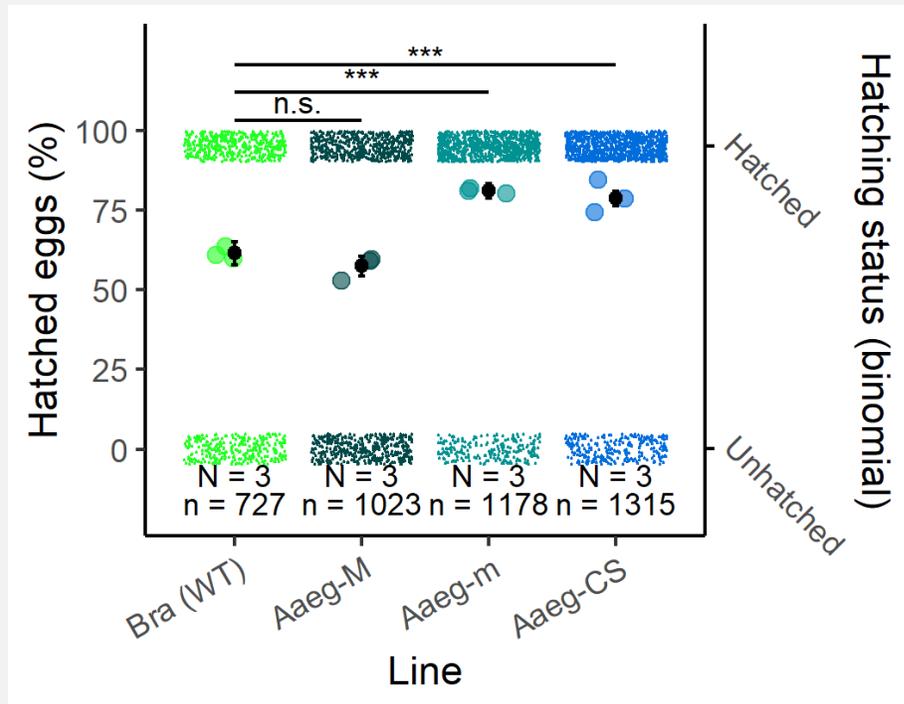


RESULTS – PERFORMANCE OF THE GSS

Egg hatching rate

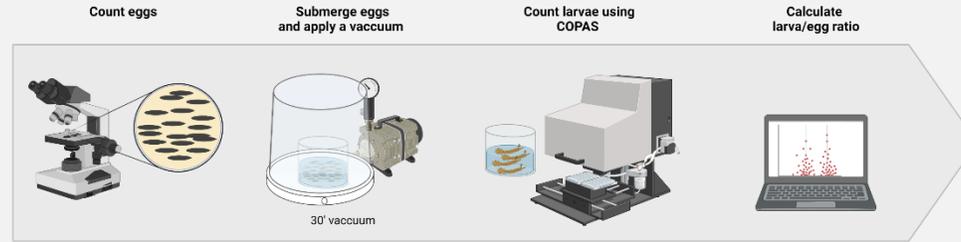


Aedes aegypti

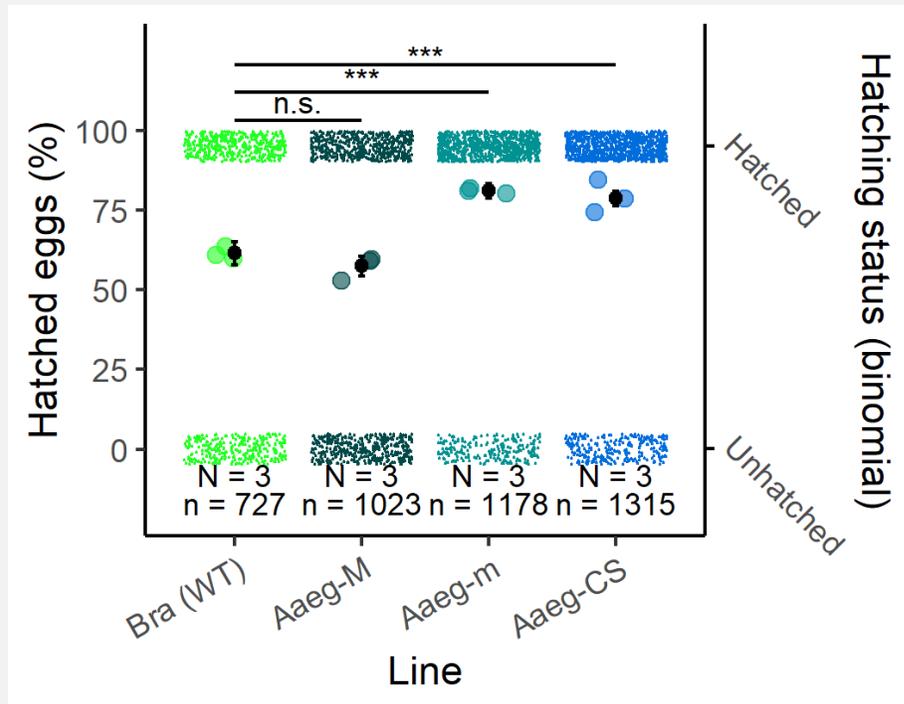


RESULTS – PERFORMANCE OF THE GSS

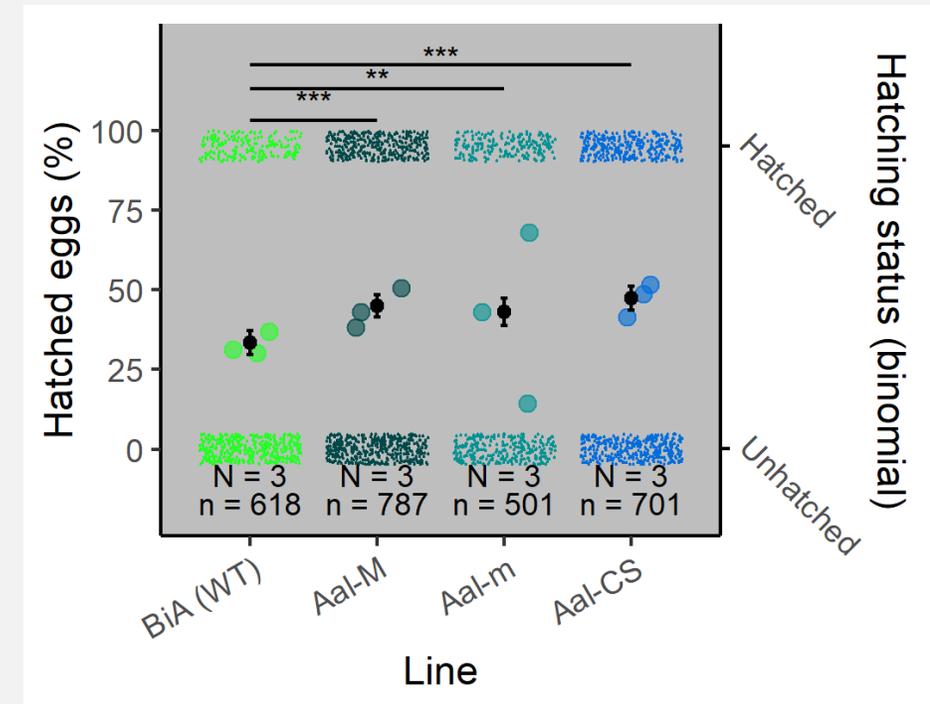
Egg hatching rate



Aedes aegypti



Aedes albopictus



RESULTS – PERFORMANCE OF THE GSS

Male competitiveness

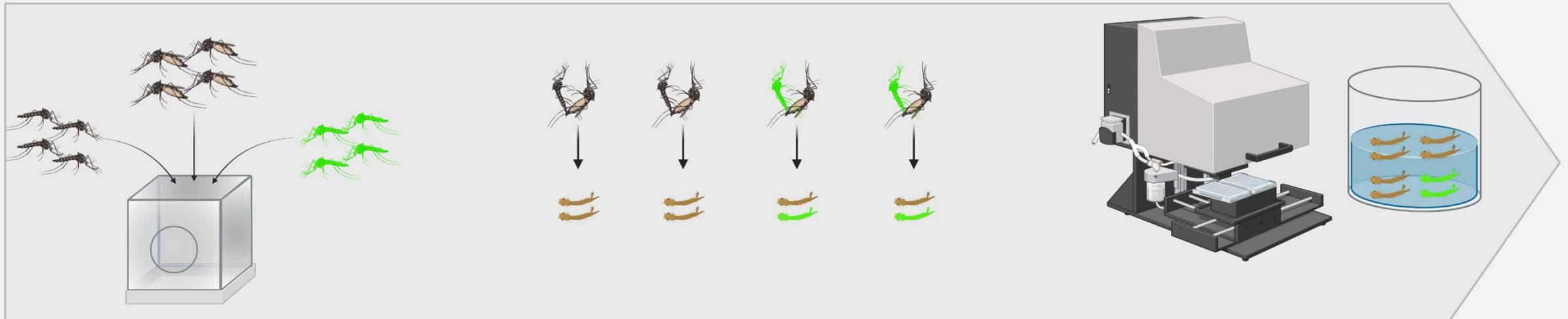
Mix equal numbers of WT females,
WT males and transgenic males

Equal competitiveness

50% of the females mate with a WT male
50% of the females mate with a transgenic male

Result:

75% non-transgenic
25% transgenic



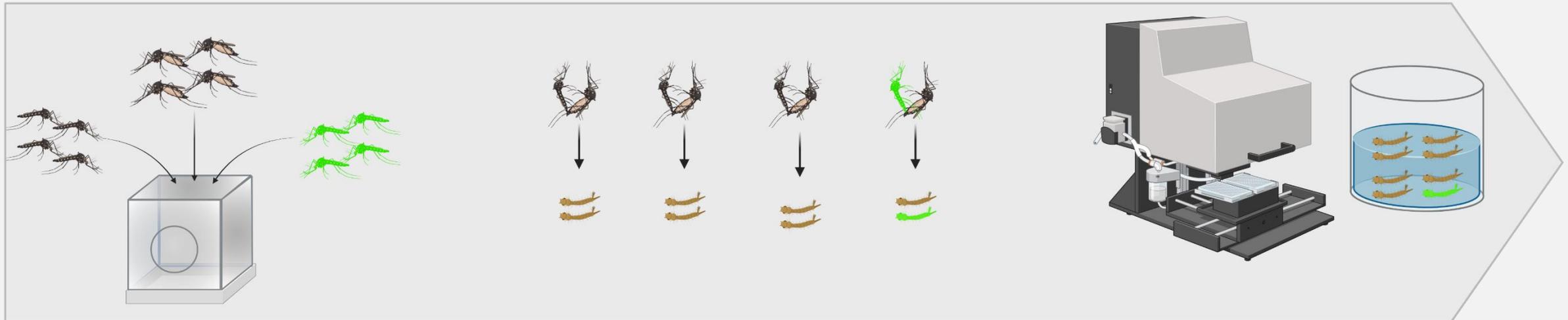
RESULTS – PERFORMANCE OF THE GSS

Male competitiveness

Mix equal numbers of WT females, WT males and transgenic males

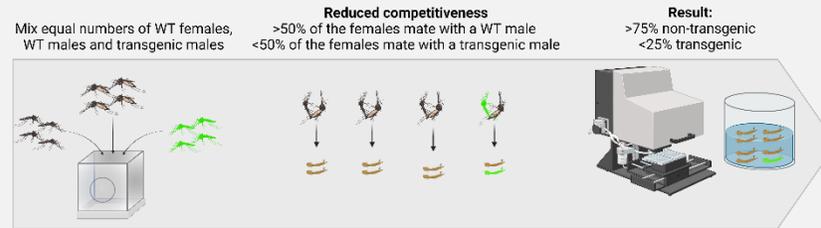
Reduced competitiveness
>50% of the females mate with a WT male
<50% of the females mate with a transgenic male

Result:
>75% non-transgenic
<25% transgenic

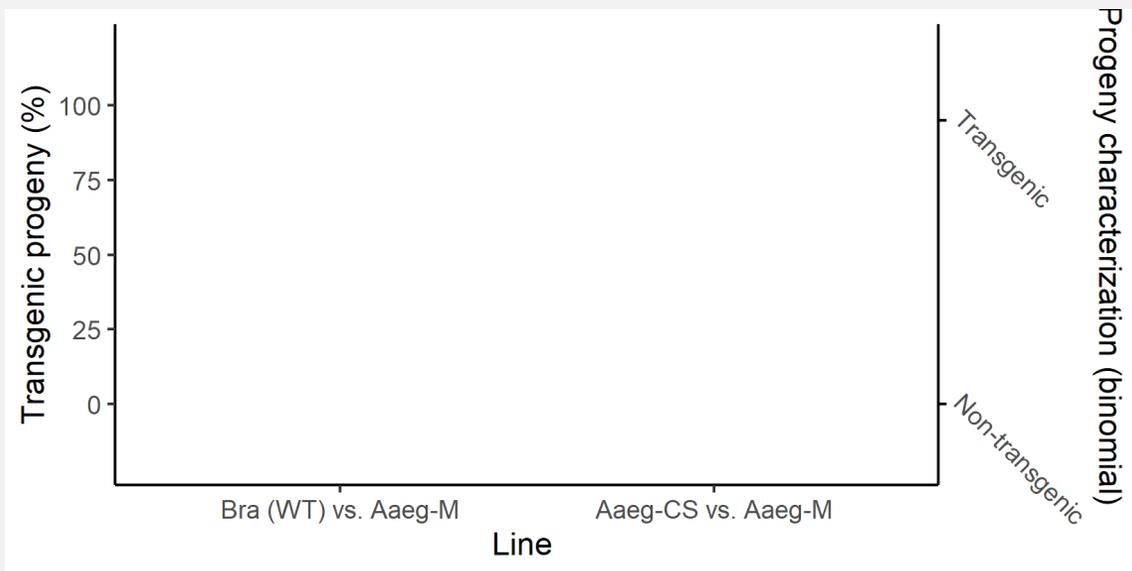


RESULTS – PERFORMANCE OF THE GSS

Male competitiveness

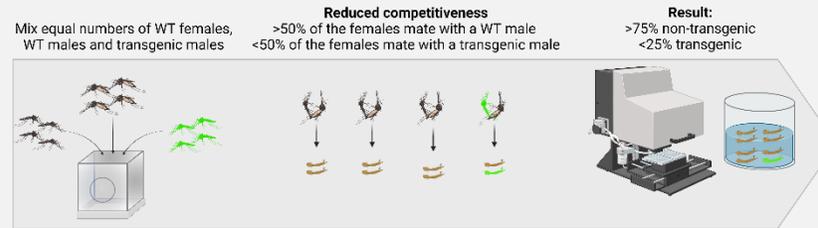


Aedes aegypti

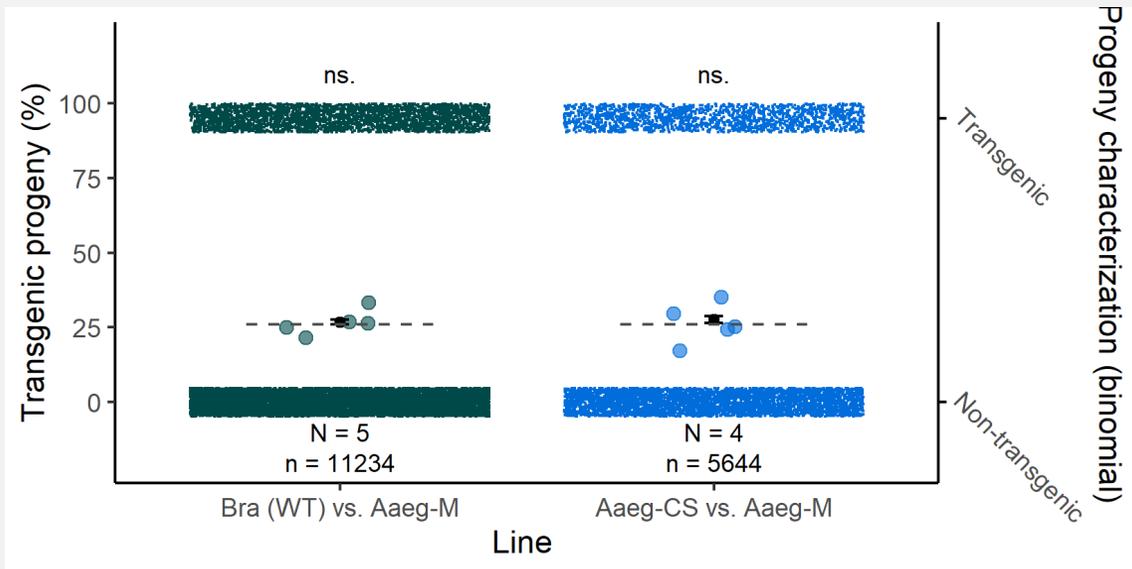


RESULTS – PERFORMANCE OF THE GSS

Male competitiveness

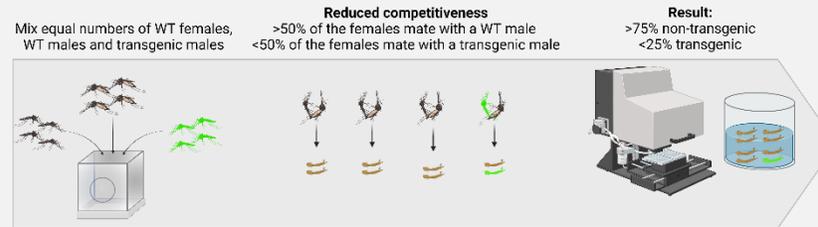


Aedes aegypti

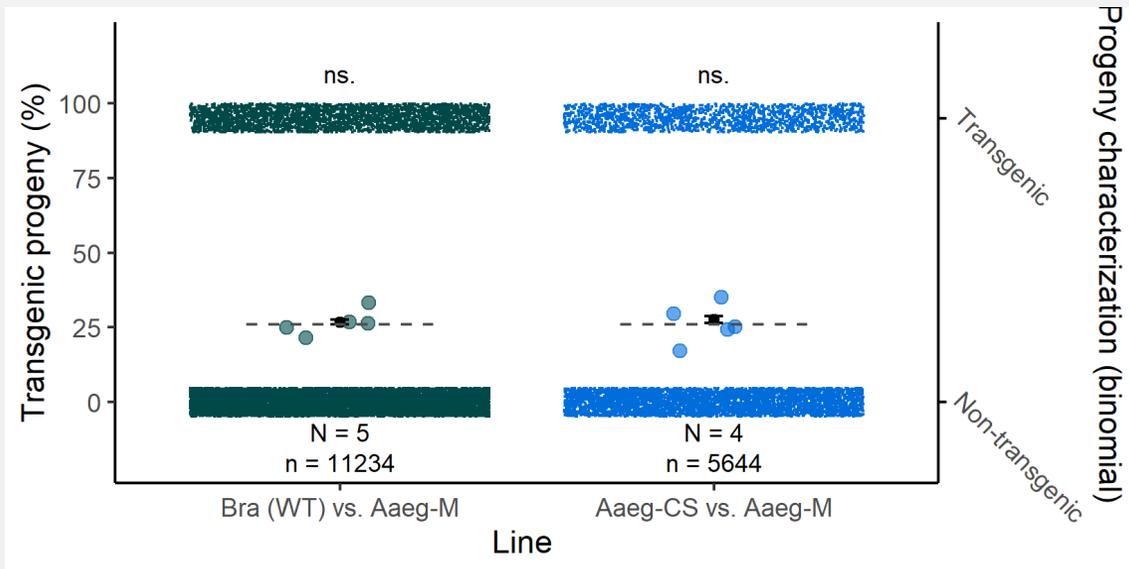


RESULTS – PERFORMANCE OF THE GSS

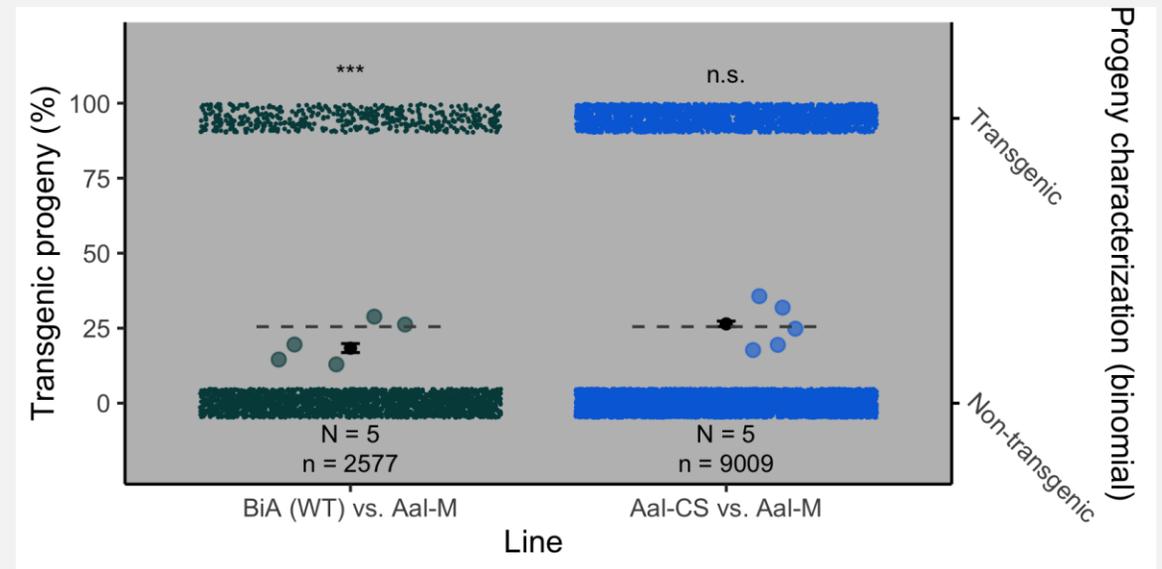
Male competitiveness



Aedes aegypti



Aedes albopictus



RESULTS

Valorisation

communications biology

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Transgenic expression of *Nix* converts genetic females into males and allows automated sex sorting in *Aedes albopictus*

[Célia Lutrat](#) [✉](#), [Roénick P. Olmo](#), [Thierry Baldet](#), [Jérémy Bouyer](#) & [Eric Marois](#) [✉](#)

Combining two Genetic Sexing Strains allows sorting of non-transgenic males for *Aedes* genetic control

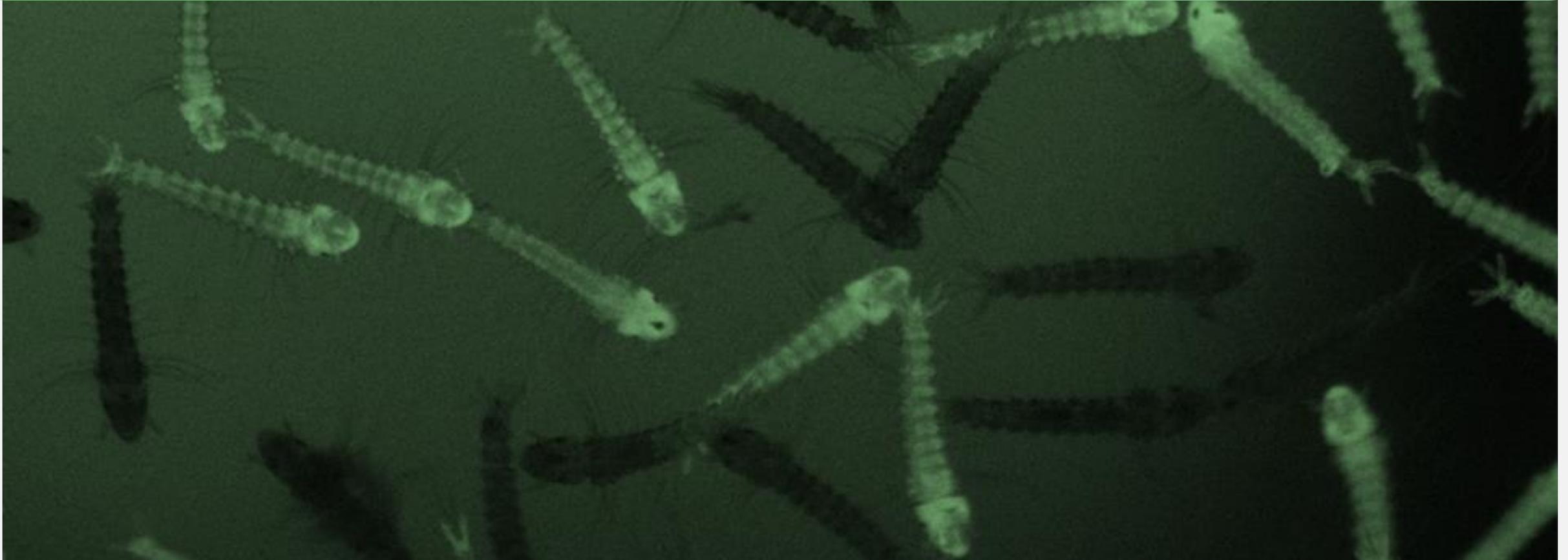
Authors

Célia Lutrat^{1,2,3,4,*}, Myriam Burekbuchler⁴, Roénick Proveti Olmo⁴, Rémy Beugnon^{5,6}, Albin Fontaine⁷, Thierry Baldet^{1,8}, Jérémy Bouyer^{1,9,10,†}, Eric Marois^{4,†,*}

Just out in Communications Biology!

Soon to be submitted

CONCLUSION



CONCLUSION

7 criteria for a good sexing method

Sorting stage

Male recovery

Female contamination

Sorting speed

Initial investment

Consumable cost

Acceptability

CONCLUSION

Evaluating our sexing method

Sorting stage
Neonate larvae

CONCLUSION

Evaluating our sexing method

Sorting stage
Neonate larvae

Male recovery
70%

CONCLUSION

Evaluating our sexing method

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Neonate larvae

Male recovery
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Female contamination
0-0.1%

CONCLUSION

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Sorting stage

Neonate larvae

Male recovery

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Female contamination

0-0.1%

Sorting speed

60 larvae / sec

CONCLUSION

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Initial investment

+++

CONCLUSION

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Consumable cost

Decreased

CONCLUSION

Evaluating our sexing method

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Initial investment

+++

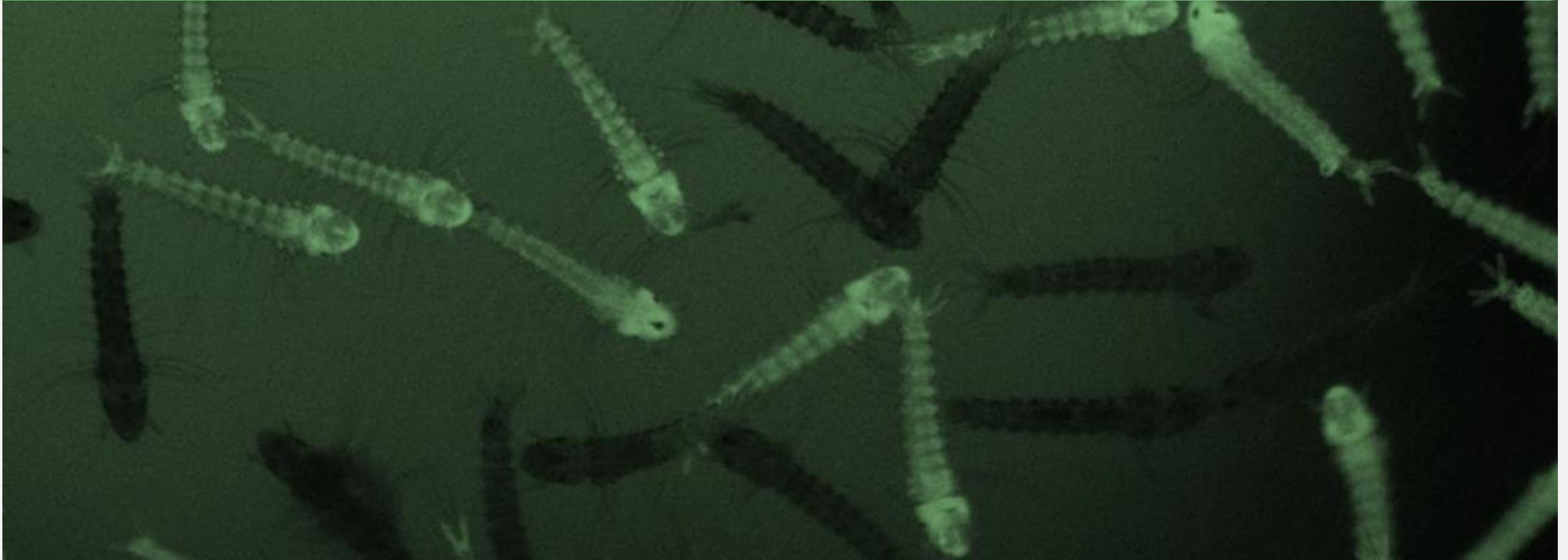
Consumable cost

Decreased

Acceptability

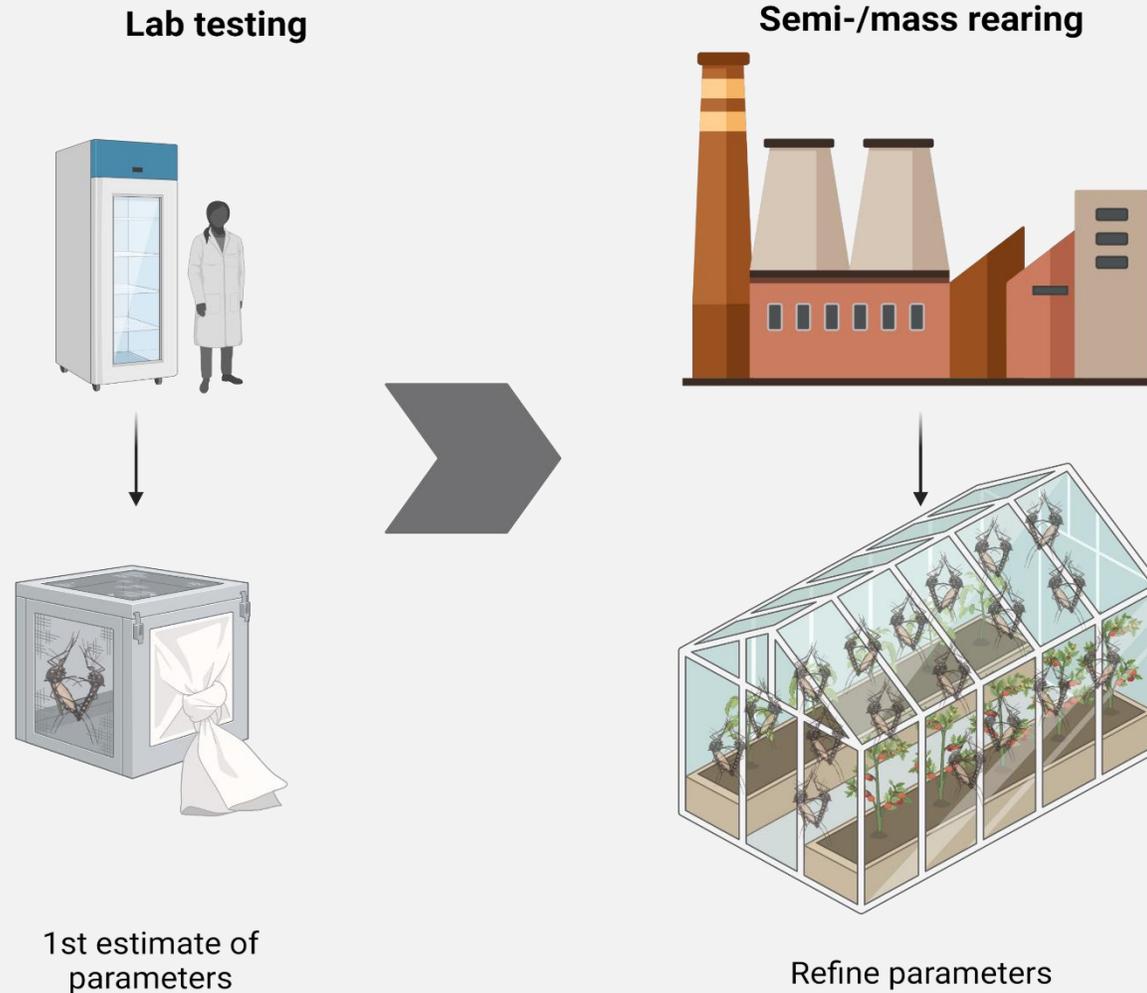
To be defined

PERSPECTIVE



PERSPECTIVE

Going further



ACKNOWLEDGEMENTS



ACKNOWLEDGEMENTS



CIRAD UMR ASTRE
Collectif vecteurs



IBMC UPR9022
Equipe moustiques

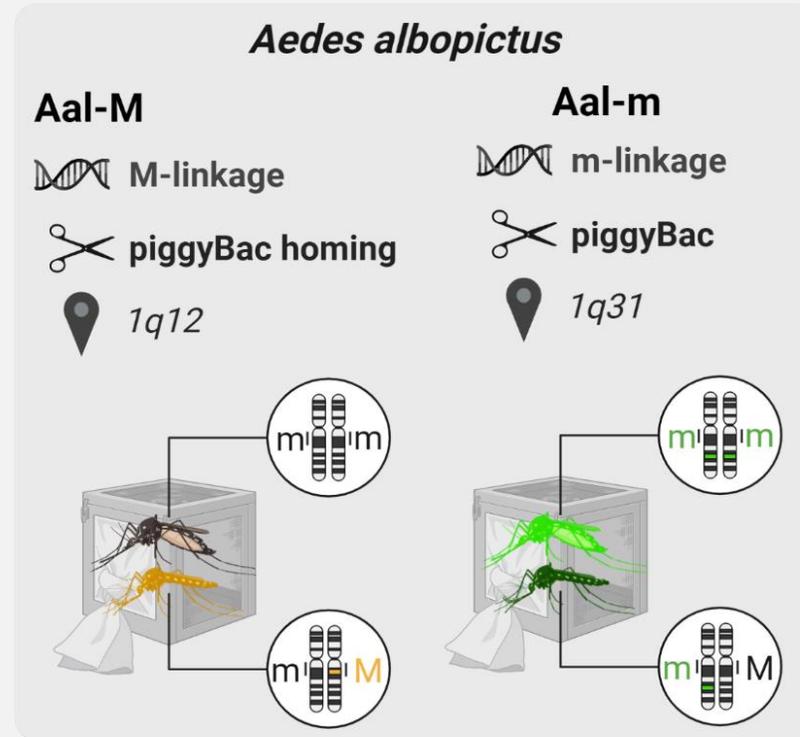
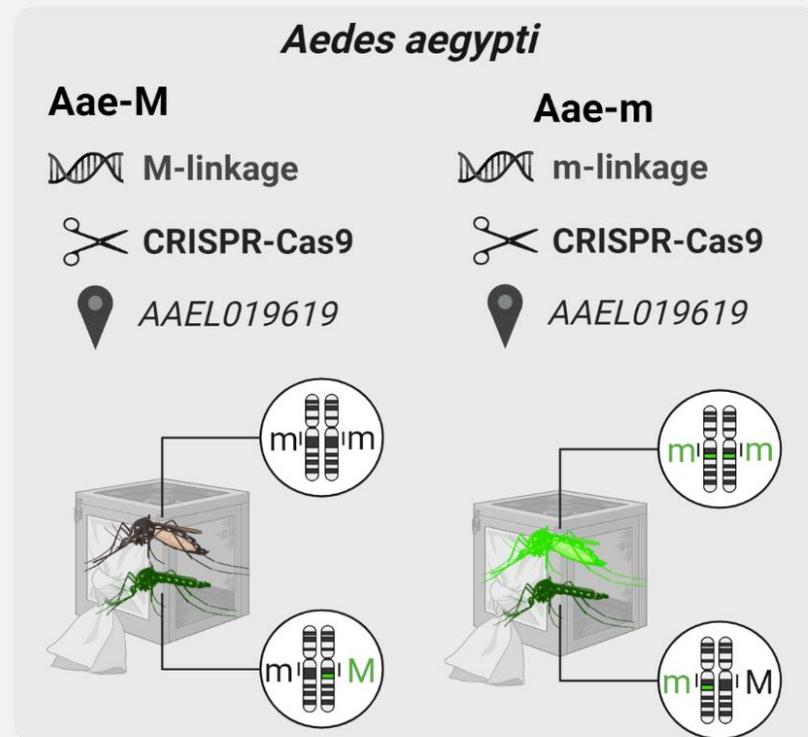
Graphical figures were designed on Biorender.com. Data figures were plotted on R.

Funding sources: EU ERC CoG—682387 REVOLINC to J.B., ANR grants #ANR-11-EQPX-0022, #ANR-19-CE35-0007 GDaMO and # 18-CE35-0003-02 BAKOUMBA to E.M. No funding was received from Union Biometrica.

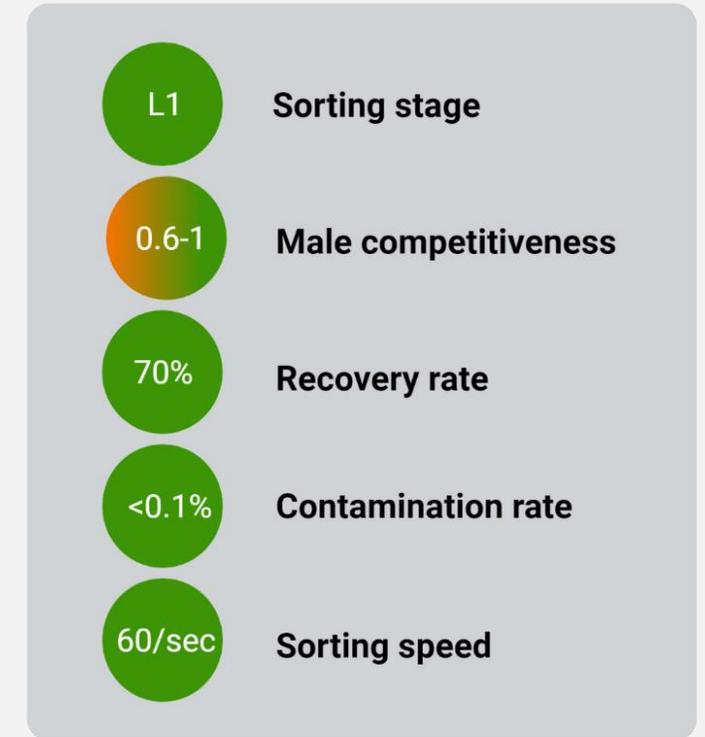
SUMMARY

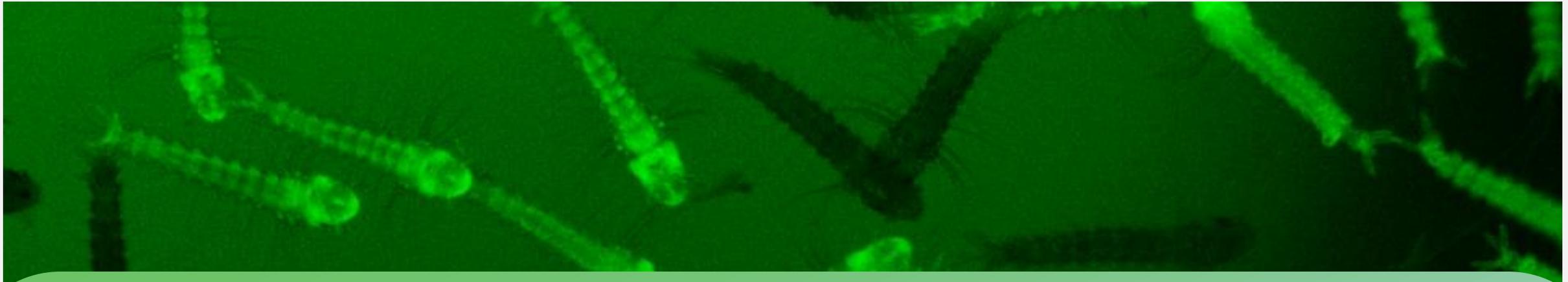
Questions, collaborations:
celia.lutrat@outlook.com

DEVELOPMENT



PERFORMANCE





Développement de méthodes innovantes de sexage pour l'application de la Technique de l'Insecte Stérile chez les moustiques du genre Aedes

Célia Lutrat

celia.lutrat@outlook.com



PERSPECTIVE

Economical interest



SPREADSHEET FOR DESIGNING AEDES MOSQUITO MASS-REARING AND RELEASE FACILITIES

Version 1.0

PERSPECTIVE

Economical interest



SPREADSHEET FOR DESIGNING AEDES MOSQUITO MASS-REARING AND RELEASE FACILITIES

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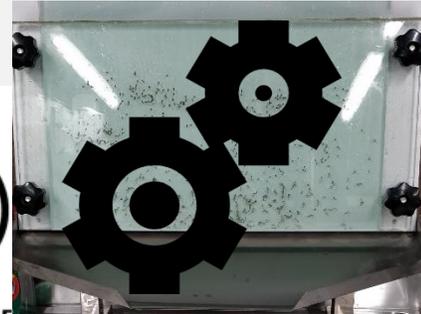
FAO and IAEA 2019
WHO 2018

PERSPECTIVE

Economical interest



Joint FAO/IAEA Programme
Nuclear Techniques in Food and Agriculture



SPREADSHEET FOR DESIGNING AEDES MOSQUITO MASS-REARING AND RELEASE FACILITIES

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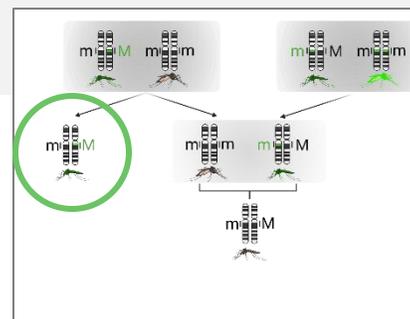
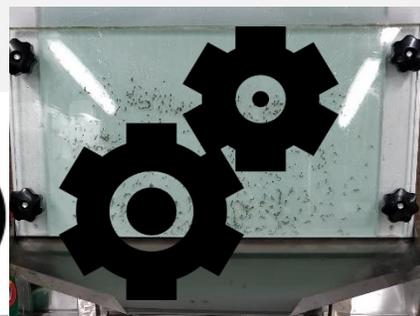
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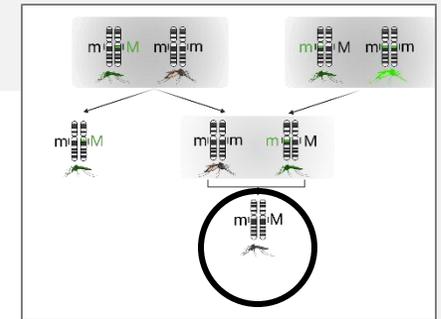
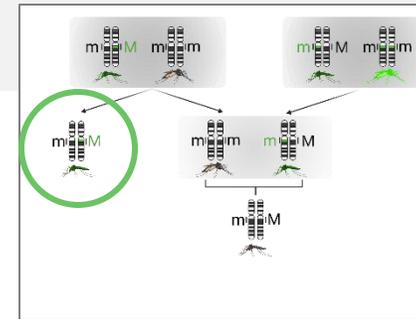
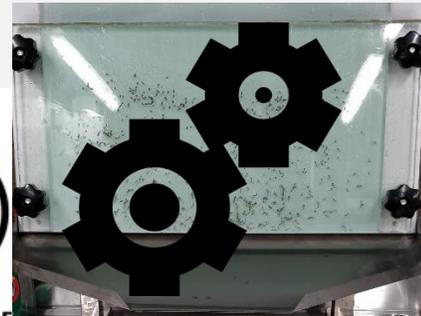
FAO and IAEA 2019
WHO 2018

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Joint FAO/IAEA Programme
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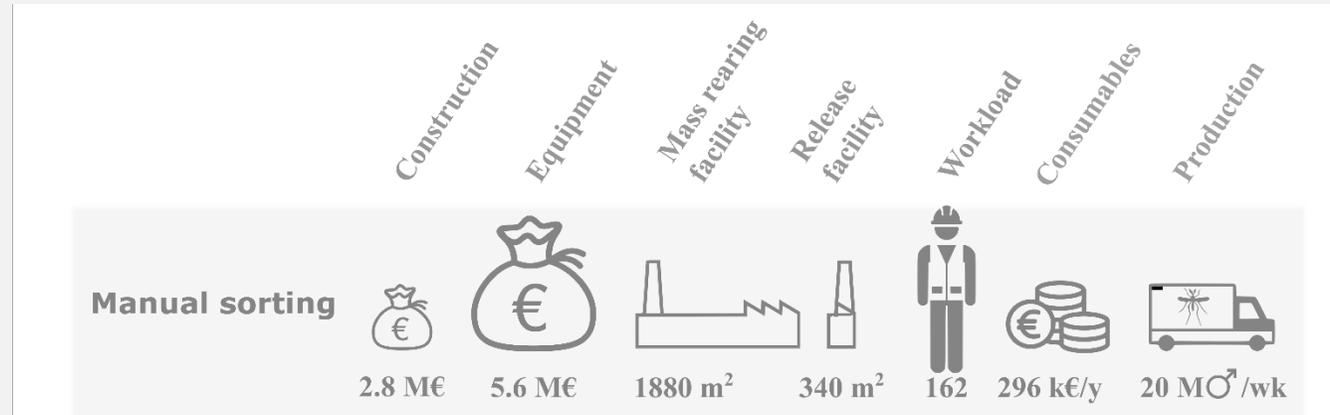
SPREADSHEET FOR DESIGNING AEDES MOSQUITO MASS-REARING AND RELEASE FACILITIES

Version 1.0

FAO and IAEA 2019
WHO 2018

PERSPECTIVE

Economical interest



PERSPECTIVE

Economical interest

	Construction	Equipment	Mass rearing facility	Release facility	Workload	Consumables	Production
Manual sorting	 2.8 M€	 5.6 M€	 1880 m ²	 340 m ²	 162	 296 k€/y	 20 MO [♂] /wk
Automated sorting	 1.6 M€	 2.8 M€	 950 m ²	 340 m ²	 83	 144 k€/y	 20 MO [♂] /wk

PERSPECTIVE

Economical interest

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GSS	 1.2 M€	 2.1 M€	 630 m ²	 340 m ²	 73	 78 k€/y	 20 MO [♂] /wk

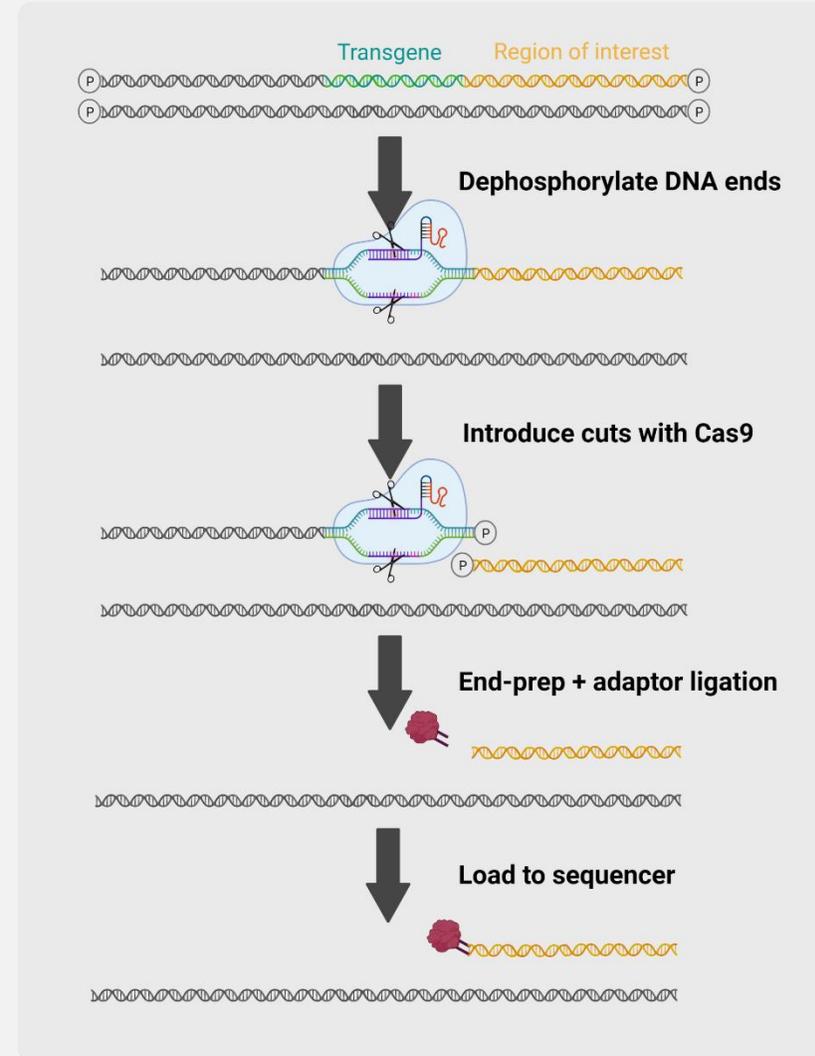
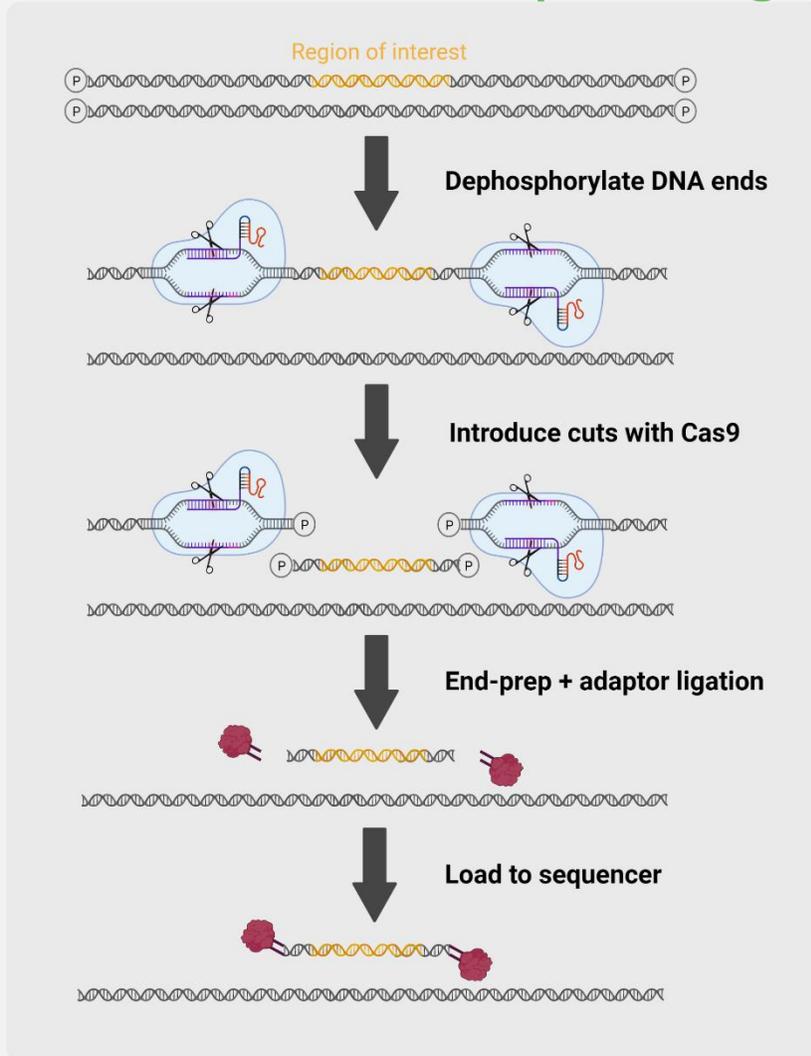
PERSPECTIVE

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SUPPLEMENTARY

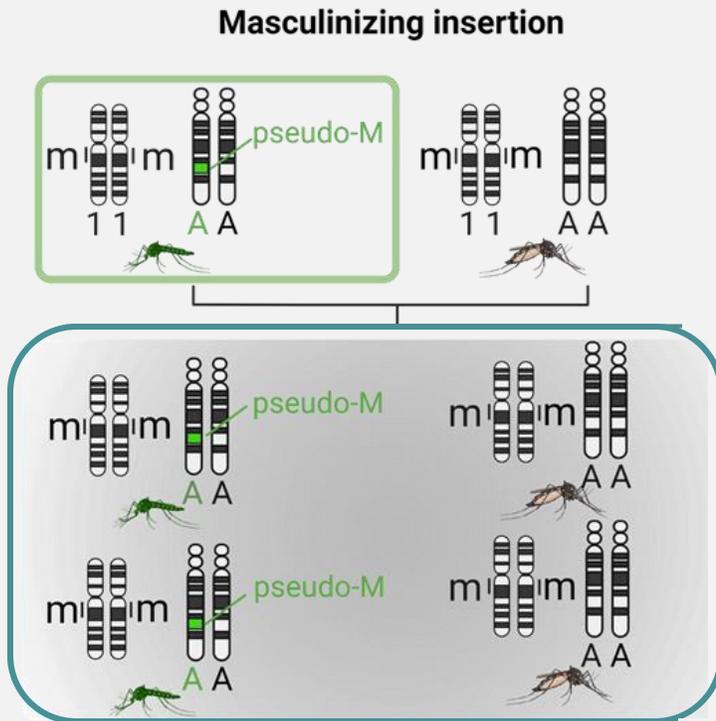
Adaptation of the nCATS sequencing method



Adapted from Gilpatrick et al. 2018

SUPPLEMENTARY

Nix ectopic expression results



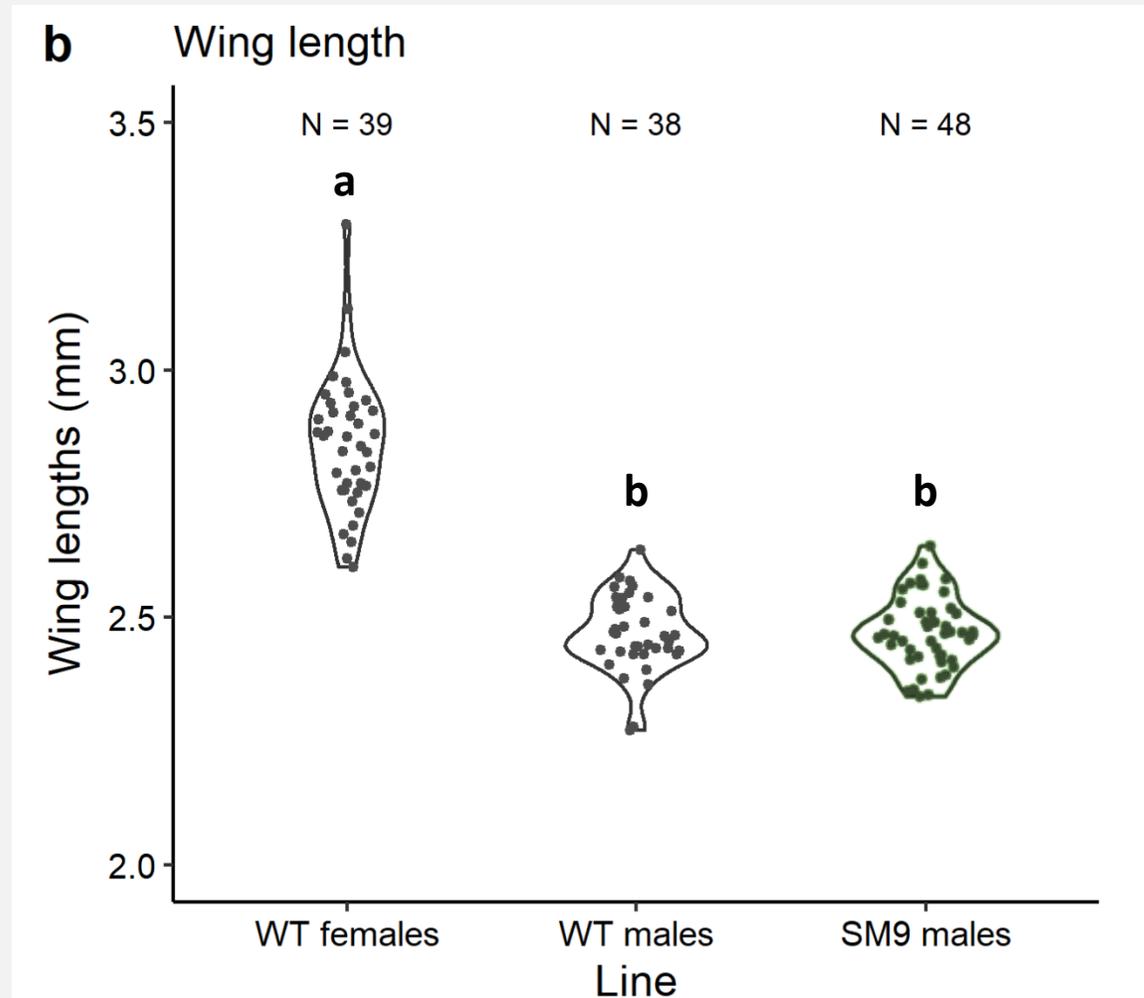
Amplification of the endogenous *Nix* gene

4 out of 13 were pseudo-M linked (GFP)

7 out of 13 were M linked (YFP and/or DsRed)

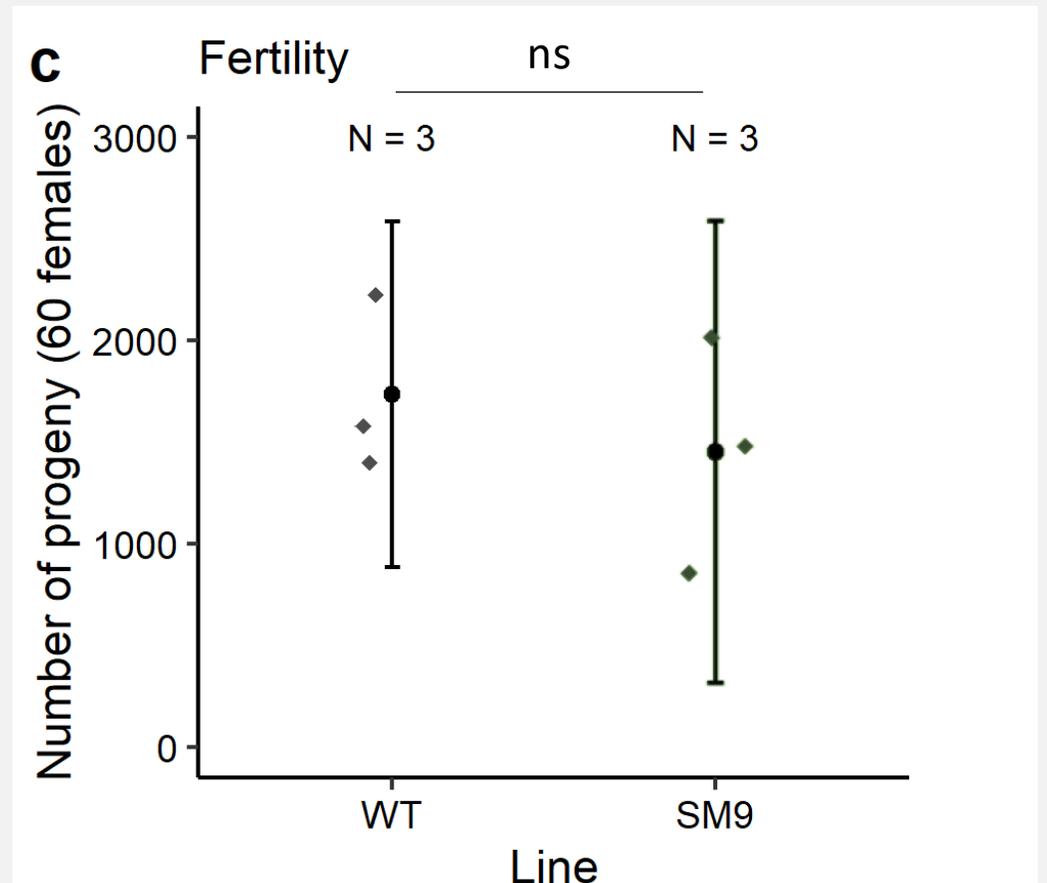
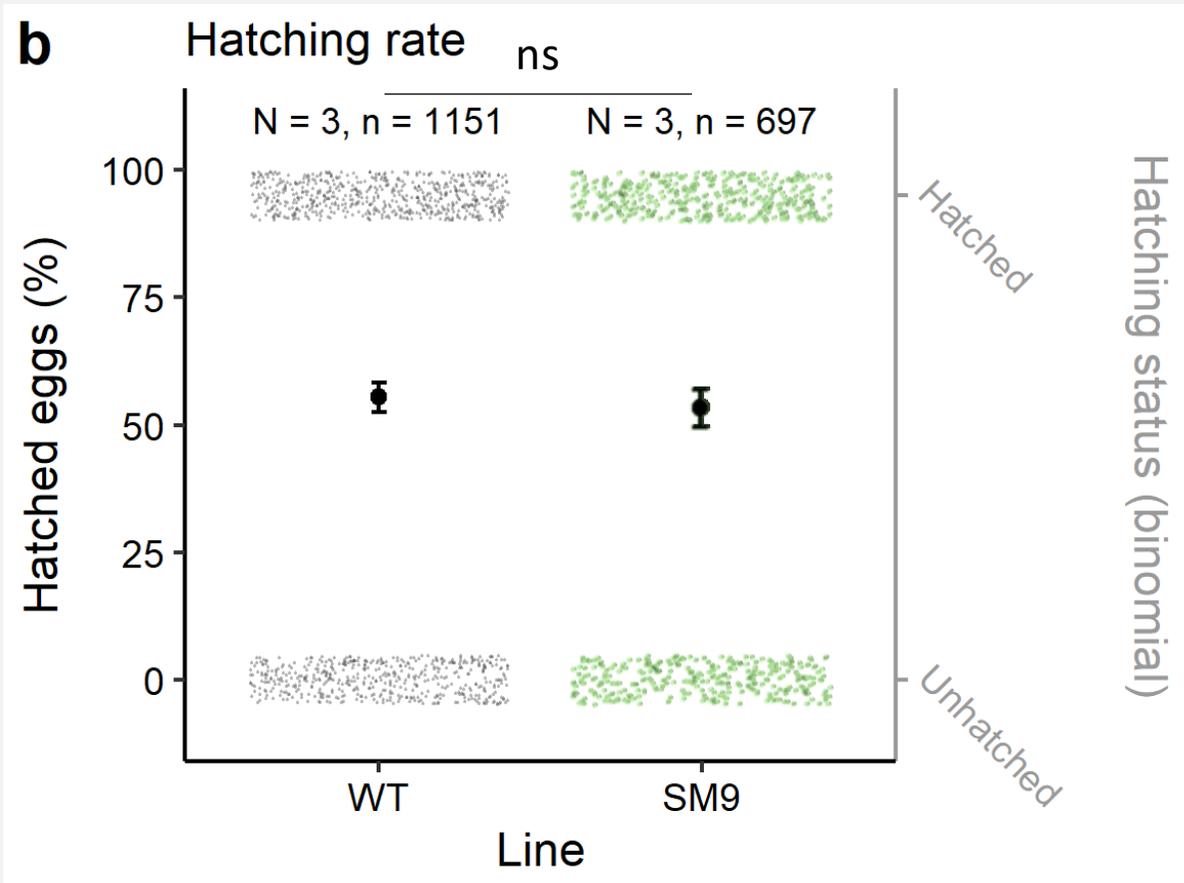
SUPPLEMENTARY

Nix ectopic expression results: size dimorphism



SUPPLEMENTARY

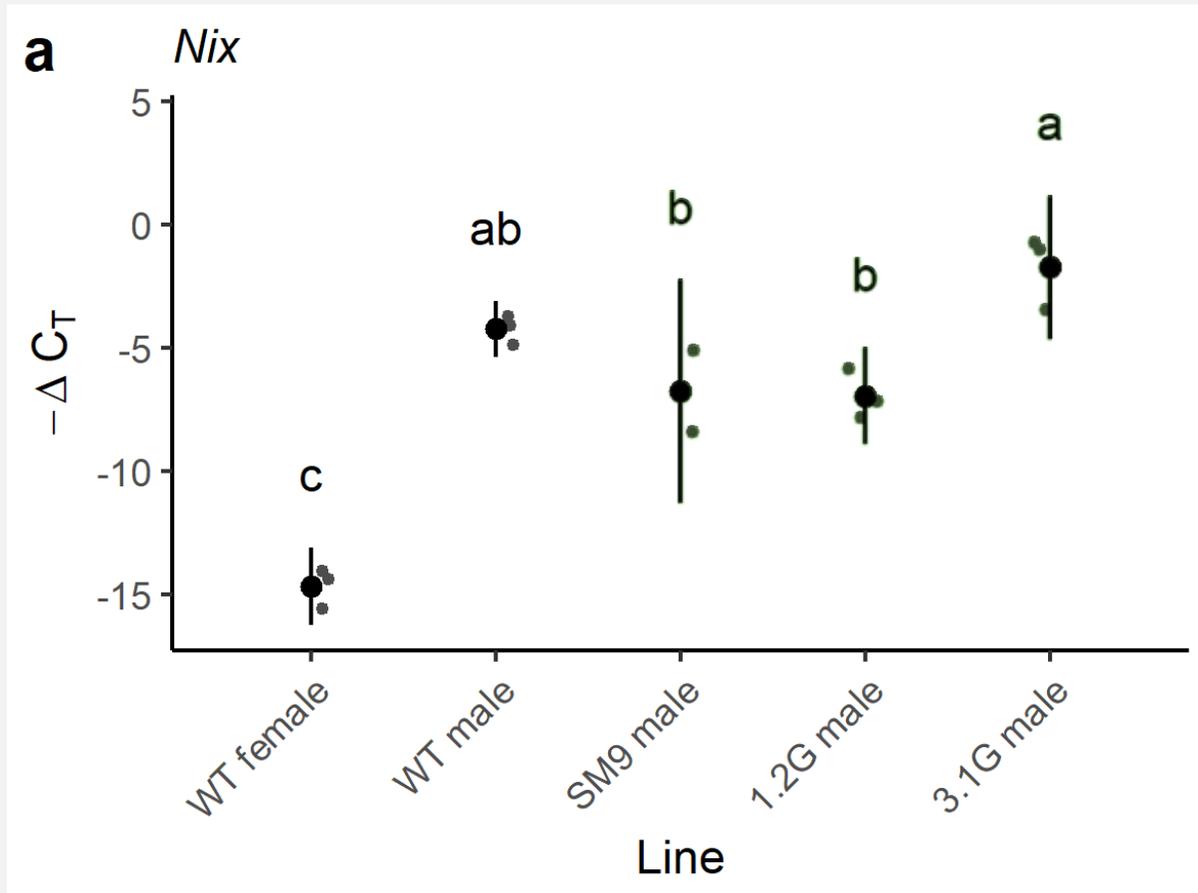
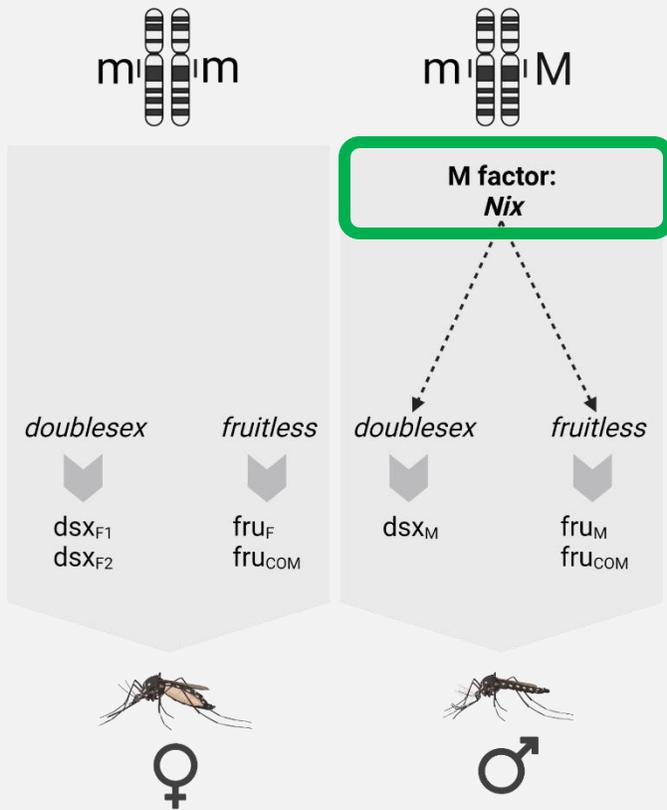
Nix ectopic expression results: fertility



SUPPLEMENTARY

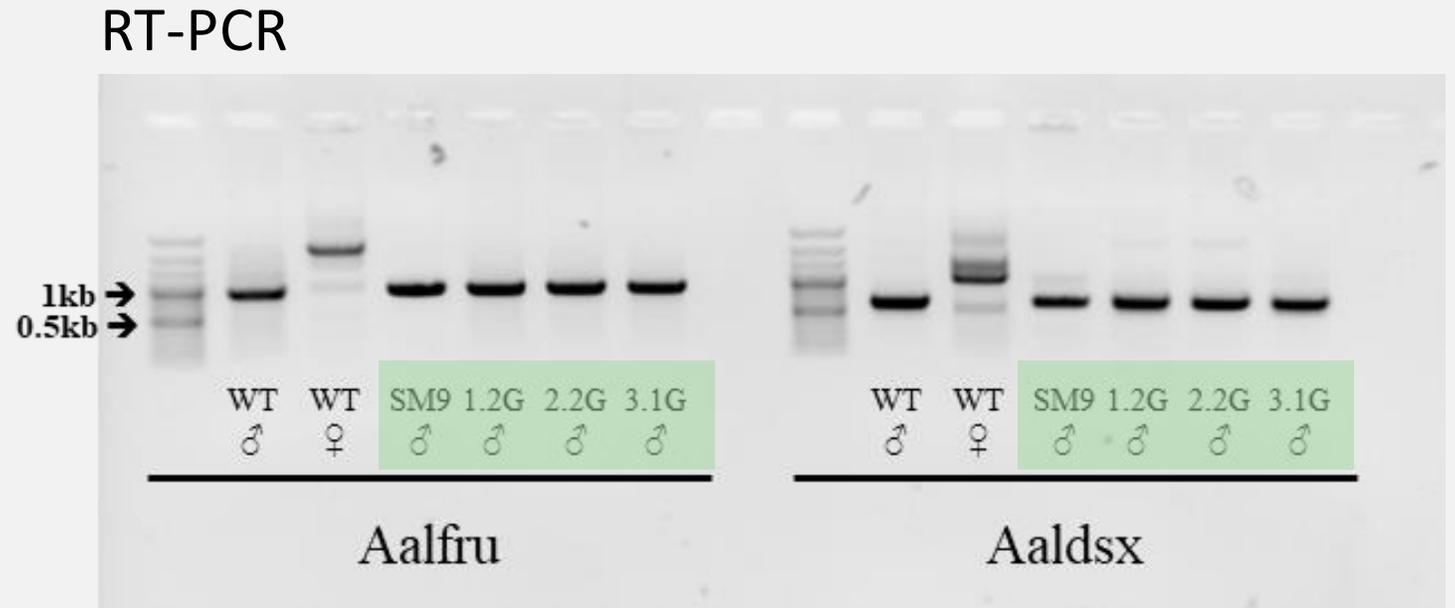
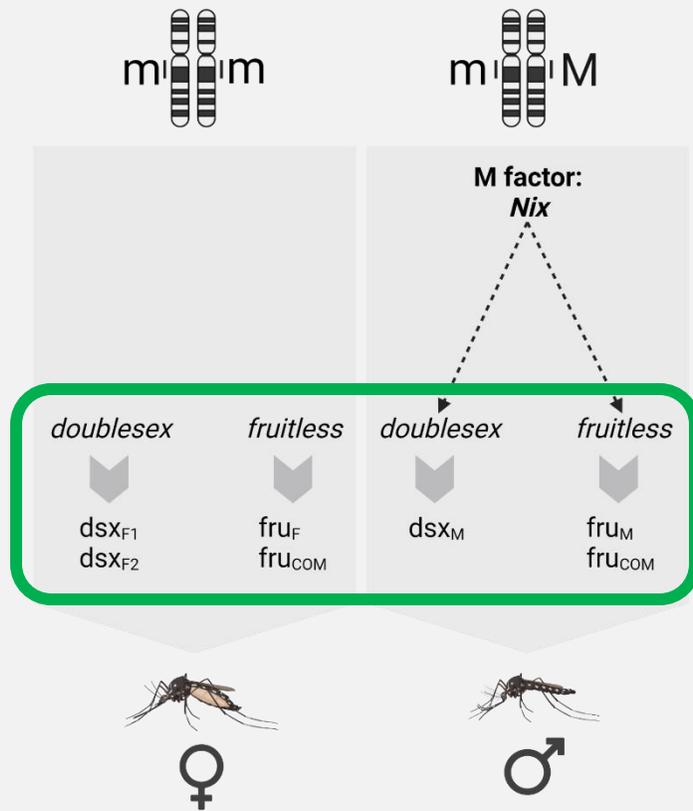
Nix ectopic expression results: expression levels of genes of interest

RT-qPCR



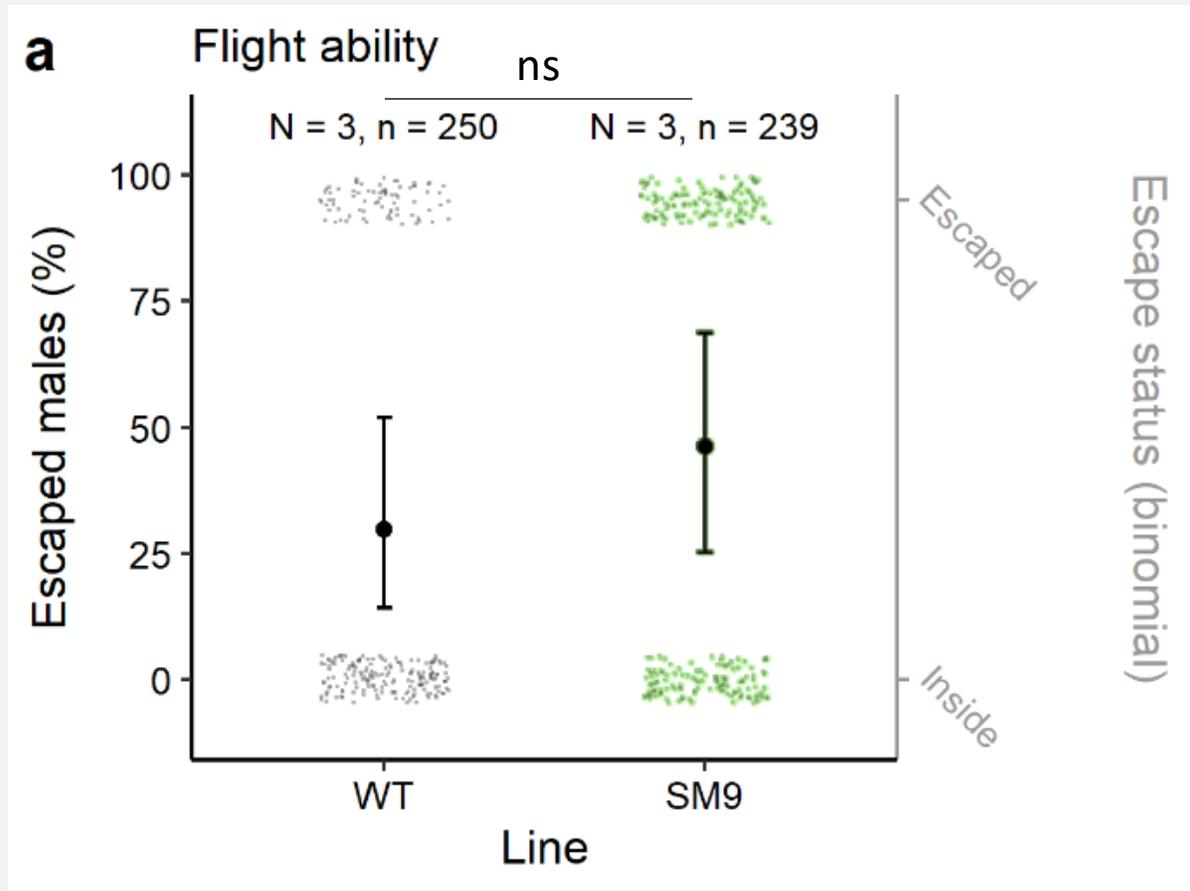
SUPPLEMENTARY

Nix ectopic expression results: expression levels of genes of interest



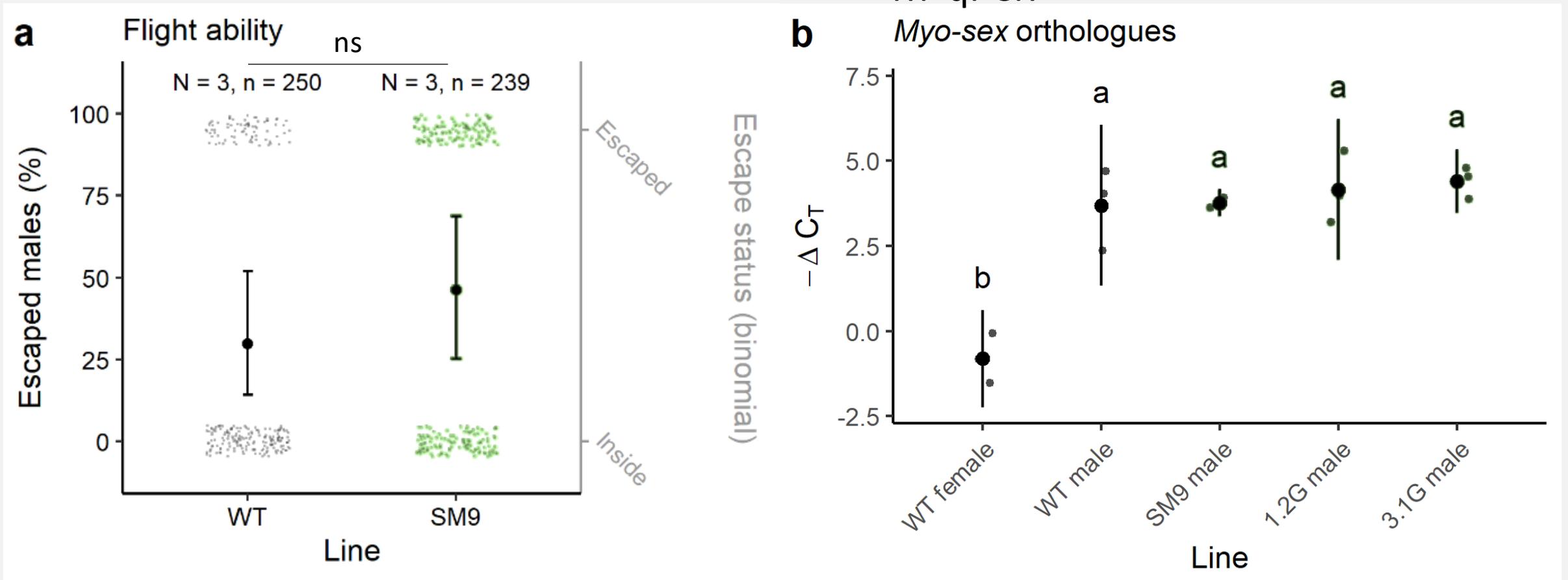
SUPPLEMENTARY

Nix ectopic expression results: expression levels of genes of interest



SUPPLEMENTARY

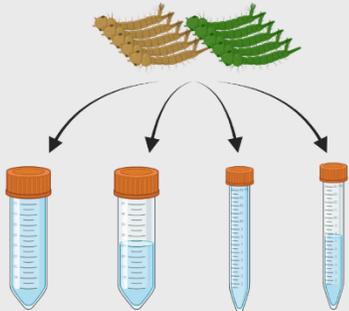
Nix ectopic expression results: expression levels of genes of interest



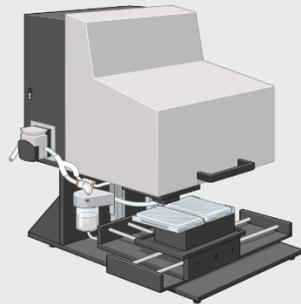
SUPPLEMENTARY

COPAS sorting speed

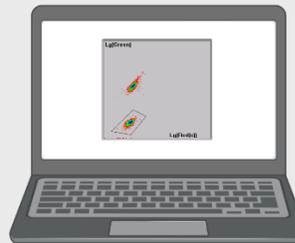
Prepare samples of a defined number of larvae at different concentrations



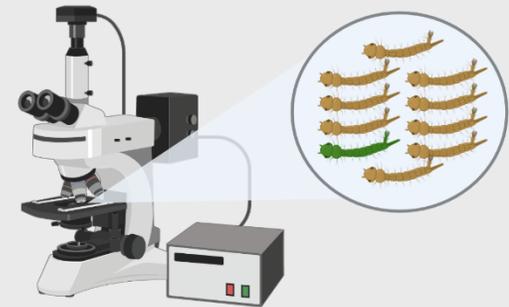
Sort by COPAS



Determine recovery rate

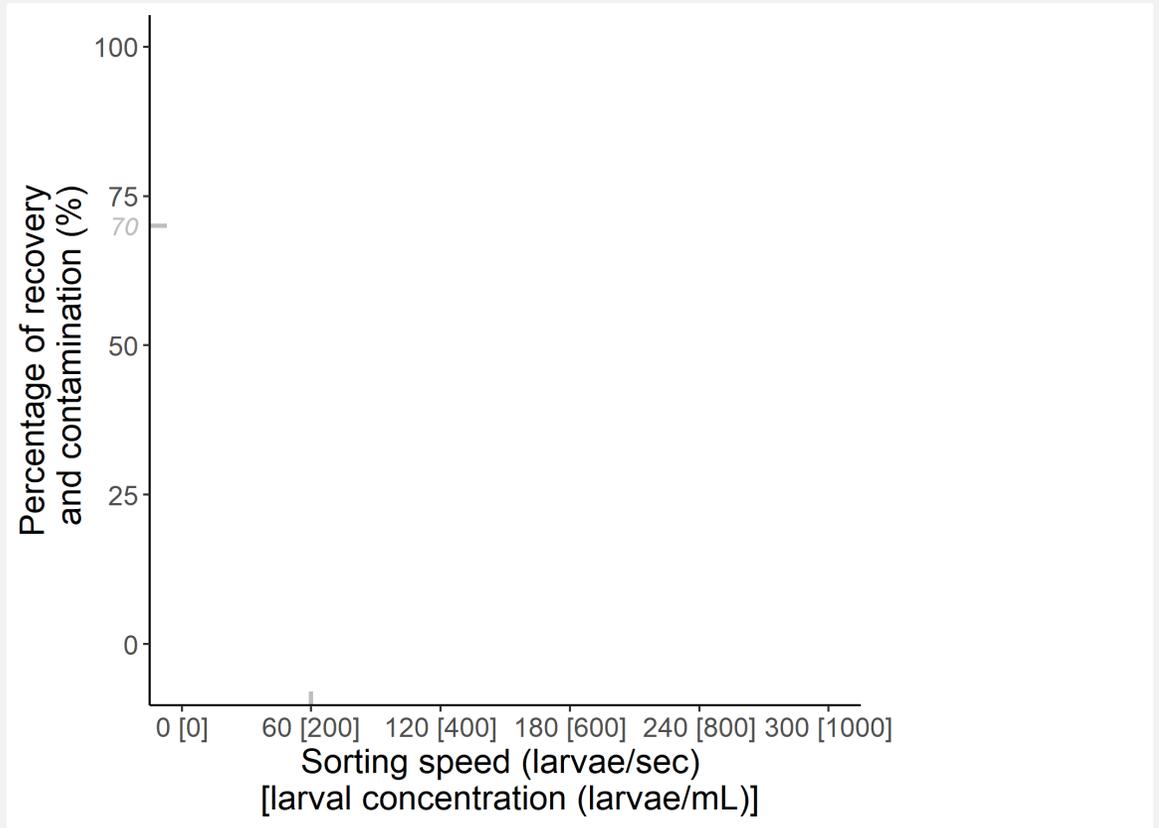
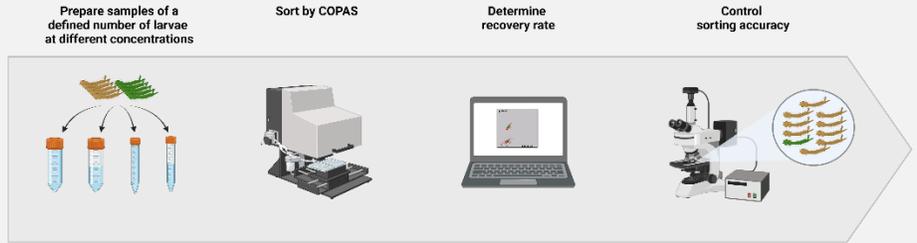


Control sorting accuracy



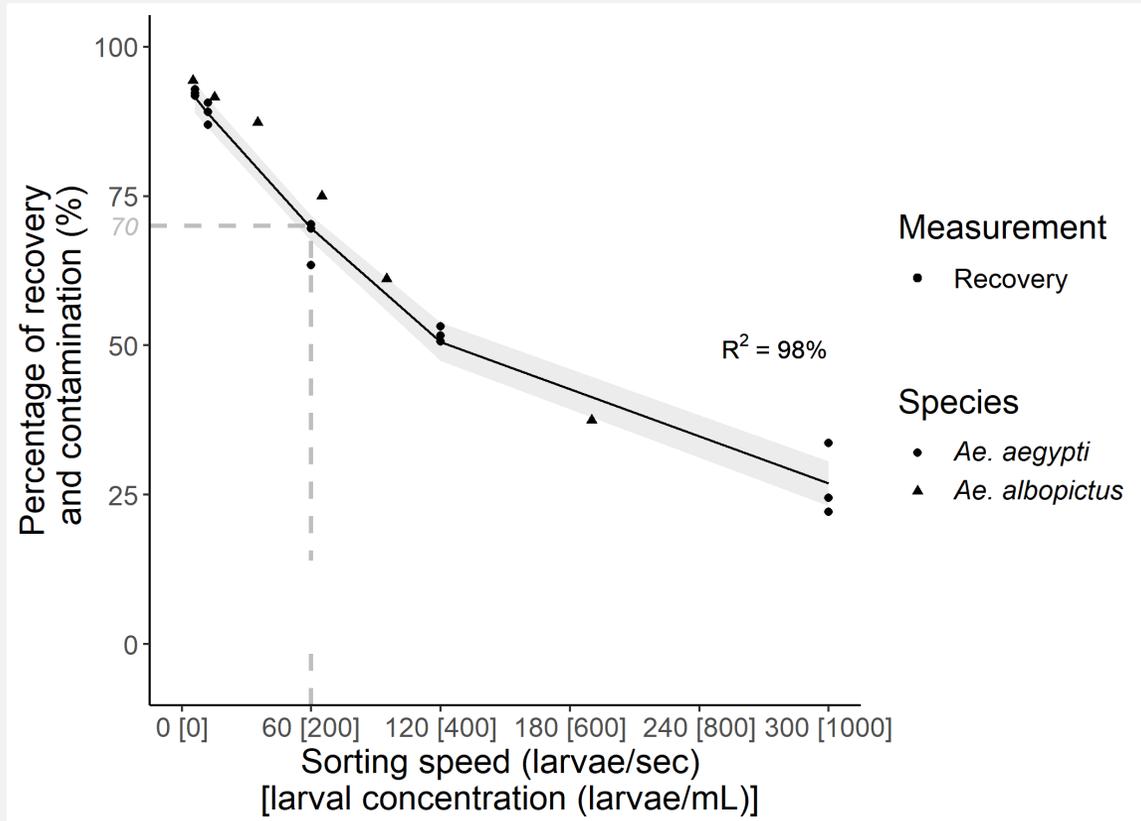
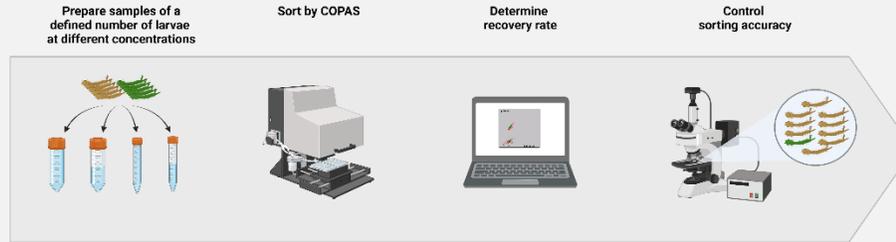
SUPPLEMENTARY

COPAS sorting speed



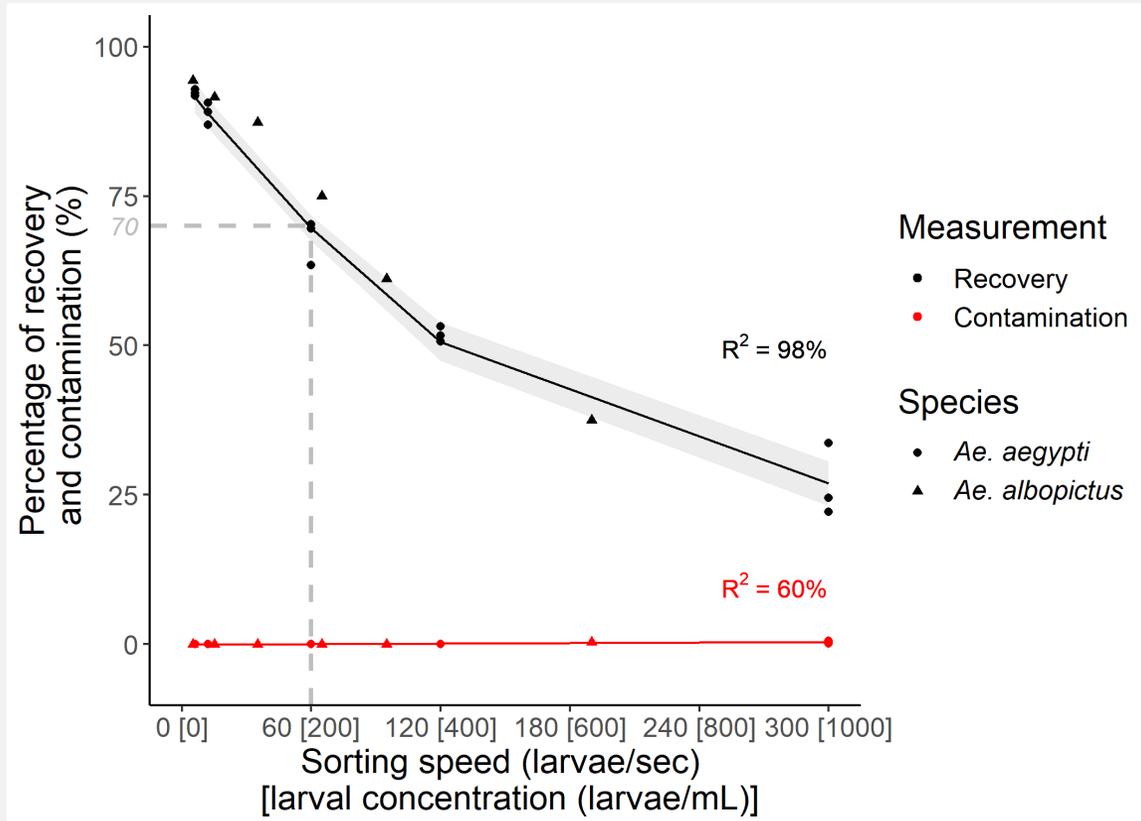
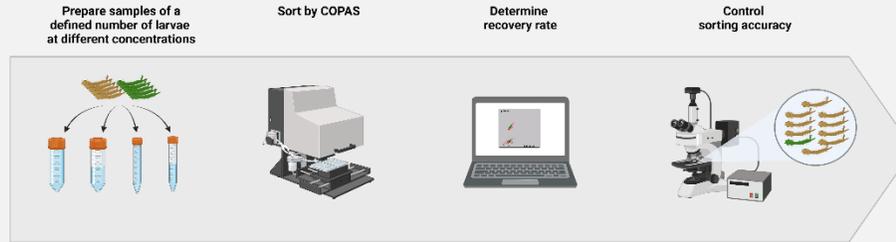
SUPPLEMENTARY

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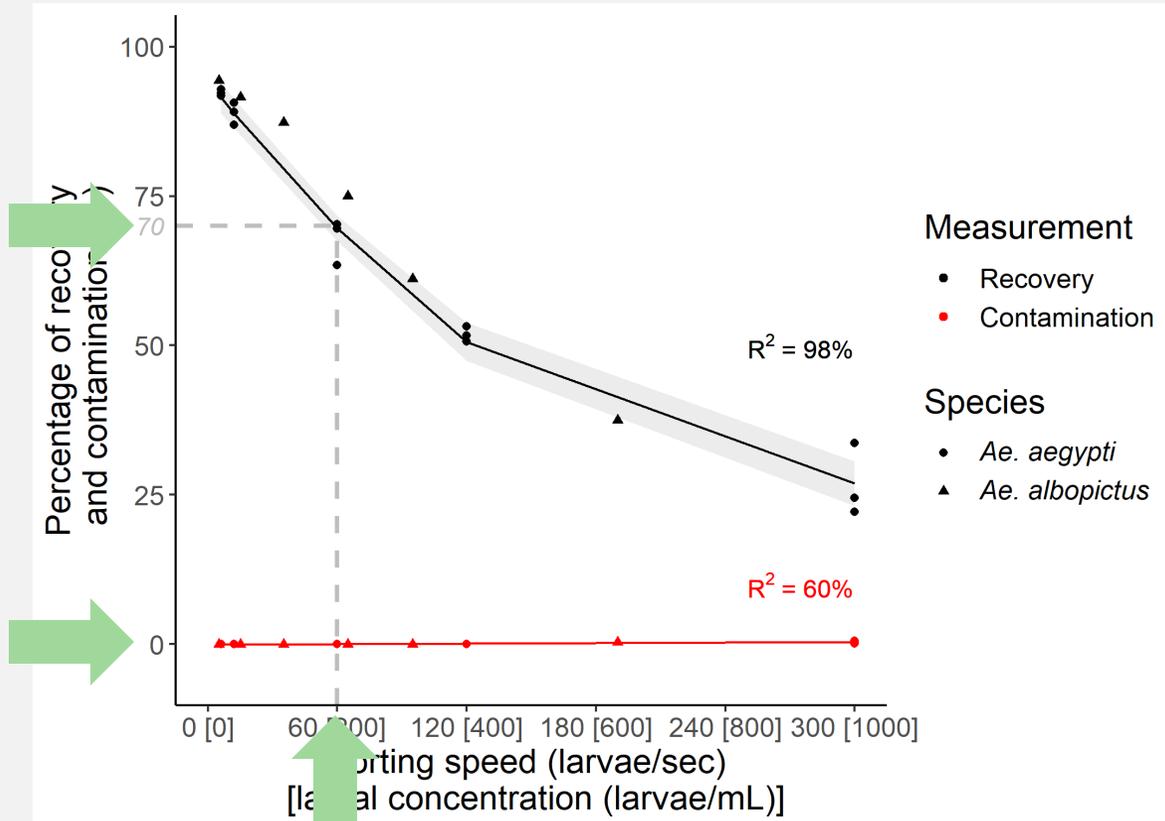
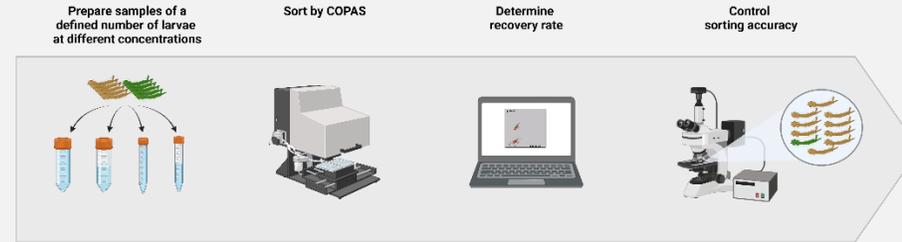
SUPPLEMENTARY

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SUPPLEMENTARY

COPAS sorting speed

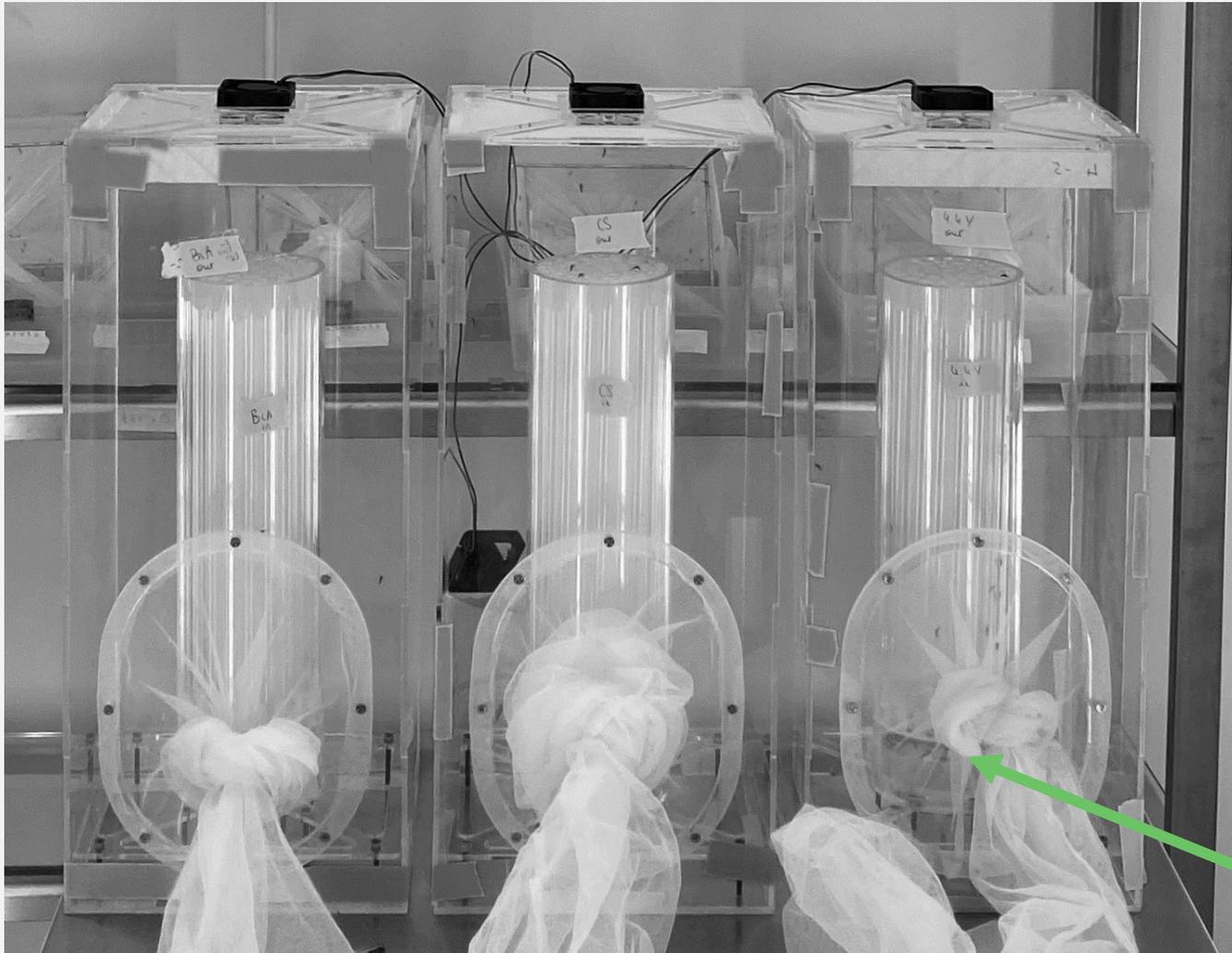


At 60 larvae / sec:
70% recovery
0% contamination

1 million males:
<15 hours

SUPPLEMENTARY

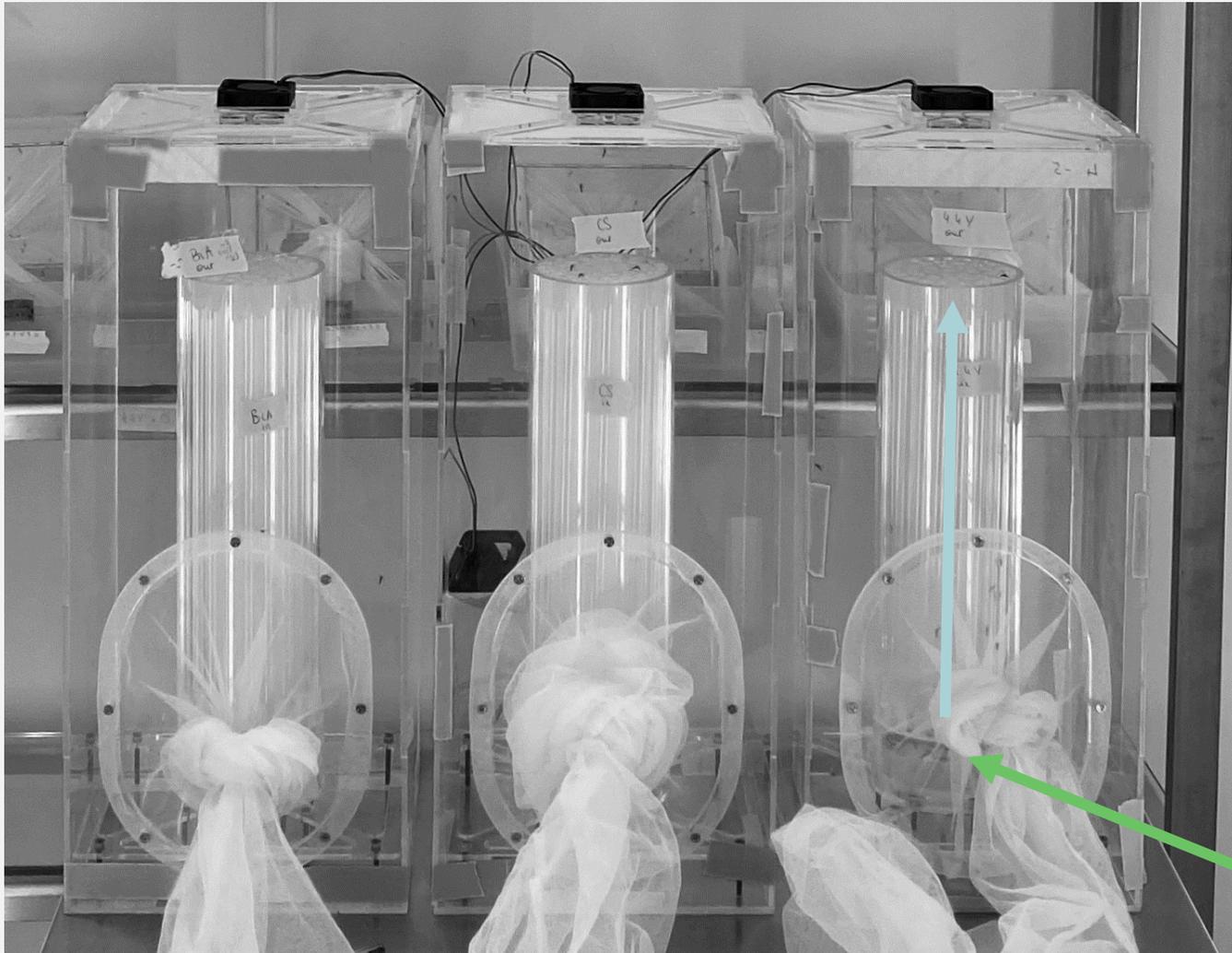
Male flight ability



Insert mosquitoes
in the bottom chamber

SUPPLEMENTARY

Male flight ability

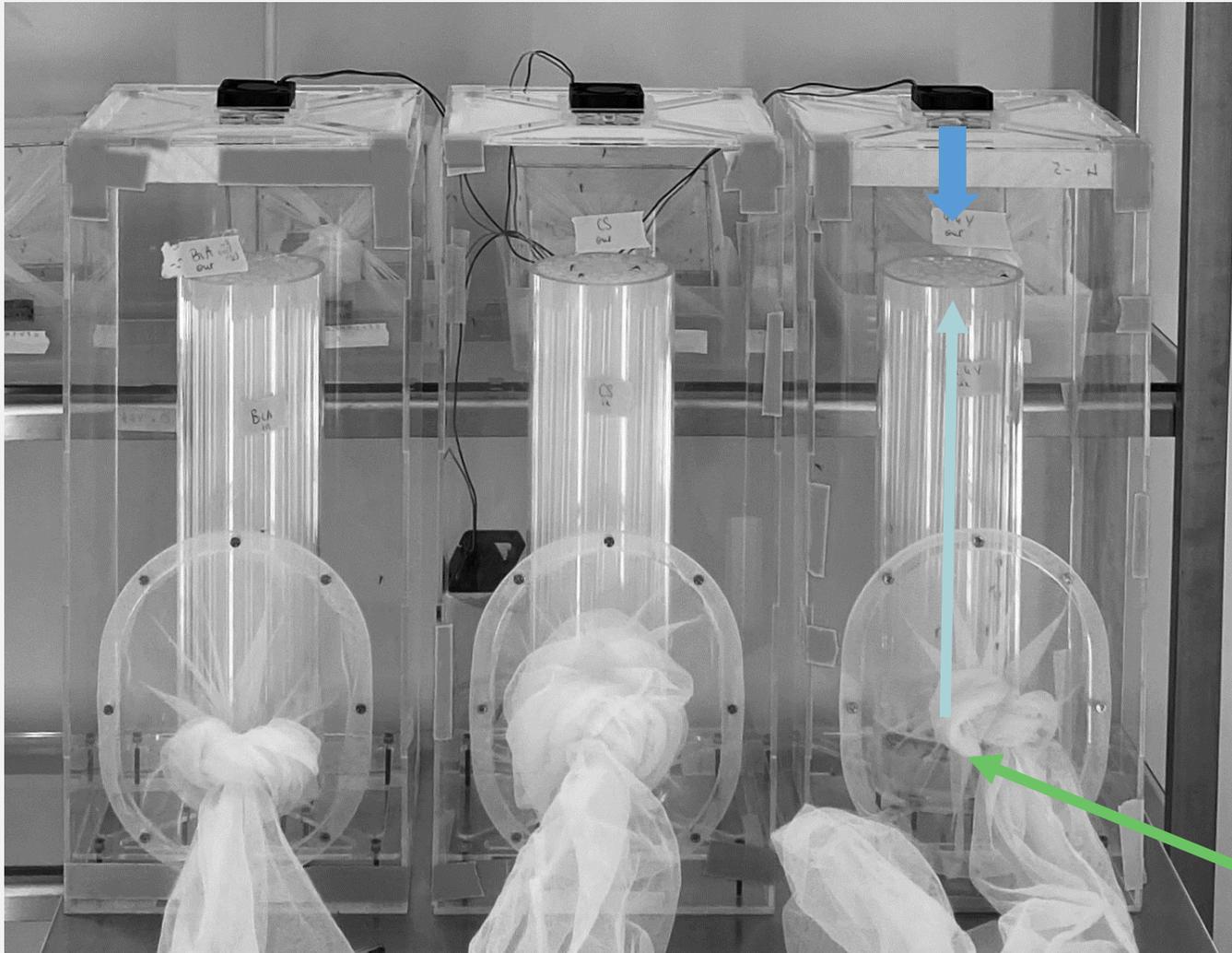


Let them fly up the tubes

Insert mosquitoes
in the bottom chamber

SUPPLEMENTARY

Male flight ability



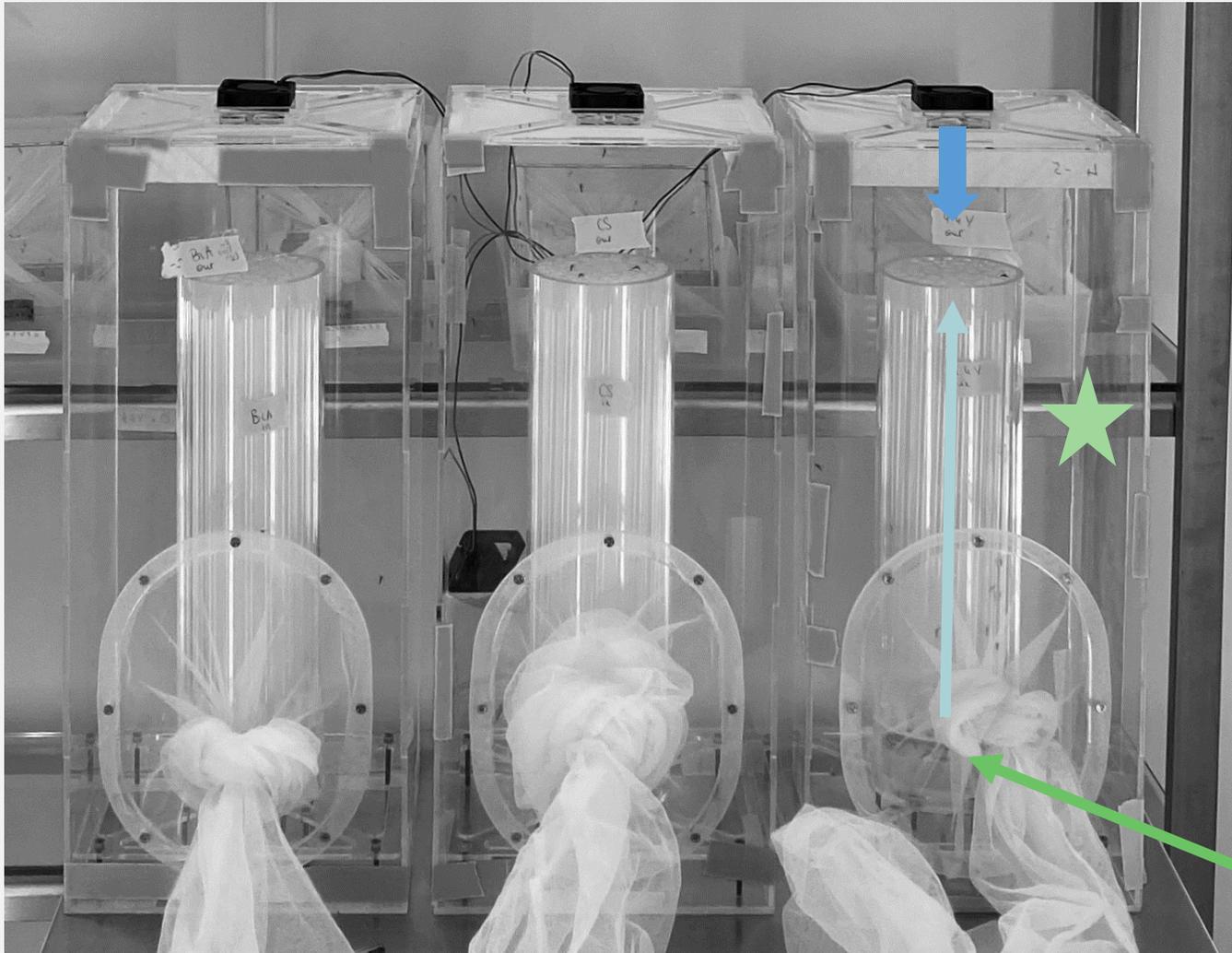
The fan blows air in the opposite direction, making it harder

Let them fly up the tubes

Insert mosquitoes in the bottom chamber

SUPPLEMENTARY

Male flight ability



The fan blows air in the opposite direction, making it harder

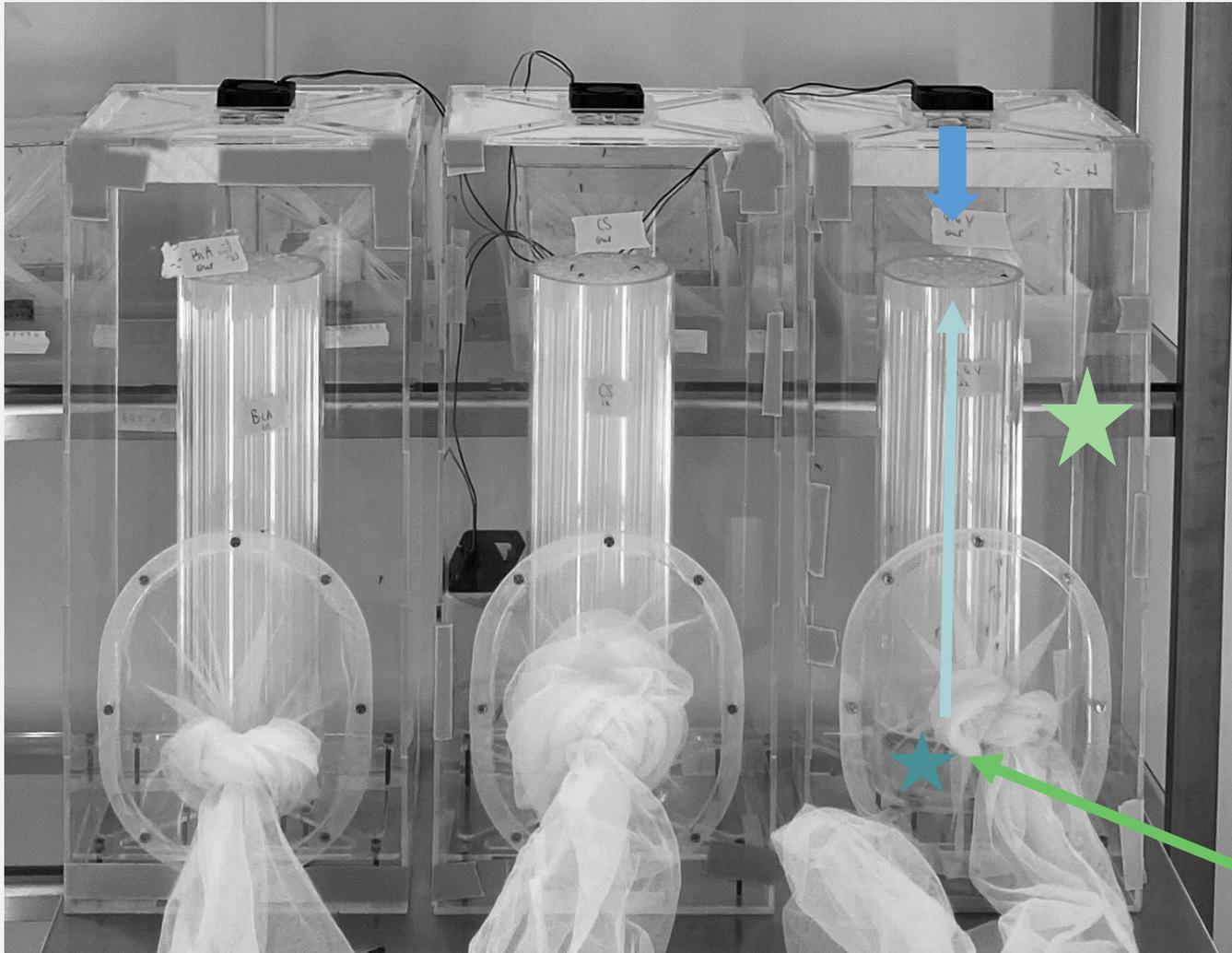
The ones that managed to make it to the bigger chamber: succeeded

Let them fly up the tubes

Insert mosquitoes in the bottom chamber

SUPPLEMENTARY

Male flight ability



The fan blows air in the opposite direction, making it harder

The ones that managed to make it to the bigger chamber: succeeded

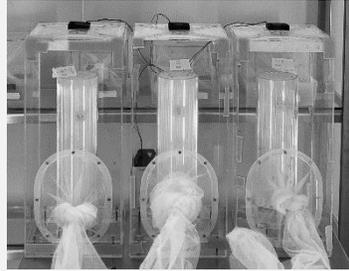
Let them fly up the tubes

The ones that stayed in the tube or in the bottom chamber: failed

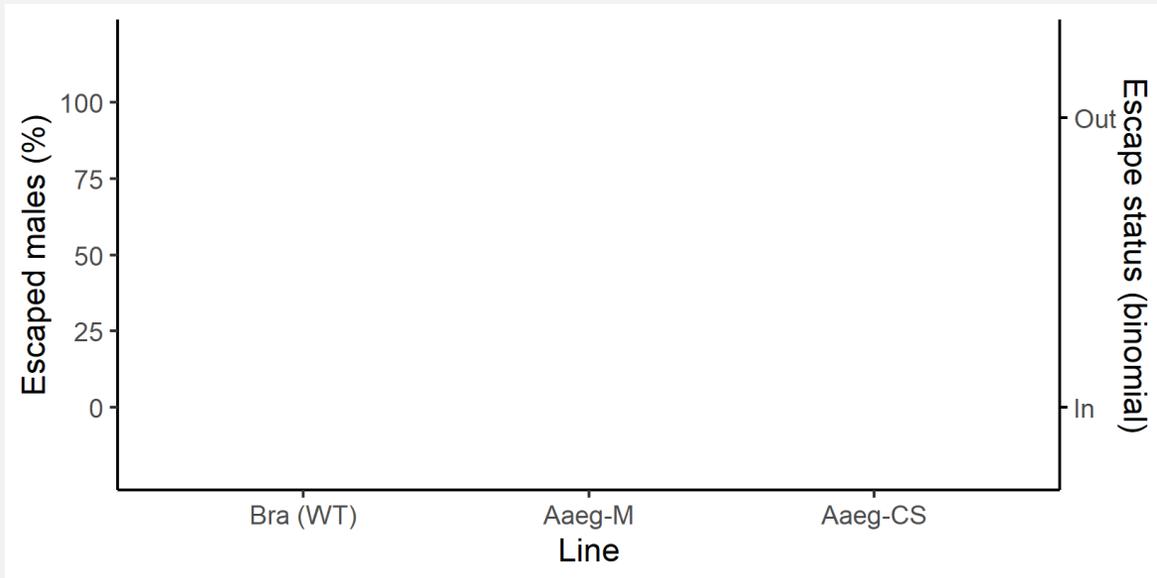
Insert mosquitoes in the bottom chamber

SUPPLEMENTARY

Male flight ability - GSS

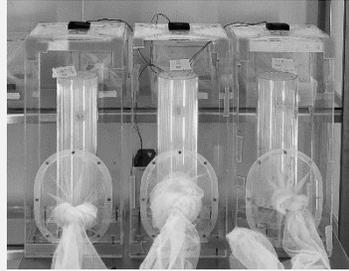


Aedes aegypti

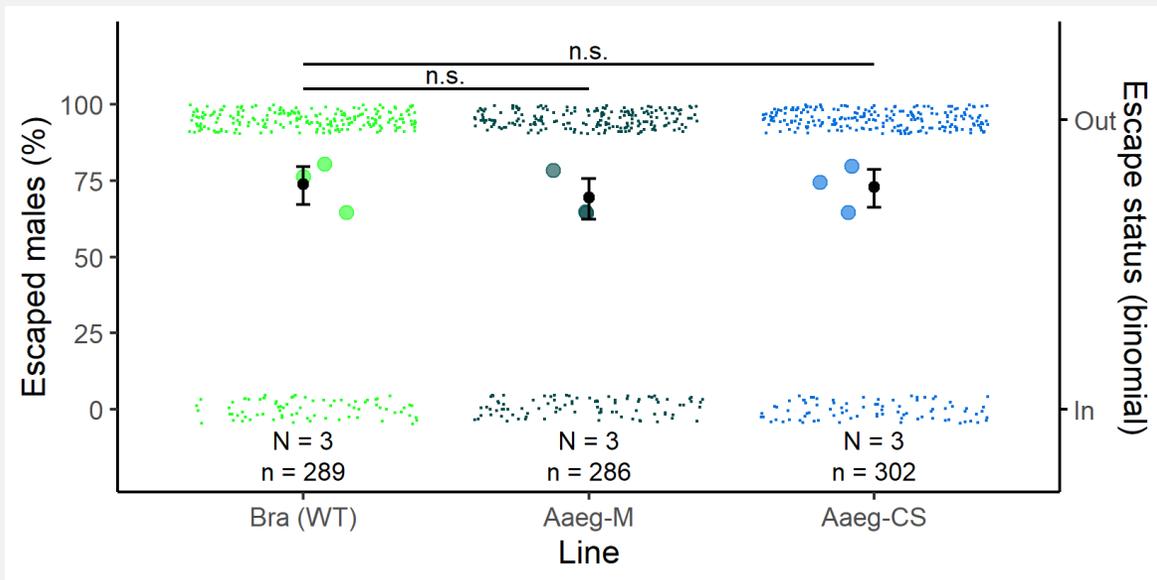


SUPPLEMENTARY

Male flight ability - GSS

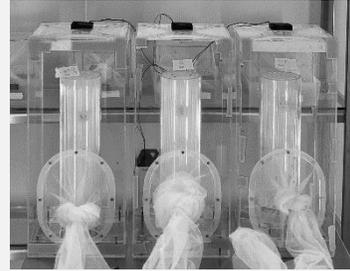


Aedes aegypti

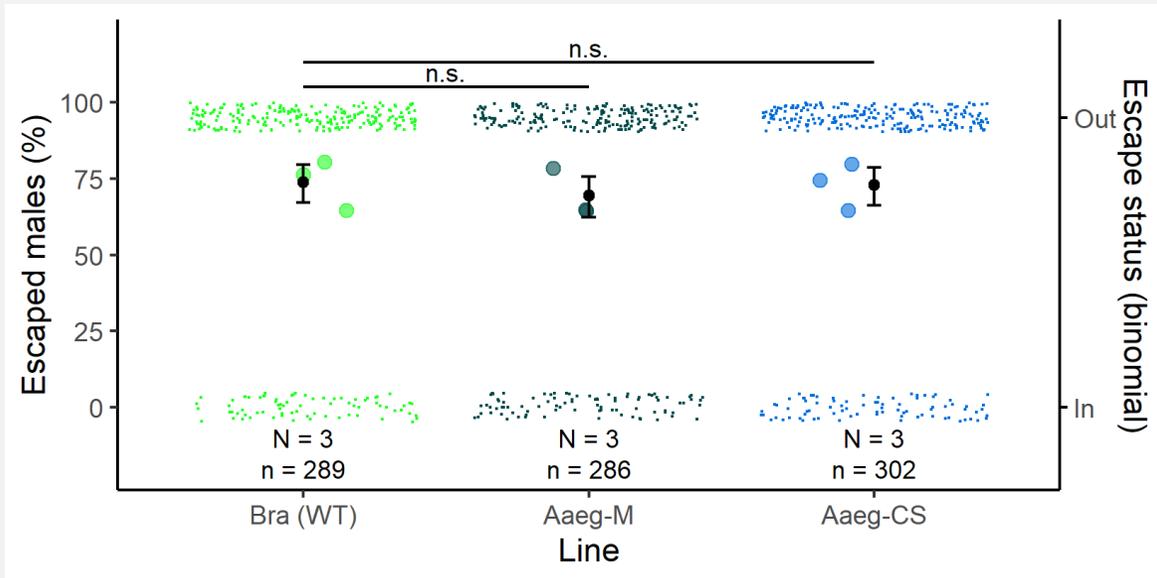


SUPPLEMENTARY

Male flight ability - GSS



Aedes aegypti



Aedes albopictus

