

A Song of Sap and Blood

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Diego Santos-Garcia, Ph.D.

Laboratory of Biometry and Evolutionary Biology
University Lyon 1 - UMR CNRS 5558

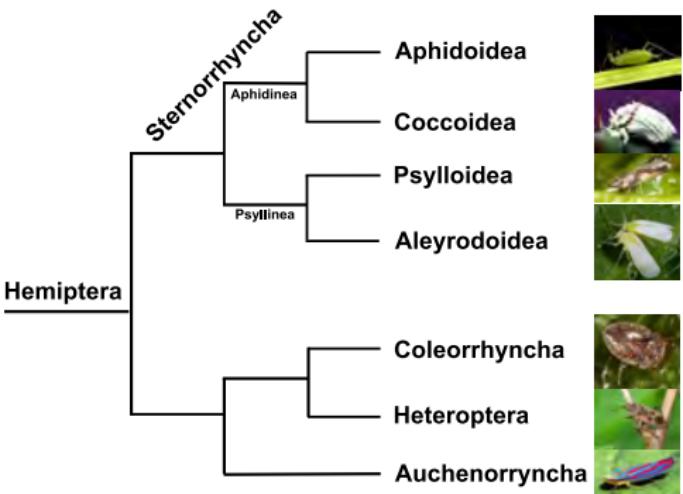
February 15th, 2022

Portiera gets wild: Genome instability provides insights into the evolution of both whiteflies and their endosymbionts

Santos-Garcia et al. 2020. *Genome Biol. Evol.* 12, 2107–2124.



Hemiptera order



- Hemiptera order is the **largest group of Hemimetabolous insects** (without metamorphosis).
- Hemiptera is divided into four suborders (Cryan and Urban, 2012).
- They are **adapted to fluid diets**, mainly plant sap: specialized mouthparts and digestive system.
- **Whiteflies: Aleyrodoidea superfamily** > Aleyrodidae family.

- Aleyrodoidea has **two recognized subfamilies**: the **Aleyrodinae** (ca. 140 genera) and the **Aleurodicinae** (ca. 20).
- Subfamilies **origin** was traced to the **Lower Cretaceous** (100-145 Mya).



Sternorrhyncha:Aleyrodoidea

- Paleotropical origin: "the aphids of the tropical regions"
 - Adults are tiny (1-4 mm) and usually covered by wax.
 - Four nymphal stages (instar):
 - 1st is the only mobile form - leaf veins hunter.
 - 2nd and 3rd are feeding stages (size increase).
 - 4th or "**red eye pupa**" is a quiescent stage.

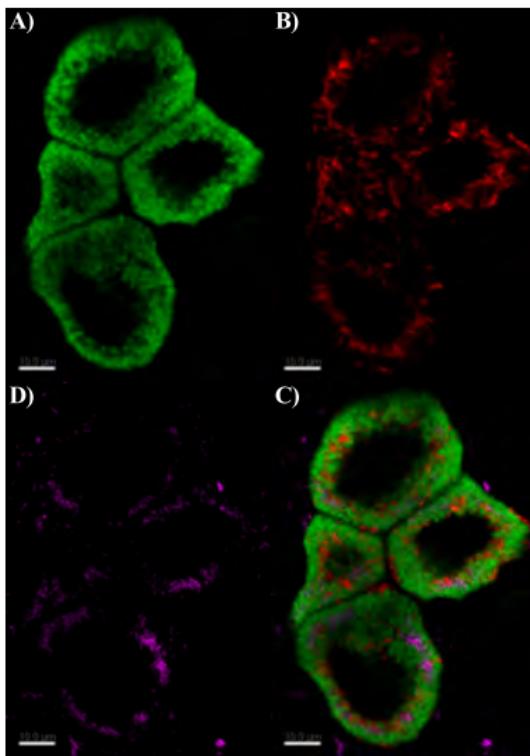


Sunendra Datta blog at <http://iucanr.edu/bloes>. 4 th instar photo by Paul de Roma.





Whiteflies endosymbionts

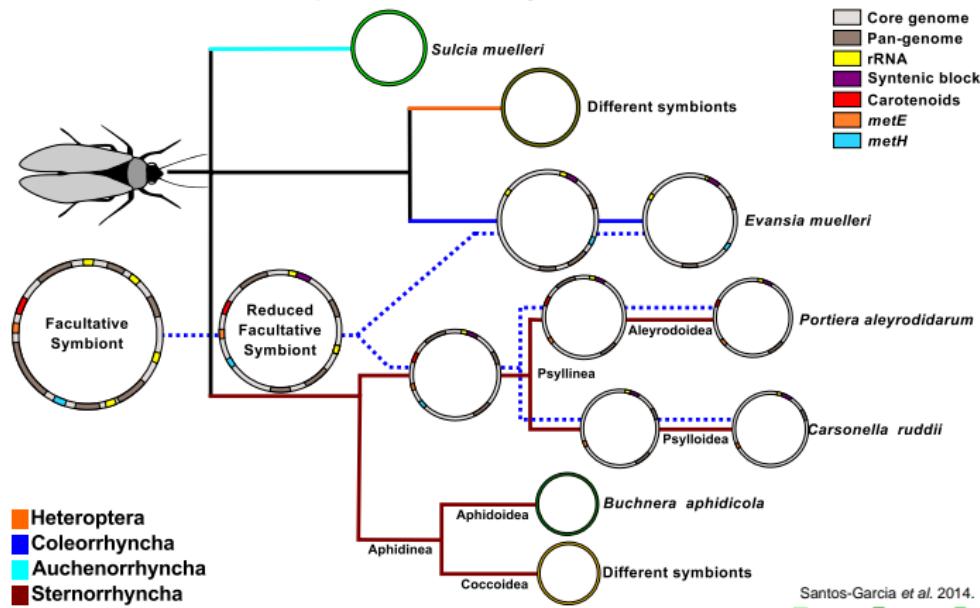


- Whiteflies always present the **P-endosymbiont *Candidatus Portiera aleyrodidarum*** (green).
- Portiera* is devoted to the **production of essentials amino acids and carotenoids**.
- A **co-primary, *Hamiltonella*** (red) or ***Arsenophonus***, supplements **B-vitamins** to their whitefly host (among other possible benefits).
- Whiteflies may also harbor other secondary endosymbionts: ***Wolbachia***, ***Rickettsia***, and ***Cardinium*** (purple).



Portiera, an ancient endosymbiont

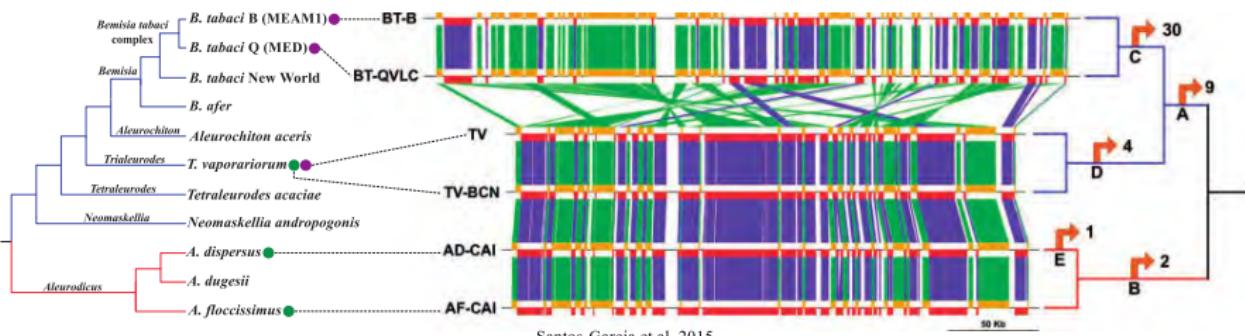
- Portiera has co-diverged with whiteflies since their origin (> 150 Myr).
 - Already a reduced endosymbiont with basic DNA replication, recombination, and repair machinery.





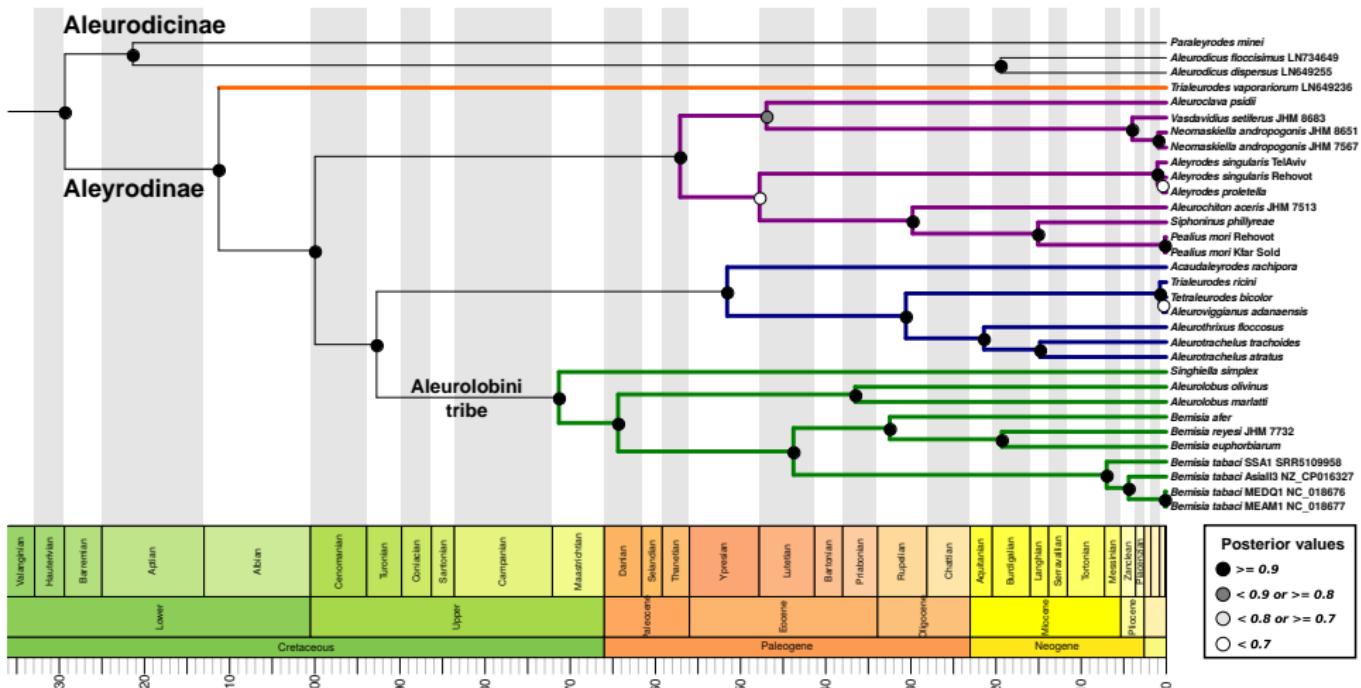
Portiera, an ancient endosymbiont

- Portiera has co-diverged with whiteflies since their origin (> 150 Myr).
 - Already a reduced endosymbiont with basic DNA replication, recombination, and repair machinery.
- Portiera genomes from most whiteflies are syntenic except in *Bemisia tabaci* → genomic instability.
- Portiera from *B. tabaci* has lost the polymerase proofreading subunit DnaQ → cause of instability? (Sloan and Moran, 2013).





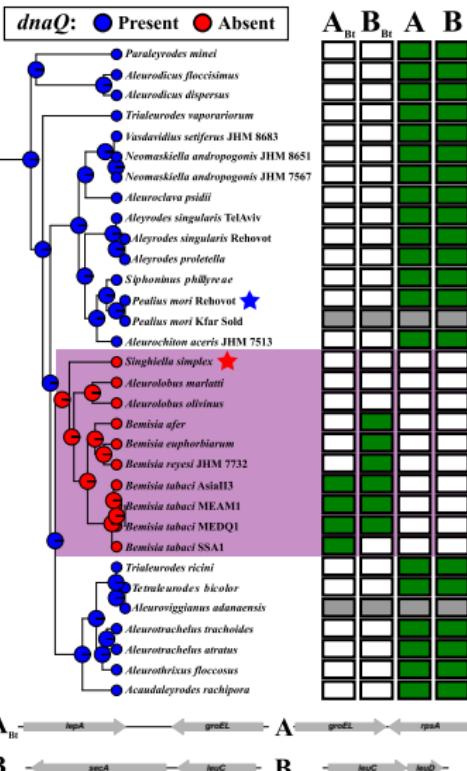
Portiera phylogeny: 16S, 23S, rpoD, dnaK, and groEL



● 27 whitefly species (19 genera) collected from Israel, La Reunion, Canary Islands, the NHM of London collection, and public databases.



Tracking *Portiera* genome instability

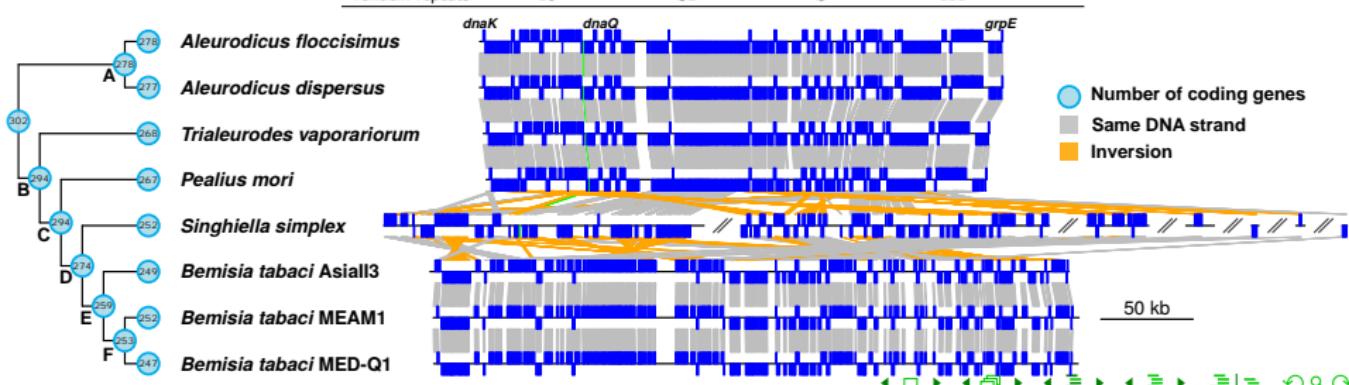


- Screening for the presence of *dnaQ* and different gene order configurations (ancestral or derived).
- dnaQ* was lost in the *Singhiella*, *Aleurolobus*, and *Bemisia* (**Aleurolobini tribe**) monophyletic clade MRCA (~70 Mya).
 - Bemisia* other than *tabaci* suffered re-arrangements compared to *tabaci*.
 - Singhiella* and *Aleurolobus* do not presented neither the ancestral or *B. tabaci* order.
- To confirm the screening results: sequencing *Portiera* from the **most basal species** in the *Bemisia* clade - the fig whitefly ***Singhiella simplex***.
- Two *Portiera* with one shot: the sample contained some individuals of *Pealius mori*.

Two new *Portiera* genomes: SiSi and PeMo

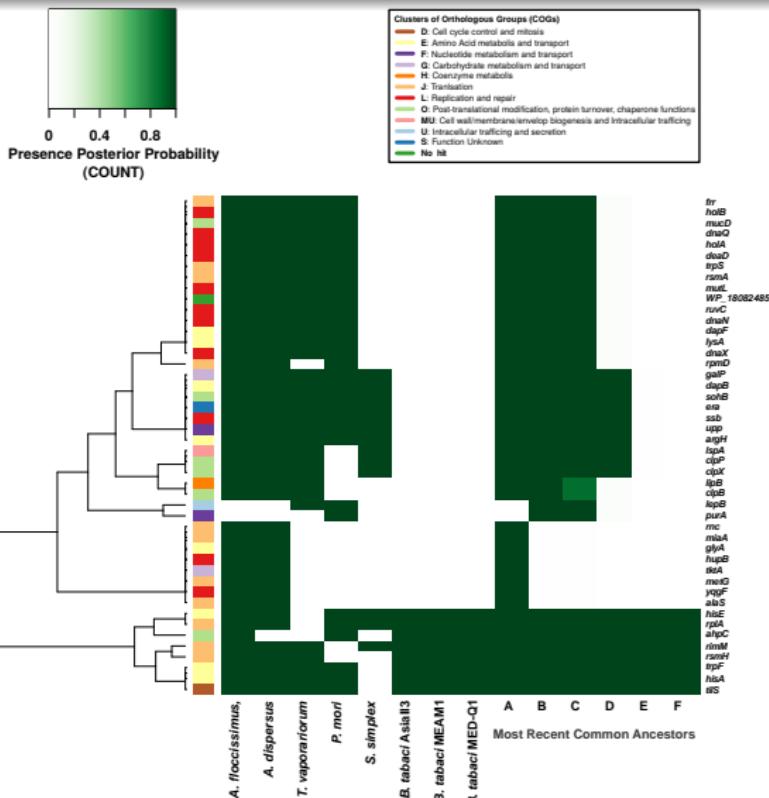


	<i>Portiera</i> TeVa	<i>Portiera</i> PeMo	<i>Portiera</i> SiSi	<i>Portiera</i> BeTa
Host	<i>T. vaporariorum</i>	<i>P. mori</i>	<i>S. simplex</i>	<i>B. tabaci</i> MED-Q1
Genome size (bp)	280822	277700	411975	357472
Contigs	1	1	9	1
Genes*	307	308	300	284
CDS	268	266	252	247
Pseudogenes (CDS)	1	3	11	7
Intergenic avg. length	62.79	51.81	715.44	524.99
Coding Density (%)	91.46	96.60	59.56	69.60
Intergenic regions (%)	8.54	3.40	40.44	30.40
rRNA	3	3	3	3
tRNA	34	34	34	33
tmRNA	1	1	1	1
RnaseP RNA	1	1	1	1
<i>dnaQ</i>	yes	yes	pseudo	no
Direct repeats	1	2	23	4
Inverted repeats	0	1	17	2
Tandem repeats	10	31	3	111



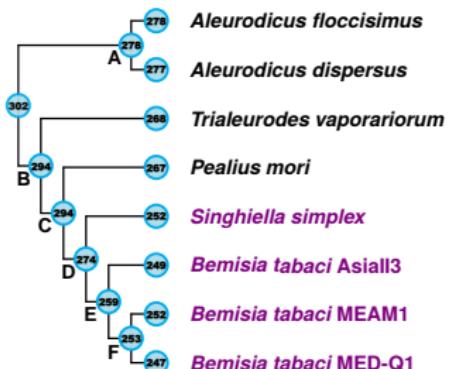


Portiera gene losses history



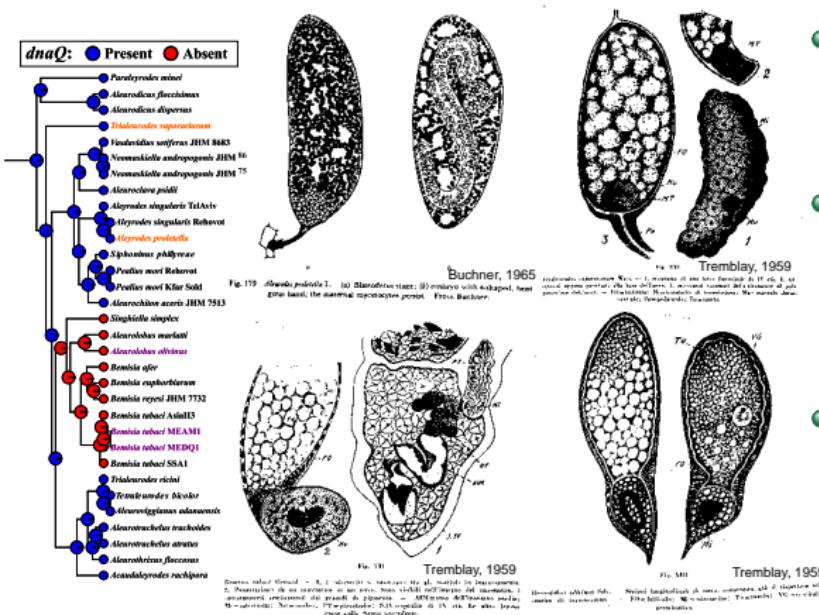
Species/MRCA	Gene name
<i>A. dispersus</i> (AIDI)	<i>ahpC</i>
<i>T. vaporariorum</i> (TeVa)	<i>rpmD</i> , <i>purA</i> , <i>ahpC</i> , <i>hisE</i> , <i>rplA</i>
<i>P. mori</i> (PeMo)	<i>clpP</i> , <i>clpX</i> , <i>lspA</i> , <i>rsmM</i> , <i>rsmH</i>
<i>S. simplex</i> (SiSi)	<i>rsmH</i> , <i>ahpC</i> , <i>hisA</i> , <i>tilS</i> , <i>trpF</i>
A	<i>purA</i> , <i>lepB</i>
B	<i>glyA</i> , <i>hupB</i> , <i>metG</i> , <i>miaA</i> , <i>rnc</i> , <i>tktA</i> , <i>yggF</i> , <i>alaS</i>
C	<i>clpB</i> , <i>lipB</i>
D	<i>purA</i> , <i>lepB</i> , <i>rpmD</i> , <i>dapF</i> , <i>deAD</i> , <i>dnaN*</i> , <i>dnaQ*</i> , <i>dnaX*</i> , <i>fri</i> , <i>holA*</i> , <i>holB*</i> , <i>lysA</i> , <i>mucD</i> , <i>mutL*</i> , <i>rsmA</i> , <i>rvuC*</i> , <i>trpS</i> , hypothetical protein
E/F	<i>clpP</i> , <i>clpX</i> , <i>lspA</i> , <i>argH</i> , <i>dapB</i> , <i>era</i> , <i>galP</i> , <i>sohB</i> , <i>ssb*</i> , <i>upp</i>

*DNA replication and repair.





The symbiont or the egg: origin of genome instability?



- In whiteflies, several (4-10) maternal bacteriocytes migrate into the oocyte.
- In *B. tabaci*, *B. gigantea*, and *A. olivinus* a single bacteriocyte enters the oocyte (derived state).
- Single-bacteriocyte inheritance decreases *Portiera*'s N_e compared to multi-bacteriocyte inheritance.

- Low N_e makes selection being unable to maintain the DNA repair and recombination machinery \Rightarrow origin of genome instability?



Take home message

- **Genome instability is rare in P-endosymbionts** and could point to interesting evolutionary events in both the symbiont and the host.
- A massive DNA replication and repair genes loss happen **in the MRCA of the Aleurolobini tribe** and is linked to *Portiera* genome instability.
- Instability is not exclusive of *B. tabaci* species: no role on diversification.
- **Single-bacteriocyte inheritance** could have **decreased *Portiera*'s N_e** , making purifying selection unable to maintain the DNA replication/repair machinery.
- Instability first → **host compensatory adaptation** to increase purifying selection each generation by **eliminating defective *Portiera*** populations.
- *B. tabaci* has acquired multiple bacterial genes to compensate for *Portiera* losses compared to *T. vaporariorum* (Bao et al., 2021; Ren et al., 2020, 2021):
 - Consequence of single-bacteriocyte inheritance or *Portiera* genome degradation?



Acknowledgments



**In memory of Prof. Dan Gerling
(1936–2016)**

Shai Morin, Natividad Mestre-Rincon

Einat Zchori-Fein



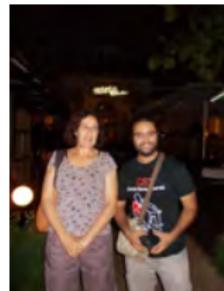
David Ouvrard

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Fundings:



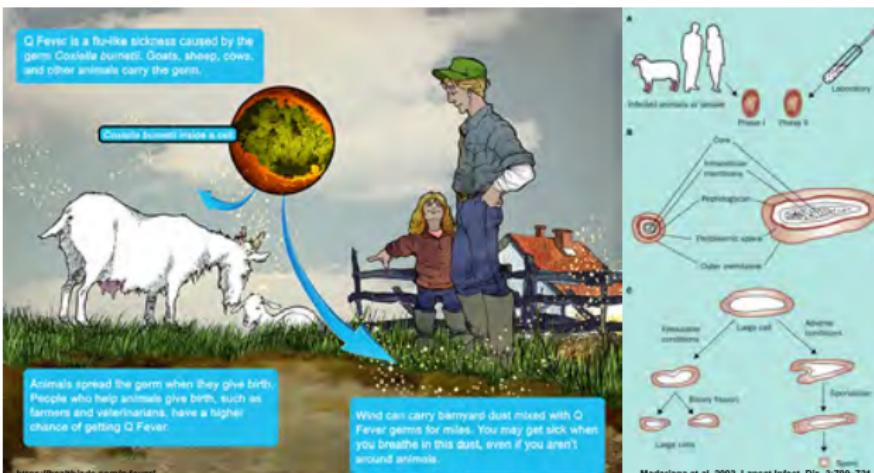
Postdoctoral Fellowships: Valazzi Pikovski Fund and The Golda Meir Fellowship Trust



Comparative genomics sheds new light on the evolution of pathogenicity in *Coxiella burnetii*.

Santos-Garcia et al. 2022. Unpublished.

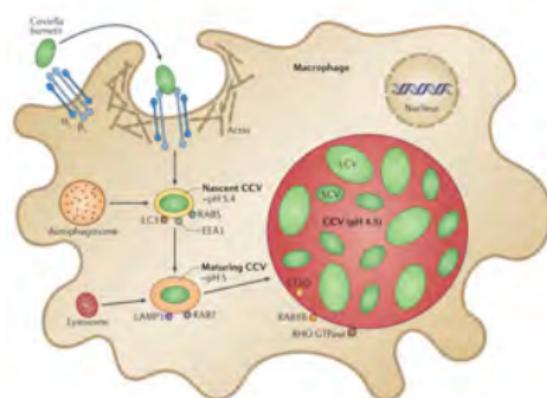
Coxiellaceae family: *Coxiella burnetii* & Q-fever



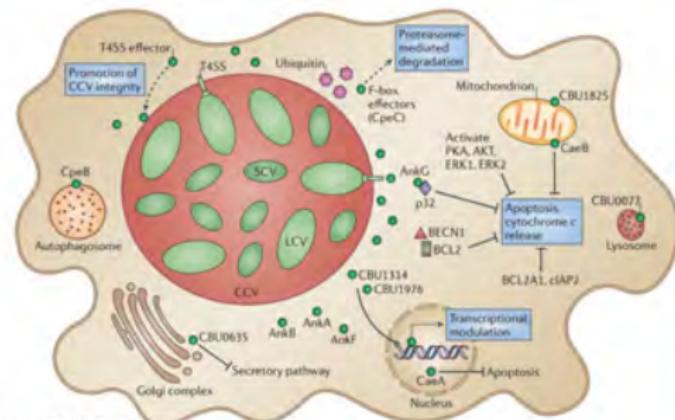
- The **Coxiellaceae family** (γ -proteobacteria) is composed of **five genera** showing lifestyles ranging from **free-living** to **mutualistic**.
- ***Coxiella burnetii*** is an **intracellular parasite** with a **biphasic life cycle**.

- The **Small Cell Variant (SCV)** is a **resistance form (~spore)**, persisting in the environment for extended periods → **infective** form.
- SCV is activated by the acid pH of the host cell phagolysosome → Large Cell Variant (**LCV**) → **replicative** and metabolically active **form**.

Coxiella burnetii pathogenesis

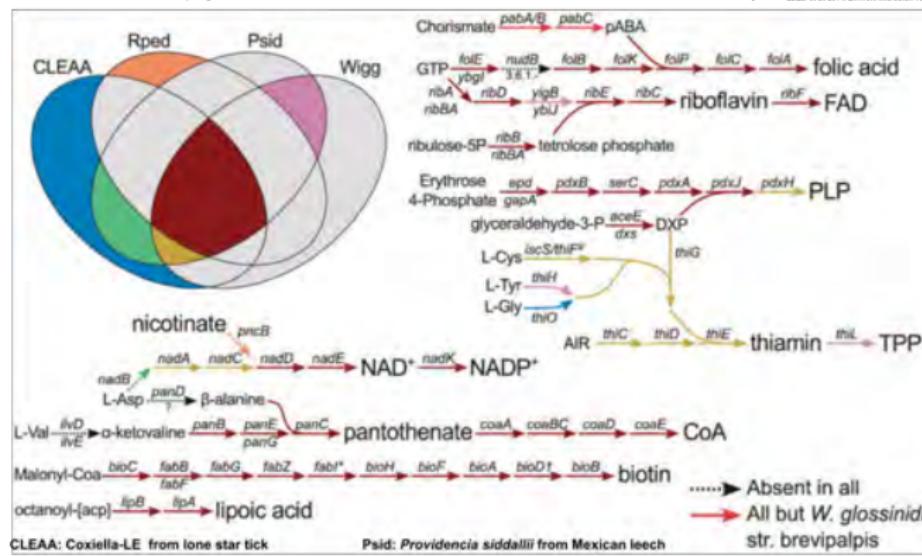
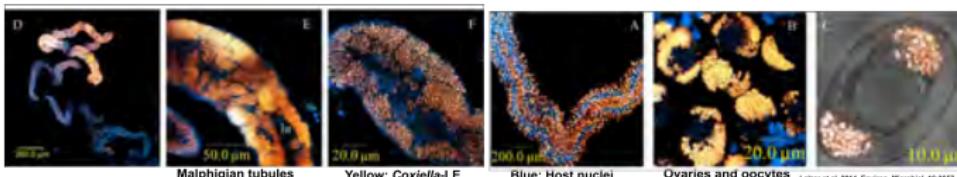


van Schaik et al. 2013. Nat. Rev. Microbiol. 11:561–73



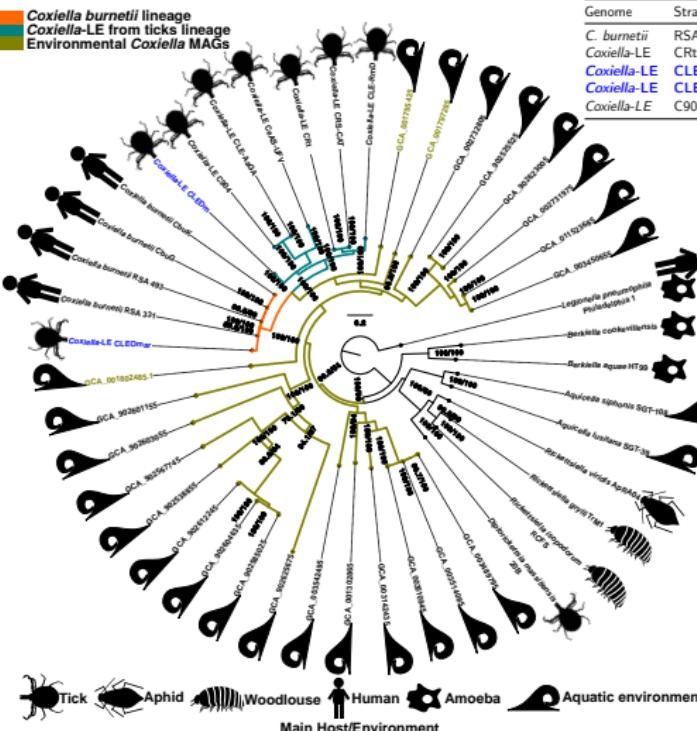
- The **SCV is activated after the phagolysosome formation** and starts to express the virulence factor Dot/Imc T4 Secretion System (SS).
- C. burnetii* uses the Dot/Imc SS to manipulate the host cell by inhibiting apoptosis and allowing the maintenance of the *Coxiella* Containing Vacuole (CCV) (~late phagolysosome - acid pH).
- The Dot/Imc SS is **essential** for *C. burnetii* intracellular survival.

The good relatives: *Coxiella*-like endosymbionts



- *C. burnetii* relatives have been found associated with ticks.
- **Coxiella-like endosymbionts (*Coxiella*-LEs or CLEs) supply their tick hosts with B-vitamins scarce in blood-based diets.**
- Are *Coxiella*-LEs domesticated pathogens?

Coxiella relatives and two new Coxiella-LEs

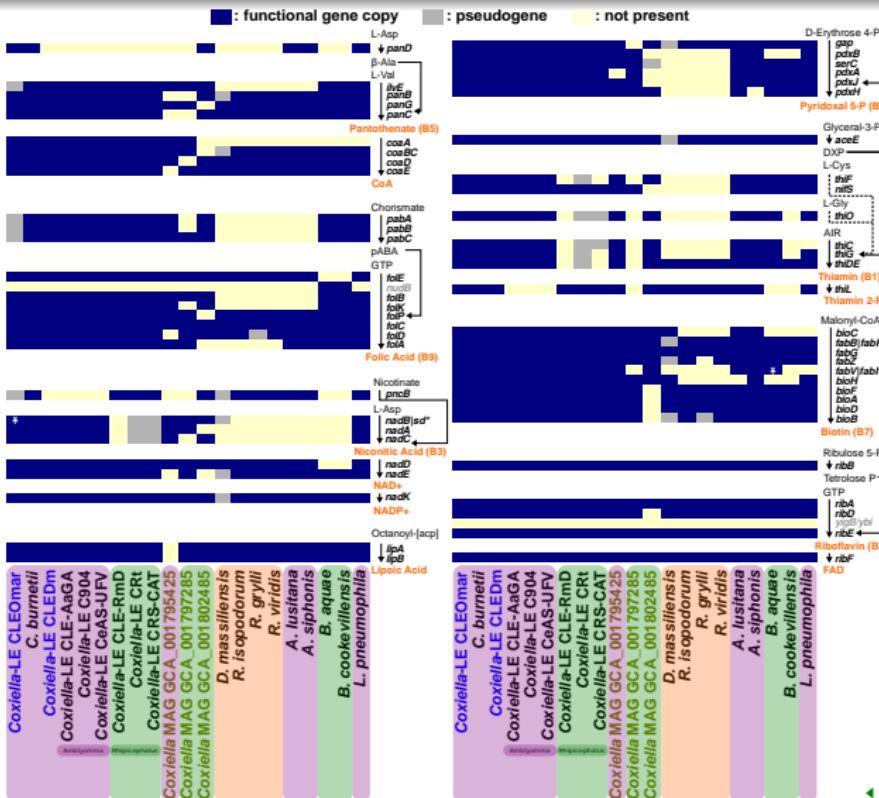


Maximum Likelihood phylogenomic tree of Coxiellaceae genomes inferred from a concatenated alignment of 143 core proteins.

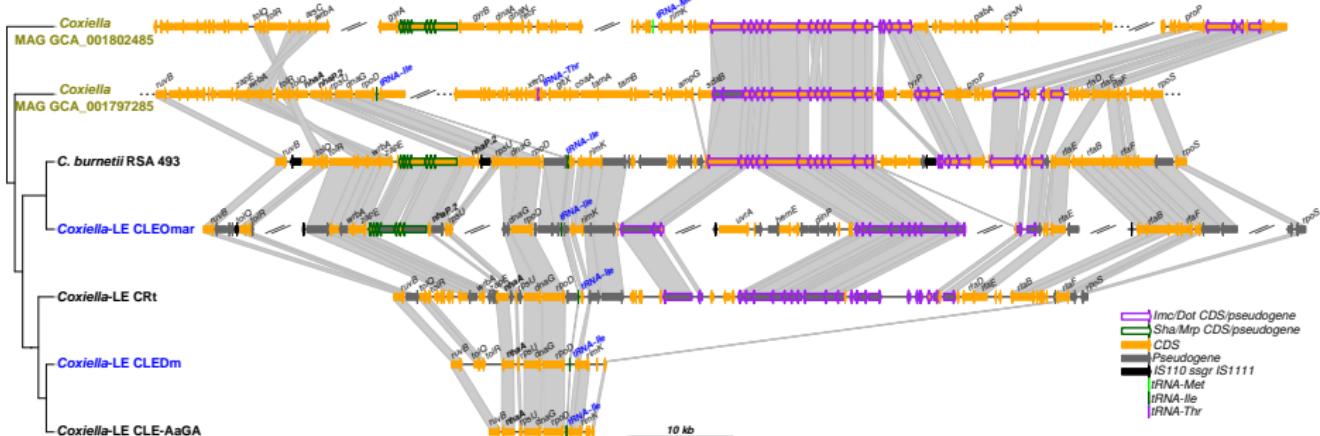
Genome	Strain	Host	Size (Mb)	CDS	Pseudogenes	IS types/copies	% GC	Contigs
<i>C. burnetii</i>	RSA493	Mammals/Amoebas	1.99	1833	207	6/32	42.6	1
<i>Coxiella</i> -LE	CRt	<i>Rhipicephalus turanicus</i>	1.73	1293	337	0/0	38.2	1
<i>Coxiella</i> -LE	CLEomar	<i>Ornithodoros maritimus</i>	1.83	976	608	4/31	41.5	112
<i>Coxiella</i> -LE	CLEDm	<i>Dermacentor marginatus</i>	0.9	658	15	0/0	35.1	1
	C904	<i>Amblyomma americanum</i>	0.66	565	3	0/0	34.6	1

- **CLEDm** is an **old endosymbiont**: small genome, low GC, no mobile elements.
- **CLEomar** is a **recent endosymbiont**: reduced genome with pseudogenes, mobile elements, and high GC.
- *C. burnetii*, *Coxiella*-LEs, and environmental *Coxiella* MAGs form a **monophyletic clade**.
- *C. burnetii*/Coxiella-LEs clade originated from aquatic bacteria able to establish symbiotic relationships.

Coxiellaceae biosynthetic potential

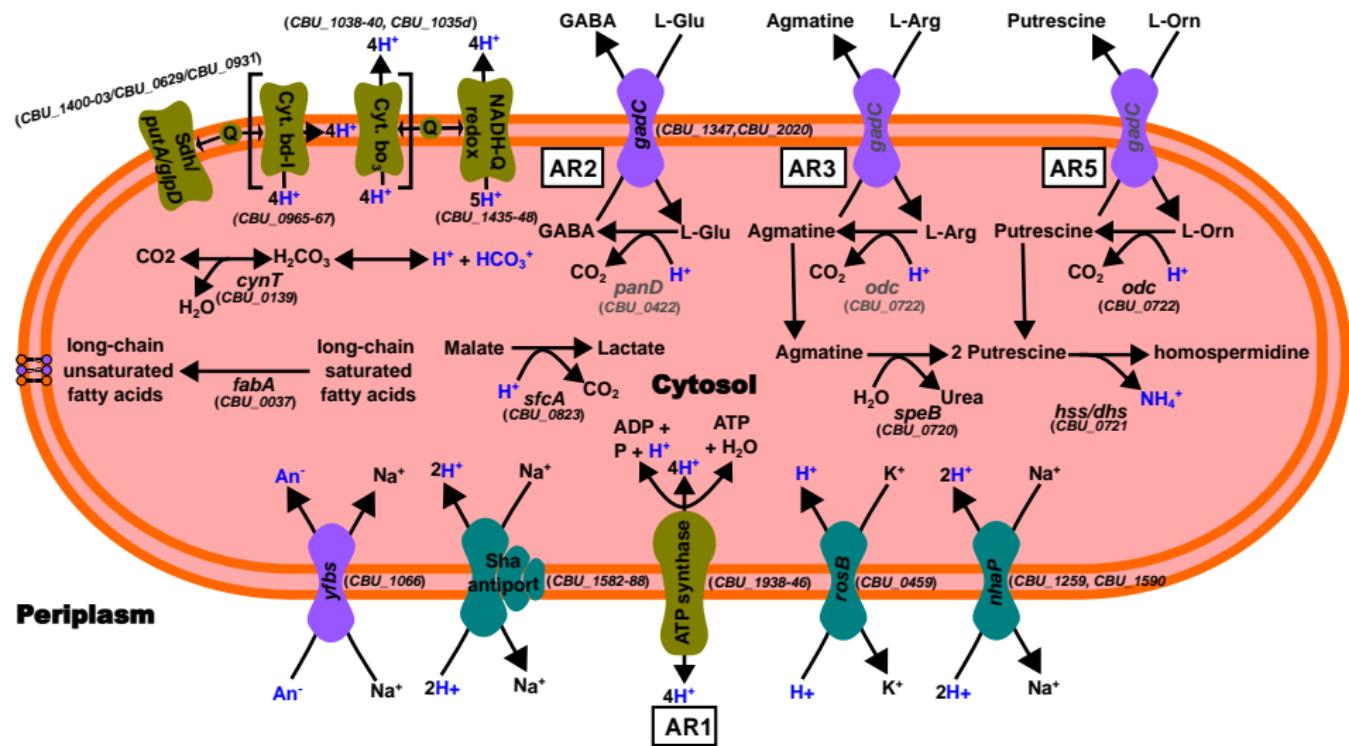


The Dot/Icm T4 Secretion System in Coxiellaceae

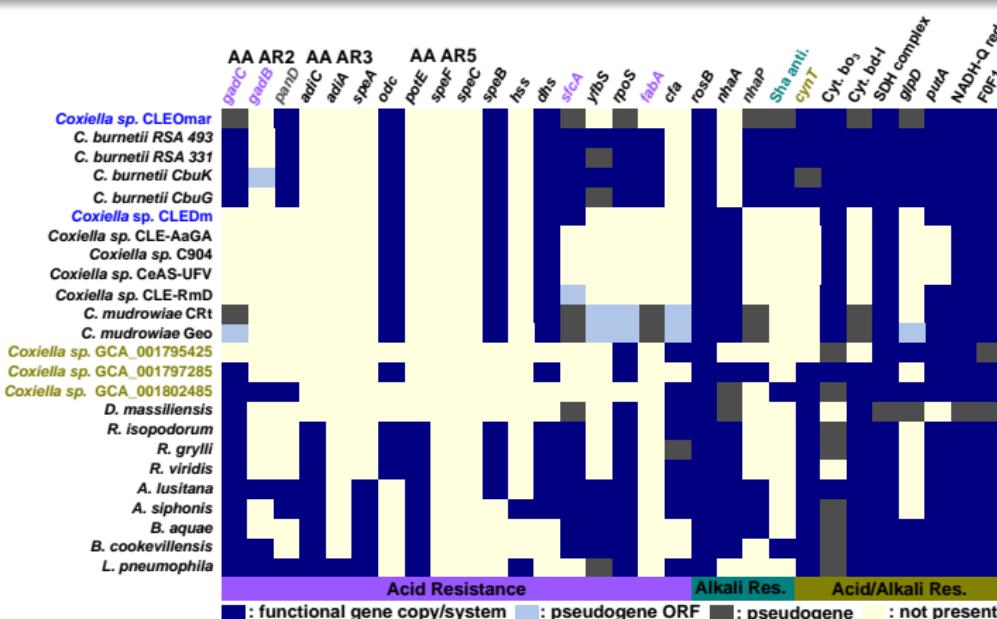


- The **Dot/Imc SS** is present in all Coxiellaceae except in reduced *Coxiella*-LEs: an ancestral trait for host cell invasion and manipulation (Brenner et al., 2021)
 - SSs losses are expected during evolution towards mutualism (Oakeson et al., 2014).
 - In *C. burnetii*, the **Dot/Icm SS** region presents signatures of **pathogenic islands** (PAIs): tRNAs, IS elements, DR, HT genes, and virulence factors (Hacker and Kaper, 2000).
 - *C. burnetii*'s PAI acquired additional genes (a **Sha/Mrp antiporter**) while in *Coxiella*-LEs is losing genes related to pathogenesis.

pH Homeostasis in *C. burnetii*



pH Homeostasis in Coxiellaceae



- *C. burnetii* potentially use three AA ARS.
- ***Coxiella*-LEs** are harbored in non-acidic host-derived vacuoles and **ARS are not required** (Brenner et al., 2021)

- **Sha/Mrp antiporter** is almost exclusive to *C. burnetii*'s lineage.

- Involved in alkali resistance, establishment of symbiotic interactions, and **sporulation** (Ito et al., 2001)

Take home message

- **Coxiella** ancestor was an **environmental facultative symbiont able to invade different hosts**, thanks to the presence of the Dot/Icm T4SS and the ability to deal with abiotic stress.
- This ancestor splitted in **two lineages**, one including ***C. burnetii*** and the other mainly **tick-associated symbionts**.
- **All *Coxiella*-LEs produce B vitamins**, which seems the reason for their mutualistic symbiosis with ticks (Duron and Gottlieb, 2020).
 - The potential to **synthesize vitamins is required for initiating mutualistic symbiosis** in ticks.
- The **Icm/Dot T4SS** is part of the same **pathogenic island** present in *C. burnetii* and *Coxiella*-LEs, although reduced in the last.
- The **Sha operon may be related to the SCV morphotype** of *C. burnetii* **and** their ability **to resist** alkaline environments found **outside the host**: future research on anti Q fever strategies?



Acknowledgments

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AGENCE NATIONALE DE LA RECHERCHE

ANR hMICMAC: Host-Microbiota
Co-Adaptations - Mechanisms
and consequences.

Projets Exploratoires Premier Soutien
(PEPS) Interdisciplinaires



Diego Santos-Garcia, Ph.D.

Olivier Duron



Valérie Noël



Karen McCoy



Koret School of
Veterinary
Medicine



Yuval Gottlieb



UPPSALA
UNIVERSITET



Lisa Klasson

Disentangling the multitrophic interactions of the supervector *Bemisia tabaci* to potentially use its symbiotic communities to reduce plants viral-vectored diseases.

GuardSym MSCA-IF Grant agreement ID 885583

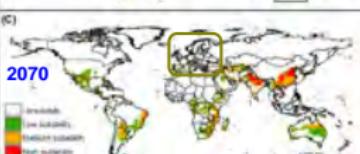
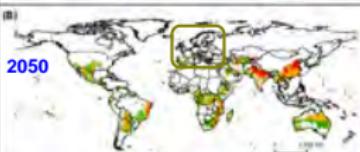
"The FAO estimates that between 20 % and 40 % of world agricultural production is lost each year due to pests.

Plant diseases cost the world economy \$220 billion annually, and invasive insects \$70 billion (www.fao.org)."

"Around 47 % of plant diseases are caused by viruses, most of them vectored by insects (Navas-Castillo *et al.* 2011)."

Intensive agriculture and Global warming

Future predicted distribution of *B. tabaci*



Ramos RS et al. 2018. PLoS ONE 13 (9): e1–20. <https://doi.org/10.1371/journal.pone.0198025>.



- Intensive agriculture: **biodiversity and beneficial insects reduction.**
 - Monocultures and irrigation.
 - Introduction of non-native crops.
 - Chemical abuse: fertilizers, herbicides, and insecticides.
- Global warming: **expansion of pests insects and vectored diseases.**
 - Increased temperatures, changes in rainfall patterns and desertification.
- Most **phyto-viruses require a vector to infect new plants:**
 - **Sap sucking insects**, such as whiteflies, are the most **efficient vectors**.

Whiteflies: a supervector



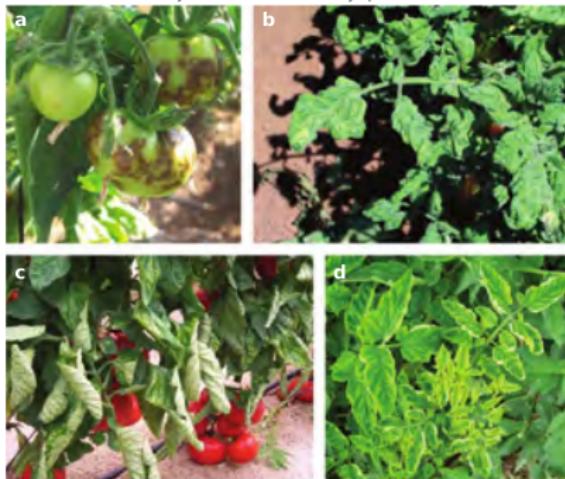
- The sweet-potato whitefly *Bemisia tabaci* is a tiny insect that feeds on plant sap through a stylet (modified mandible).
- Many emergent phyto-viral diseases are linked to the expansion of *B. tabaci* (Gilbertson *et al.* 2015).

- Feeds on more than 600 plants, including crops such as:

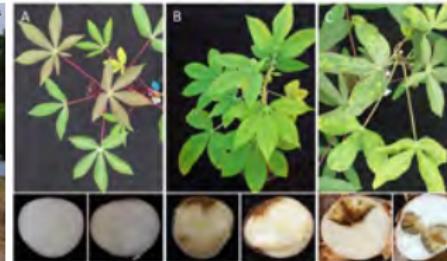
- Cassava** (Malpighiales) -> Staple food in Africa.
- Potato, sweet-potato, tomato, pepper, aubergine... (Solanaceae).
- Pumpkin, zucchini, melon... (Cucurbitaceae).
- Beans, broad beans, soybeans, alfalfa, peanuts... (Legumes).
- Cauliflower, broccoli, cabbage... (Brasicaceas).

Whiteflies: a supervector

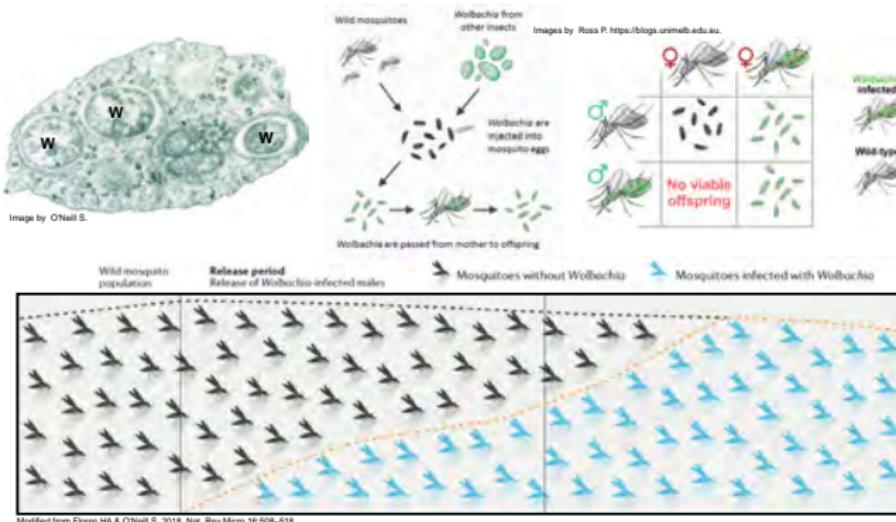
Navas-Castillo J et al. 2011. Annu. Rev. Phytopathol. 49:219-248.



- Develops resistance to pesticides.
- Transmits more than 300 phyto-viruses: global pandemic in tomato (TYLCV) and cassava (CSBD and ACMV).
- Need for Integrated Pest Management (IPM): set of strategies and practices that respect the environment, **minimizing the use of pesticides**.



Mosquitoes, dengue, and population replacement (IPM)



- In Queensland (Australia) it has been possible to **eliminate dengue** since 2011 using ***Wolbachia*** (299Km^2).
- *Wolbachia* is an intracellular bacterium present in $>50\%$ of insects.

- *Wolbachia* is **able to invade natural mosquito populations** by manipulating their reproduction (Cytoplasmic Incompatibility or CI).
- *Wolbachia*-infected mosquitoes do not transmit dengue, Zika, chikungunya, and yellow fever viruses (Heather & O'Neill 2018).

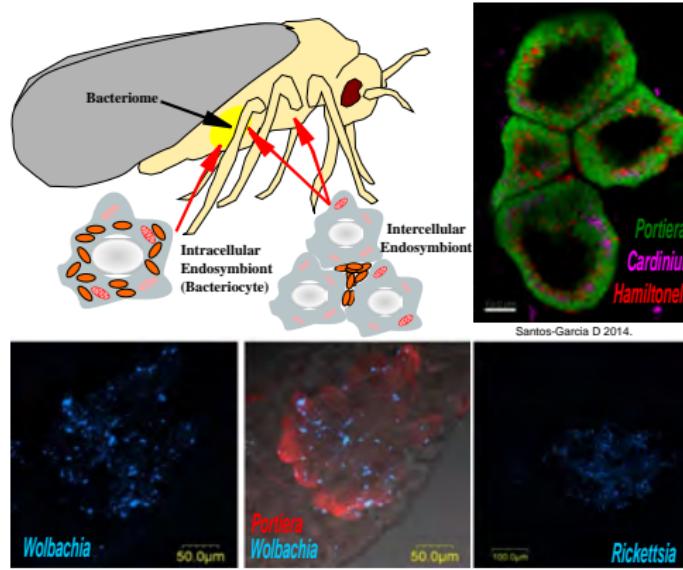
The Guardian Symbionts (GuardSym) project focuses on identifying **symbiotic bacteria** naturally present **in whiteflies** that may confer **viral resistance**.

GuardSym tries to lay the foundations for the **development of population modification strategies** in which natural populations are replaced by others unable to vector viral diseases.

Insects artificially infected with natural strains of *Wolbachia* (or other symbiotic bacteria) are not considered Genetically Modified Organisms - **permitted use** in the European Union.

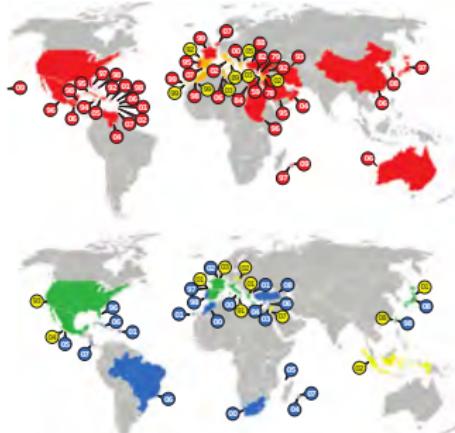
Symbiotic bacteria in whiteflies

- All whiteflies host **various symbiotic bacteria**.
 - Bacteriocyte: cells specialized in harboring symbiotic bacteria.
 - *Portiera* is essential to the insect and is always present.



GuardSym: Basic Objectives

- Study **insect-vectored viral diseases** as a set of **multitrophic interactions** between insects, bacteria, viruses, and plants.
- Establish the whitefly *B. tabaci*, one of the worst invasive species and present in Europe, as a viral vector model.
- Unravel multitrophic interactions in two of the most widely distributed tomato model viruses: TYLCV (Begomovirus) and ToCV (Crinivirus).

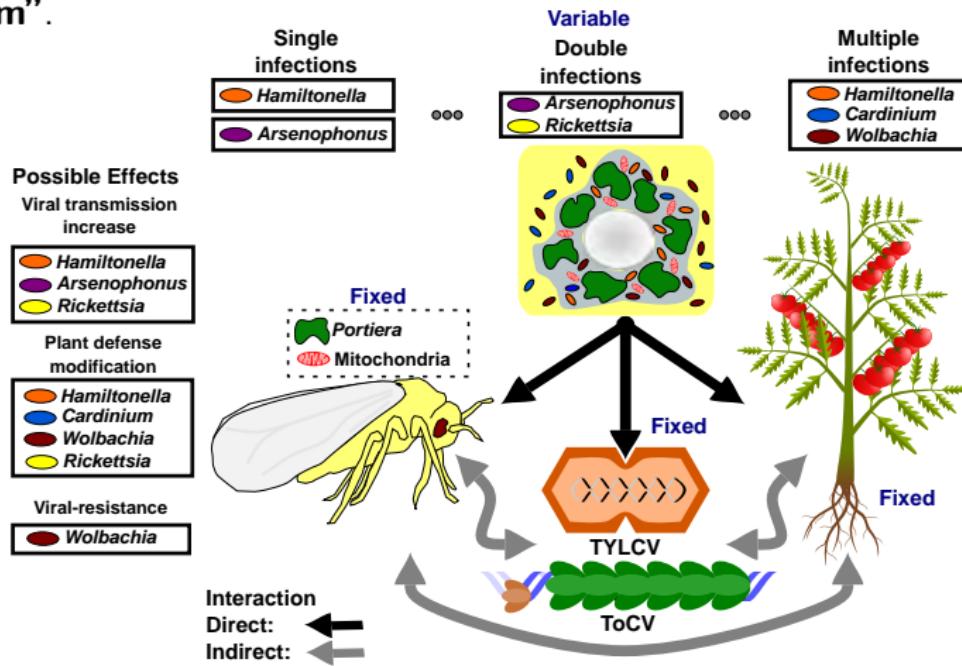


Supervector	Genus	Properties		Mode of transmission	Number of species*
		Virion shape and size	Genetic material		
<i>Bemisia tabaci</i> (sweet potato/tobacco whitefly)	<i>Begomovirus</i>	Geminate, 18 × 30 nm	ssDNA	Persistent circulative	288
	<i>Ipomovirus</i>	Flexuous, filamentous, 12–15 × 800–950 nm	ssRNA	Semipersistent	6
	<i>Crinivirus</i>	Flexuous, filamentous, 10–13 × 650–900 nm	ssRNA	Semipersistent	13
	<i>Carlavirus</i>	Flexuous, filamentous, 12–13 × 470–1,000 nm	ssRNA	Semipersistent	2 (of 52) ^b
	<i>Torradovirus</i>	Spherical, 28–30 nm	ssRNA	Semipersistent	2 (3) ^c

Gilbertson RL et al. 2015. Annu. Rev. Virol. 2:67–93.

GuardSym: Applied Objectives

- Identify useful symbiotic bacteria, or their combinations, for IPM strategies in whiteflies and other sucking insects by fixing the rest of “variables” in the “system”.



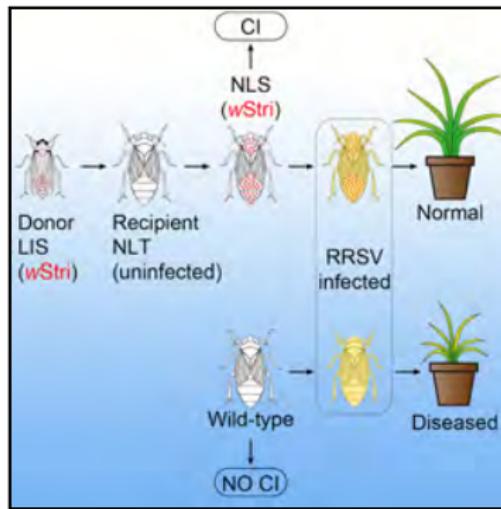
Use of symbionts in other agricultural pests

Article

Current Biology

Stable Introduction of Plant-Virus-Inhibiting Wolbachia into Planthoppers for Rice Protection

Graphical Abstract



Authors

Jun-Tao Gong, Yongjun Li,
Tong-Pu Li, ..., Xiao-Yue Hong,
Ary A. Hoffmann, Zhiyong Xi

Correspondence

xyhong@njau.edu.cn (X.-Y.H.),
ary@unimelb.edu.au (A.A.H.),
xizy@msu.edu (Z.X.)

In Brief

Gong et al. report the first successful transfer of a *Wolbachia* endosymbiont into a pest planthopper. The endosymbiont self-spreads into host populations and inhibits transmission of a rice plant virus by *Nilaparvata lugens*. The findings open the way for a new approach to limiting crop damage caused by agricultural pests.



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That's all Folks!



Have fun!



Index



1 *Portiera* gets wild: Genome instability provides insights into the evolution of both whiteflies and their endosymbionts

- Aleyrodidae superfamily or whiteflies
- Whiteflies endosymbionts
- Whiteflies-*Portiera* co-evolution

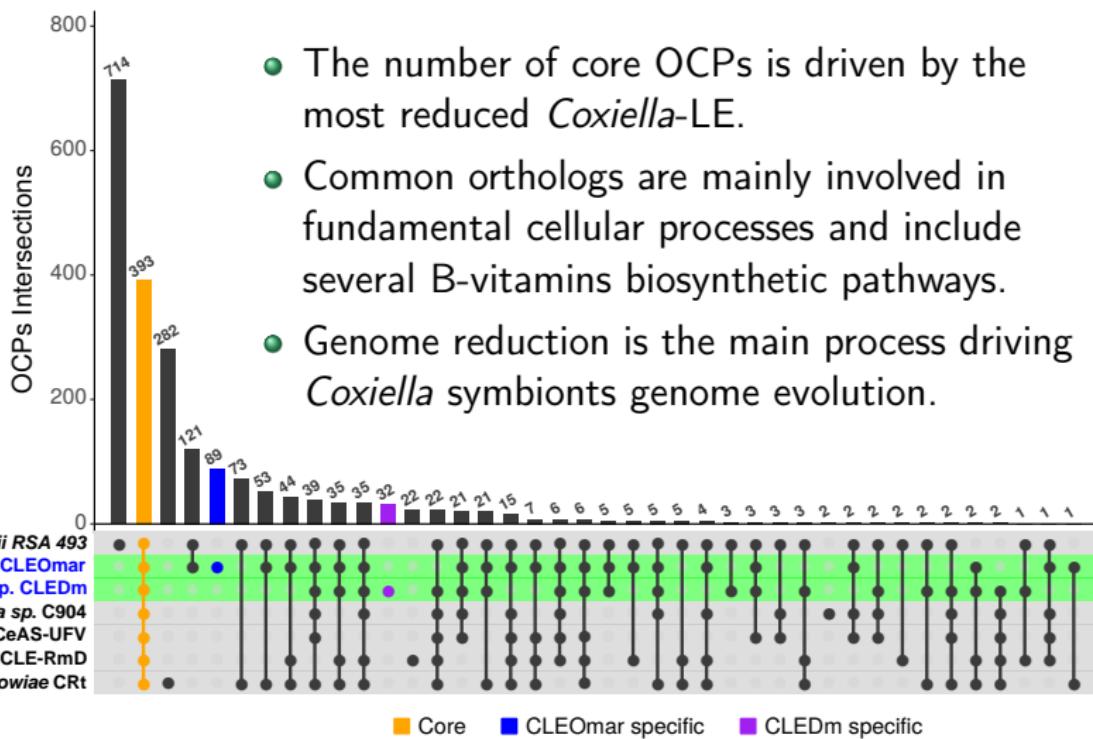
2 Comparative genomics sheds new light on the evolution of pathogenicity in *Coxiella burnetii*.

- The Coxiellaceae family
- Coxiellaceae genome evolution
- Pathogenesis evolution & pH homeostasis

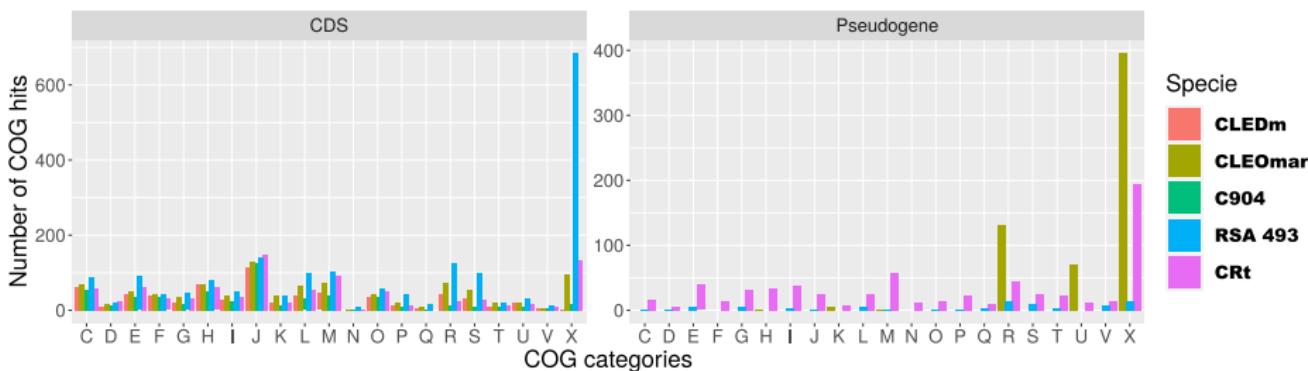
3 Disentangling the multitrophic interactions of the supervector *Bemisia tabaci* to potentially use its symbiotic communities to reduce plants viral-vectored diseases.

- Viral emergent diseases
- Whiteflies: a supervector
- Guardian Symbionts MSCA

C. burnetii & *Coxiella*-LE OCPs

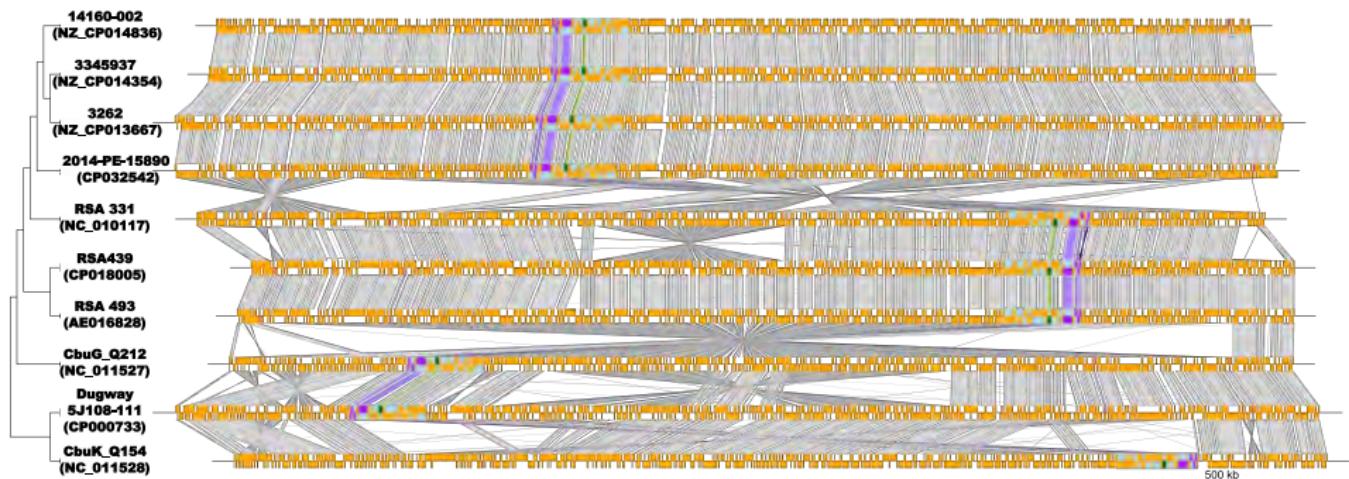


COG categories

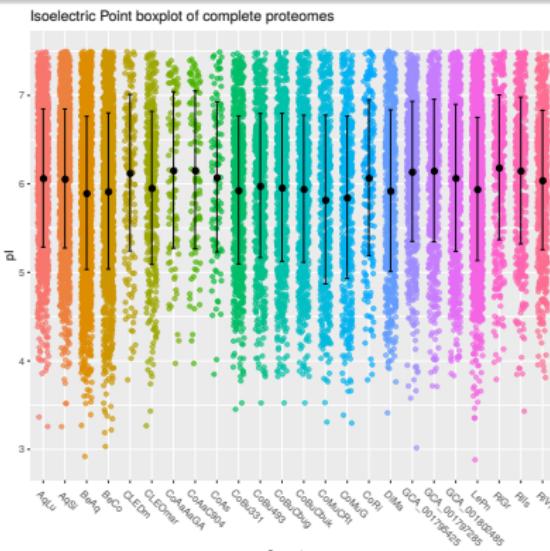
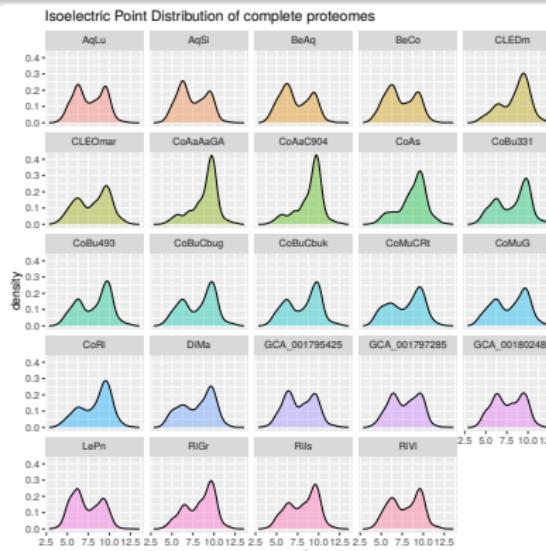


- Most *C. burnetii* species specific OCPs have unknown functions - few pseudogenes.
- CLEDm similar to CoAaC904 (*Amblyomma*), CLEOmar similar to CoMuCRt (PCA?)
- CoMuCRt pseudogenes: largest amount of inactivations are in X category
- CIEOmar pseudogenes: mainly R, U and X → environmental related gene

PAI synteny in *C. burnetii*

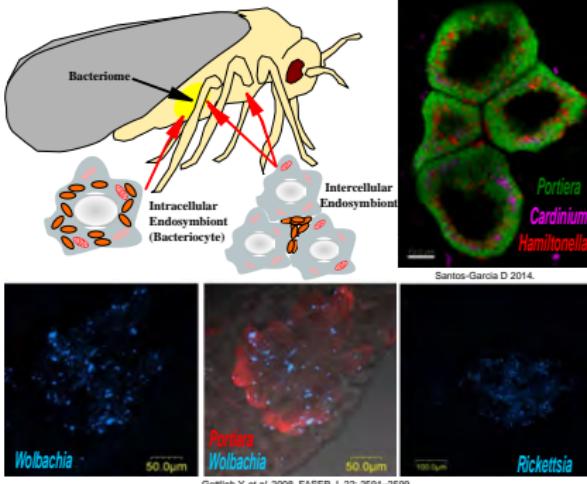


pH homeostasis: protein buffering



- Acid-resistance may be conferred by basic proteins - “proton” buffering.
- *C. burnetii* encodes large number of basic proteins (average pl ~ 8.25) compared to other bacterial sequenced genomes (e.g. *Helicobacter pylori*).
- Average pl values and distribution are not so different from other *Coxiellaceae*.

Symbiotic bacteria in *B. tabaci*



- *B. tabaci*, and all whiteflies, exhibit various symbiotic bacteria.
- *Portiera* and *Hamiltonella/Arsenophonus* are indispensable for *B. tabaci*: produce amino acids and essential vitamins absent in the sap (Santos-Garcia D et al. 2018).
- *Hamiltonella* reduces plant defenses against *B. tabaci* (Su Q et al. 2015).

- *Hamiltonella*, *Arsenophonus* and *Rickettsia* facilitate the direct or indirect transmission of different phytopiruses (Milenovic et al. 2021).
- *Cardinium*, *Wolbachia* and *Rickettsia* can be injected into plants and/or directly interact with phyto-viruses (Czosnek H et al. 2017)

Guardian Symbionts: introgression

