

# Évaluer le risque d'émergence de zoonoses à travers le prisme de l'immunoécologie et de l'immunogénétique



## Les rongeurs et hantavirus responsables de fièvres hémorragiques



CBGP – 08 janvier 2019

nathalie.charbonnel@inra.fr



MERCI

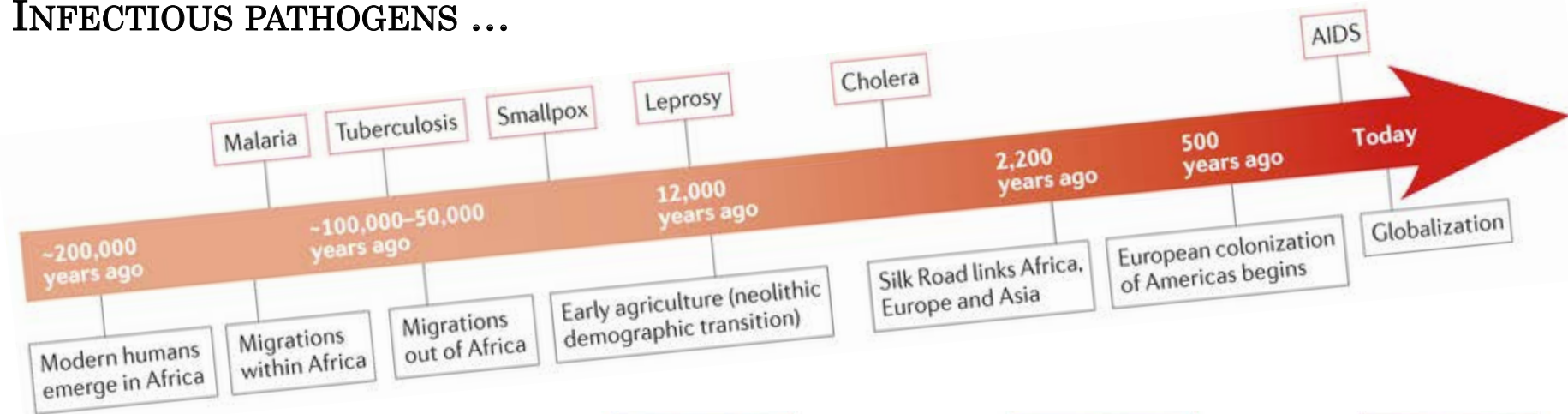




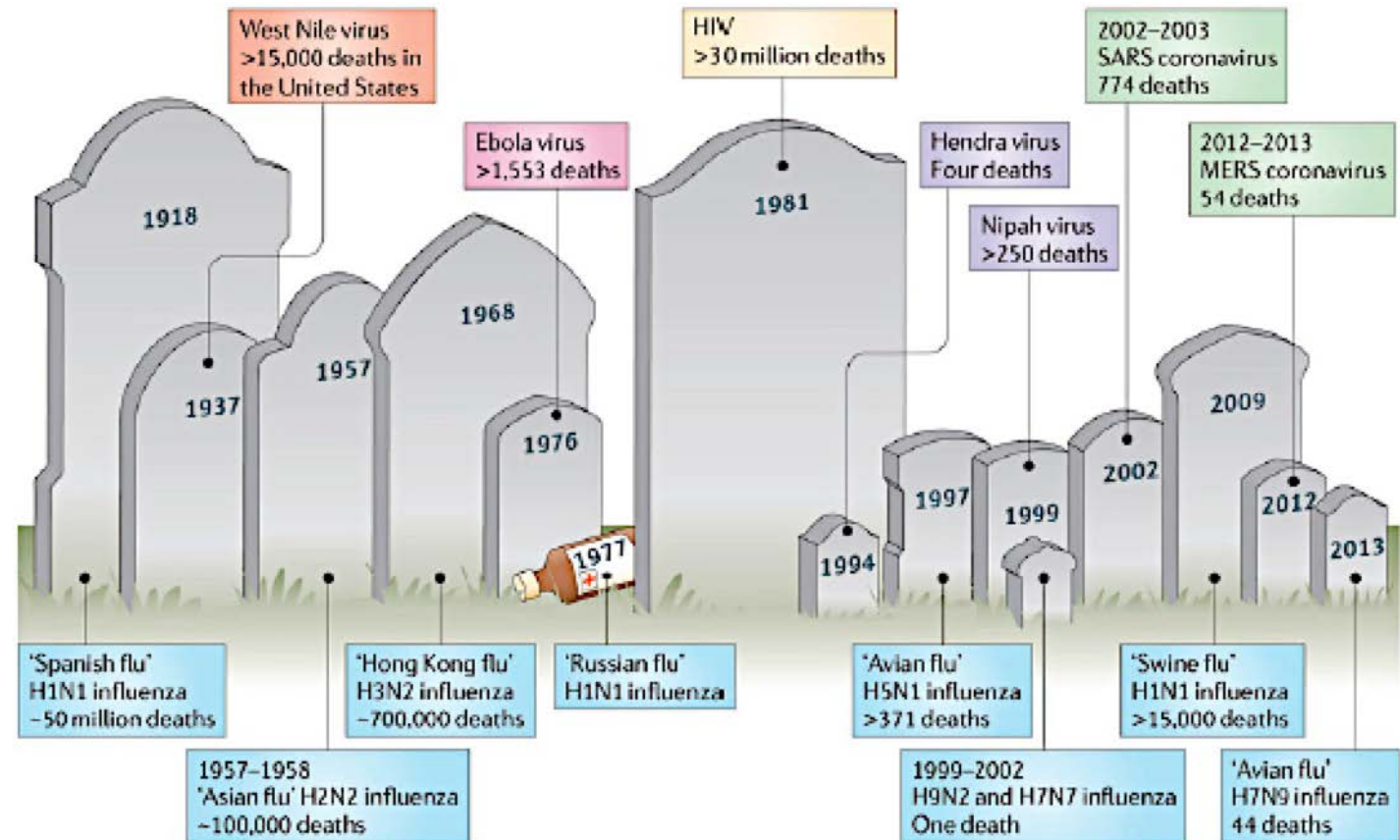
**CONTEXT**



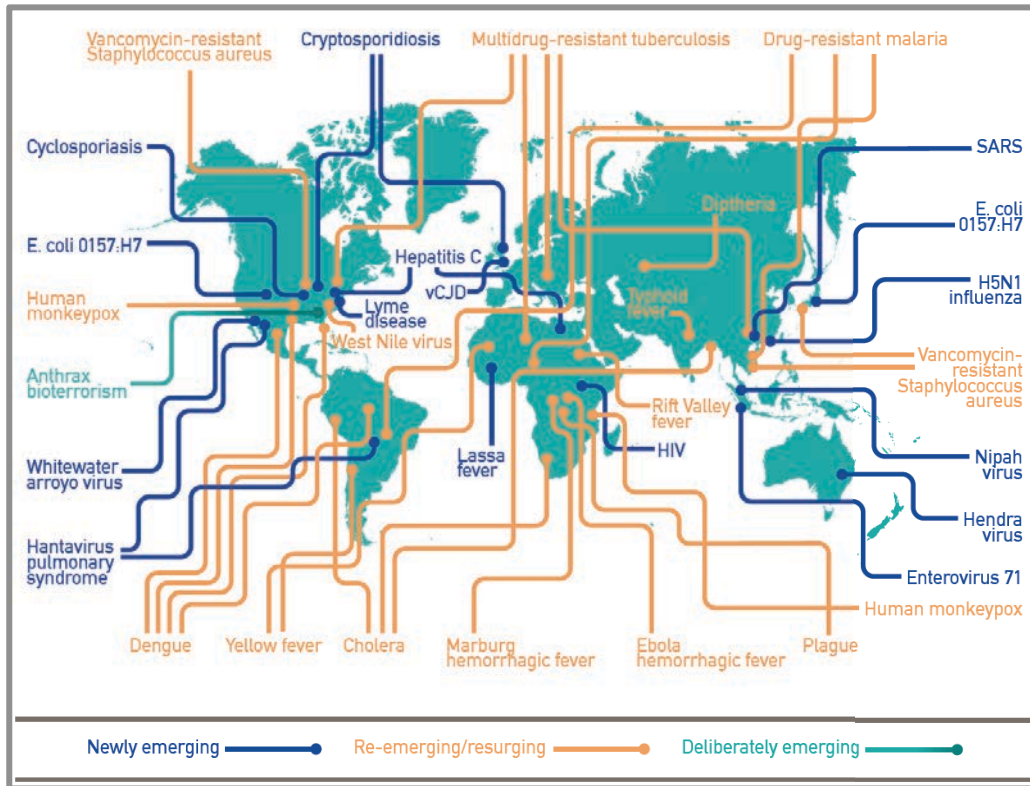
# INFECTIOUS PATHOGENS ...



Karlsson et al 2014

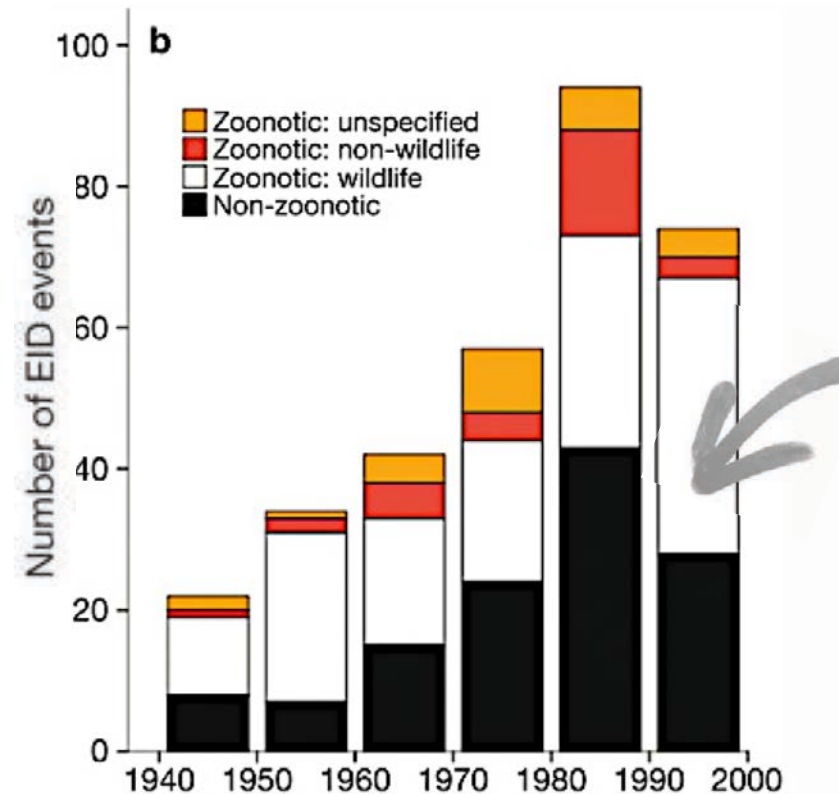


## ... EMERGING INFECTIOUS DISEASES



1 EID discovered every 8 months !

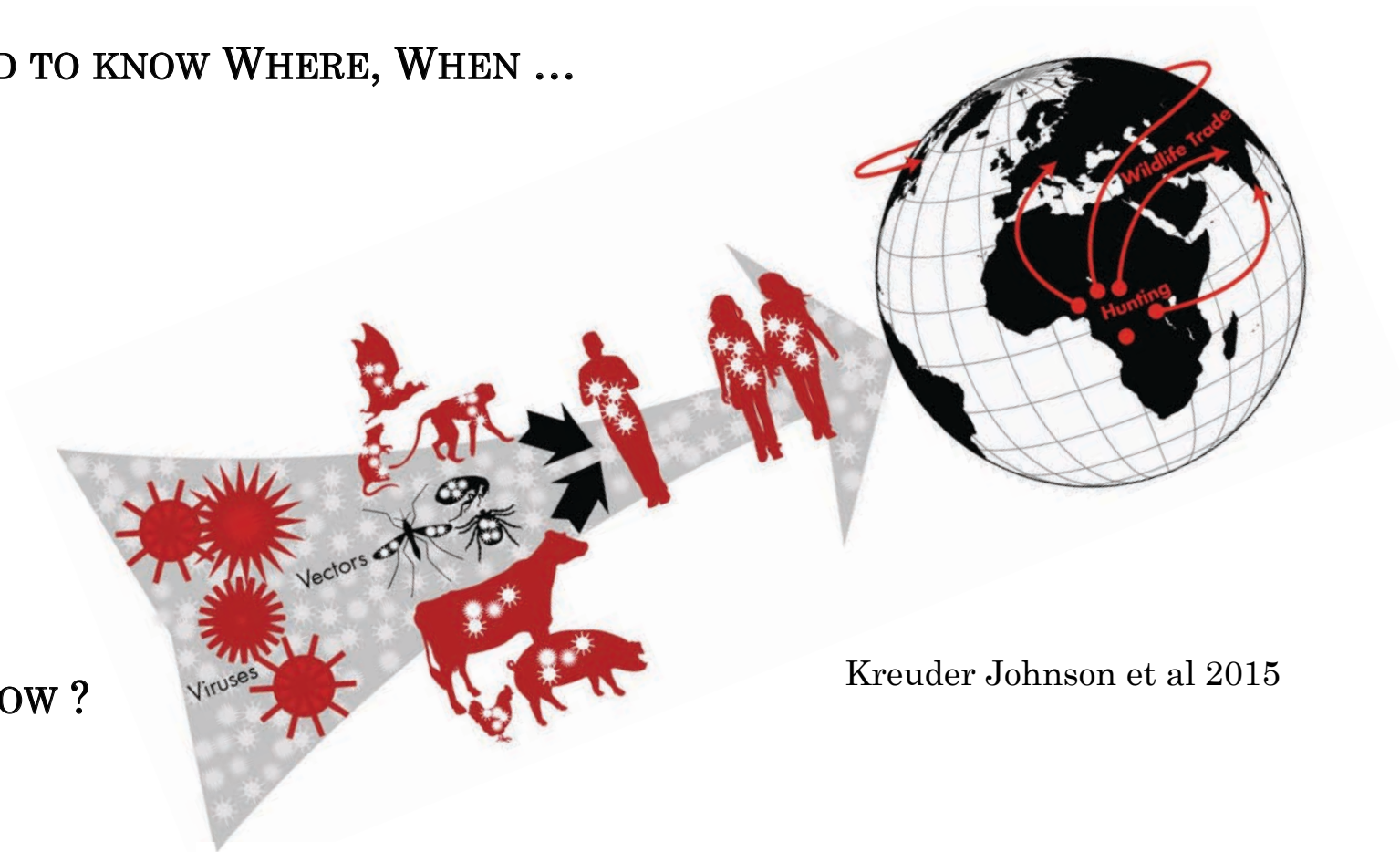
## ... and Zoonoses



Jones et al 2008

60% of EID are of animal origin  
75 % are associated with wildlife

➔ NEED TO KNOW WHERE, WHEN ...



➔ ... HOW ?

Kreuder Johnson et al 2015

## Reservoir Host Immune Responses to Emerging Zoonotic Viruses

Judith N. Mandl<sup>1,7,\*</sup>, Rafi Ahmed<sup>2</sup>, Luis B. Barreiro<sup>3</sup>, Peter  
Herbert W. Virgin<sup>5</sup>, and Mark B. Feinberg<sup>6</sup>

## Rodent reservoirs of future zoonotic diseases

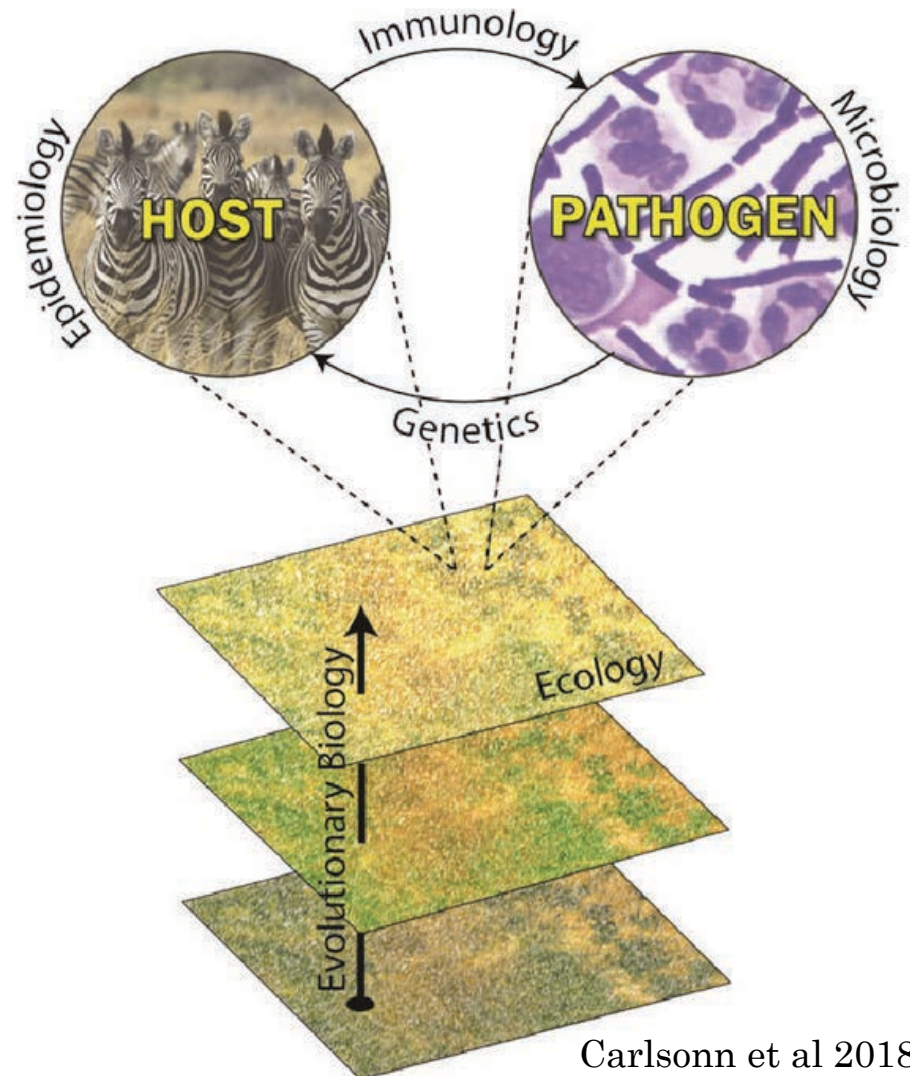
Barbara A. Han<sup>a,1</sup>, John Paul Schmidt<sup>b</sup>, Sarah E. Bowden<sup>b</sup>, and John M. Drake<sup>b</sup>



# IMMUNE RESPONSE VARIABILITY

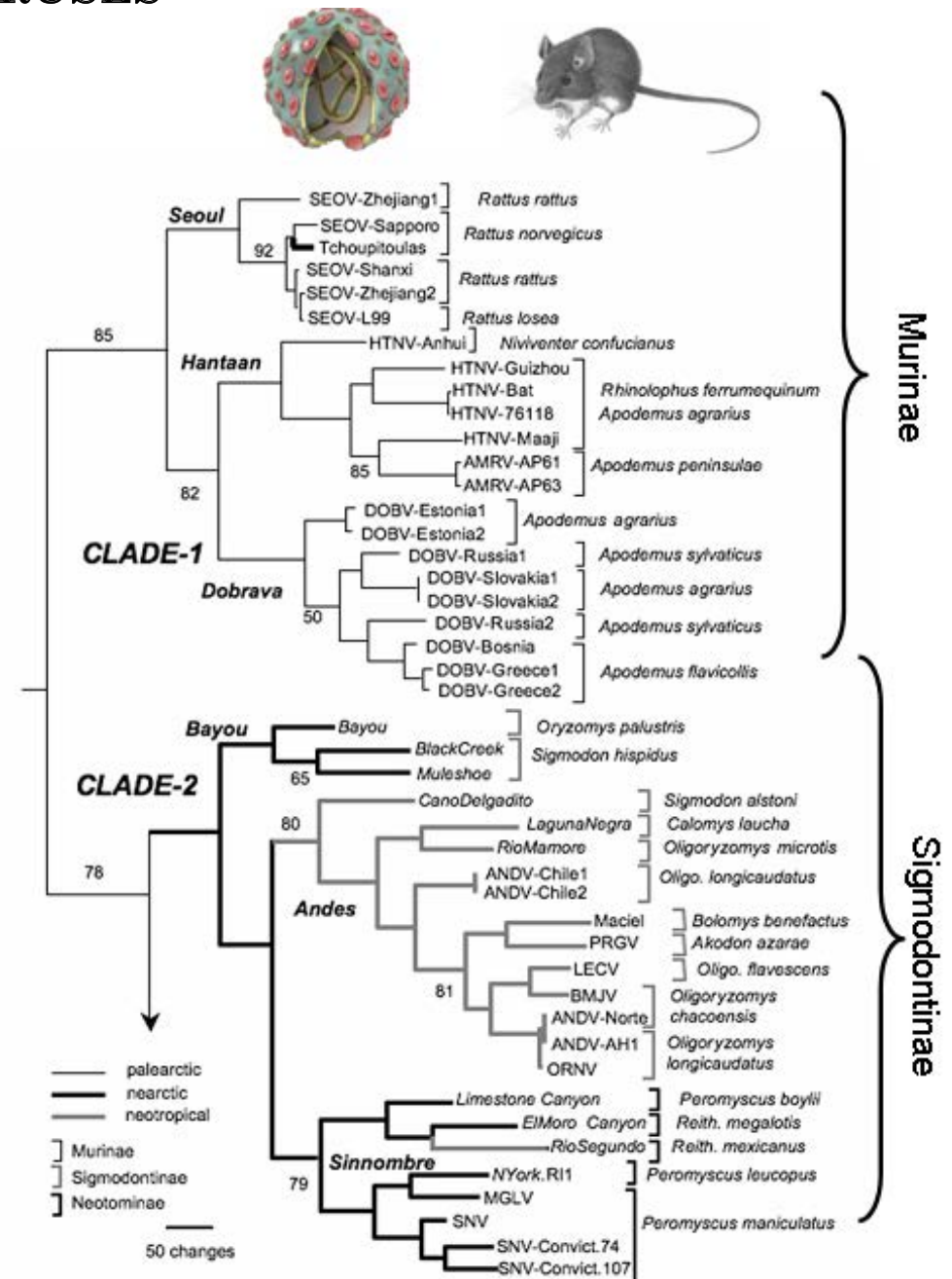
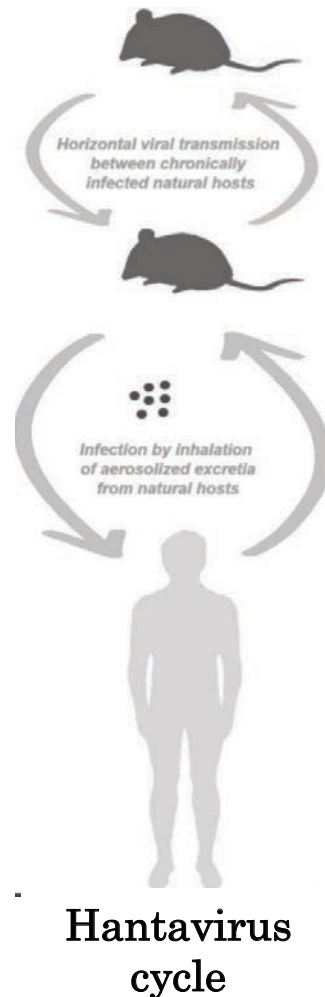
Need for studying immune variations  
in an evolutionary ecology context

Between population variability  
is still underlooked in wildlife



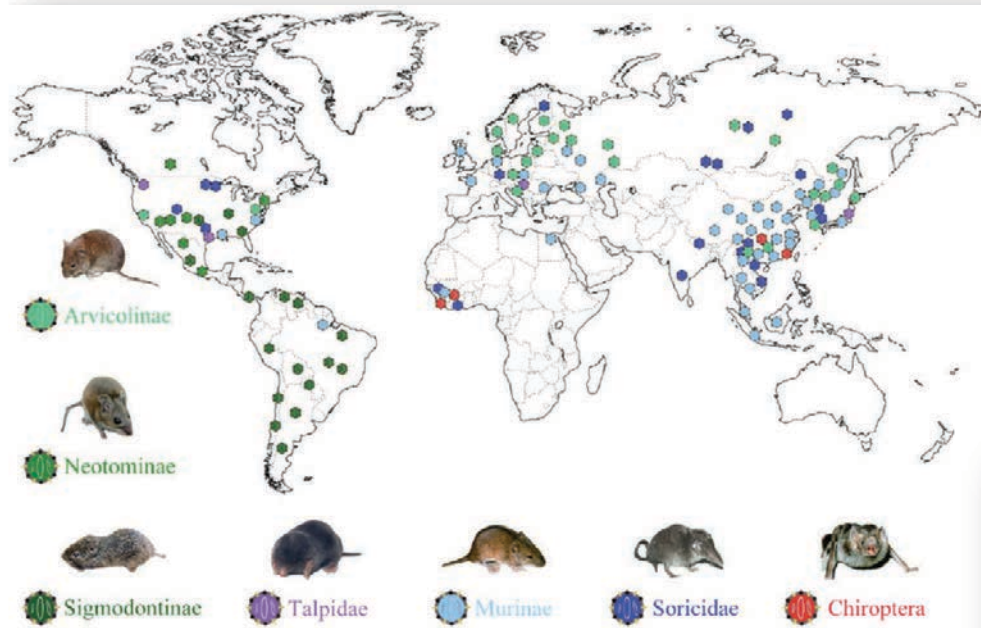
Carlson et al 2018

# THE PARTICULAR CASE OF HANTAVIRUSES

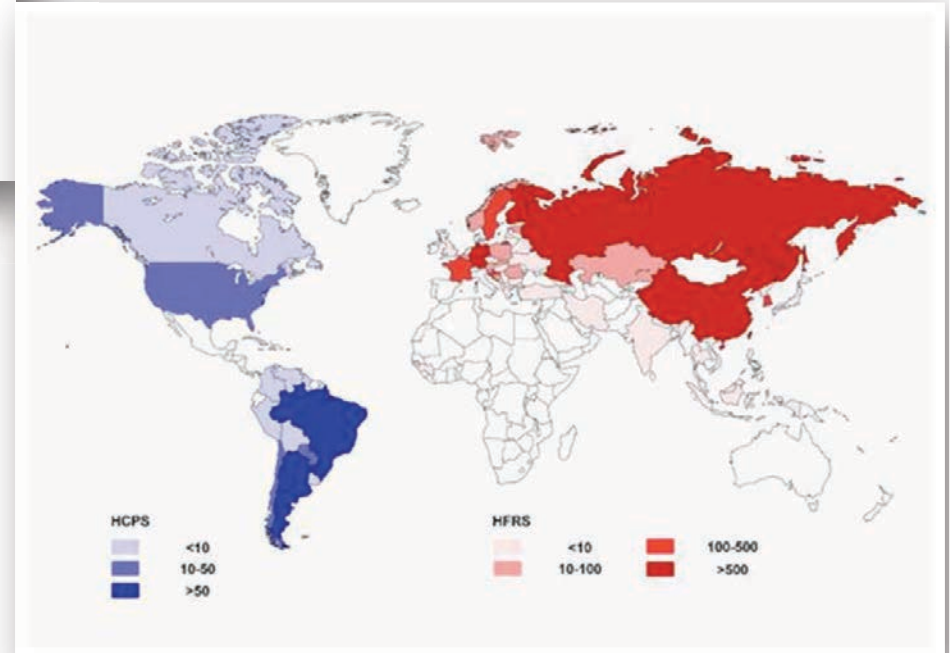




# HETEROGENEOUS DISTRIBUTION OF HANTAVIRUS CASES WORLDWIDE

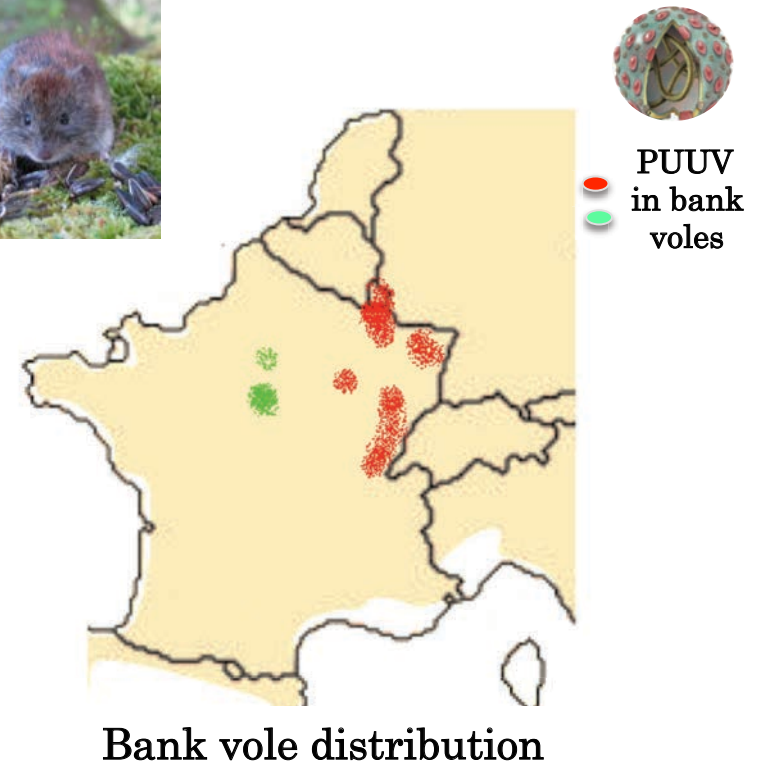
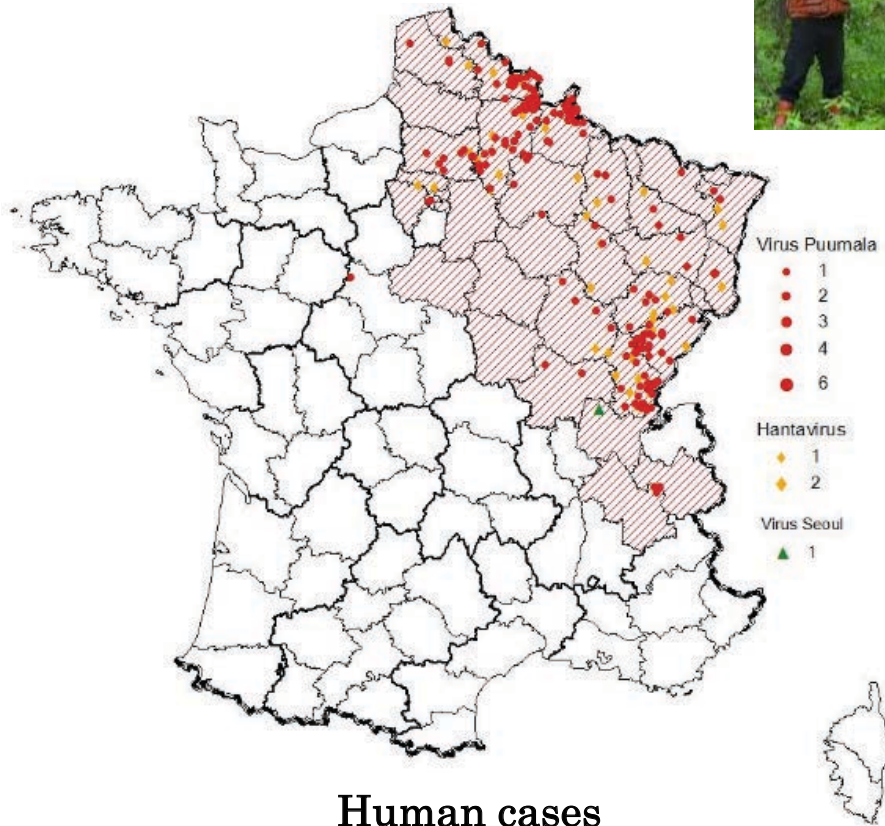
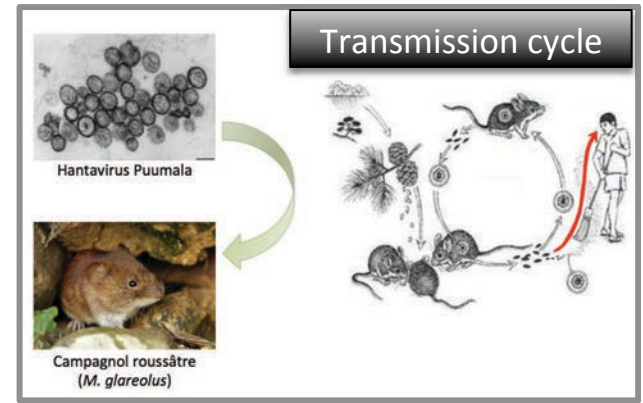


Guo et al. 2013

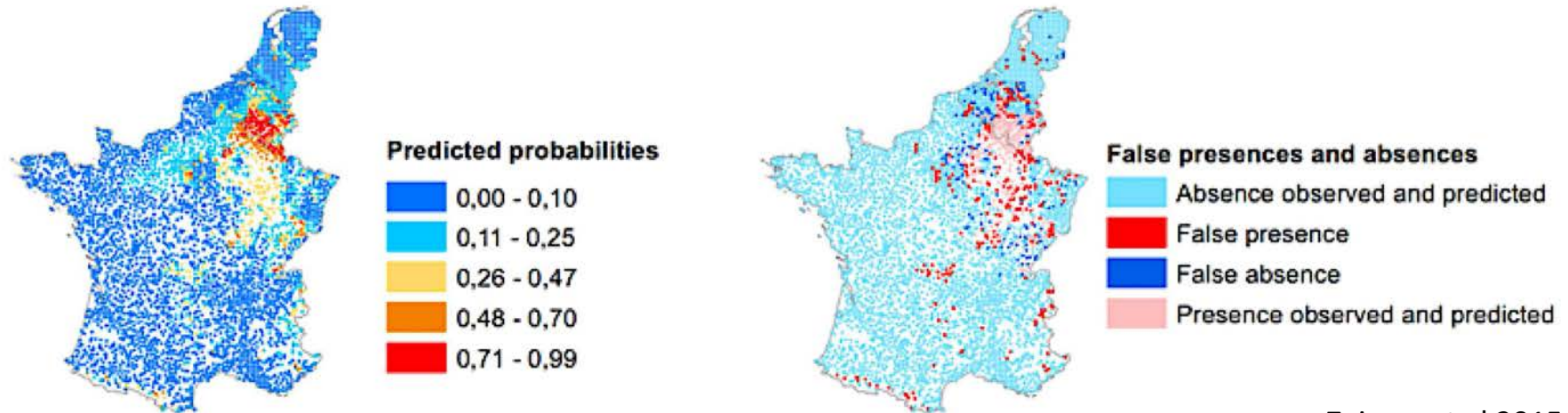


Jiang et al. 2017

# HETEROGENEOUS DISTRIBUTION Ex : NEPHROPATHIA EPIDEMICA CASES (PUUV HANTAVIRUS) IN FRANCE



# MODELING AND PREDICTING WHERE AND WHEN SHOULD PUUV HUMAN CASES OCCUR



Zeimes et al 2015



Abiotic factors (climate, environmental features) do not alone enable to predict the distribution of PUUV



Do biotic factors (host, virus, pathobiome) improve our understanding of the distribution and risk of PUUV ?



# THE IMMUNE PERSPECTIVE ...

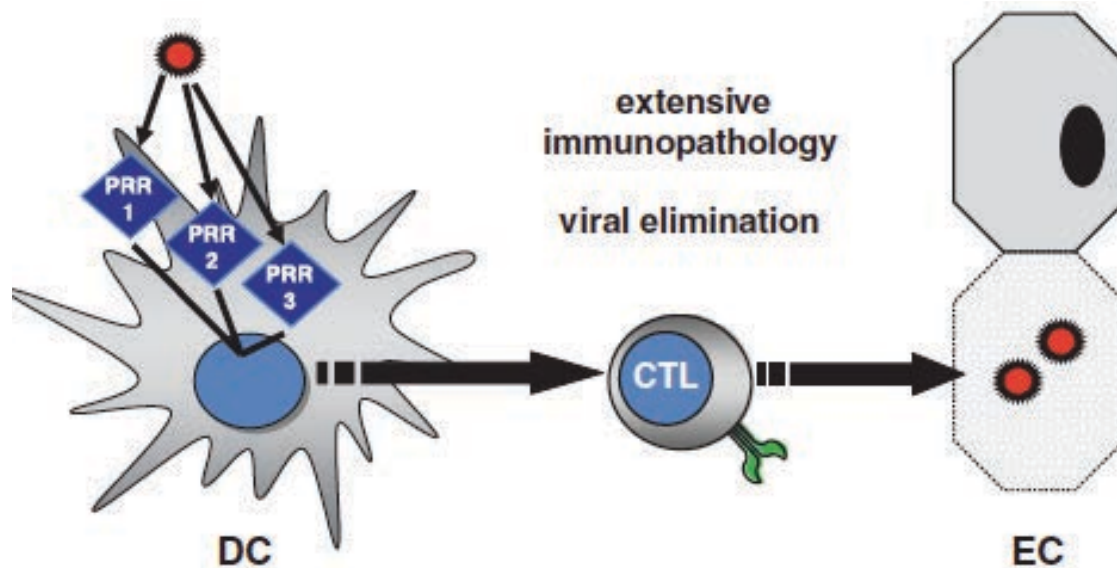
In humans

No replication of hantaviruses

Pulmonary / renal syndroms due to immunopathologies

Higher than normal numbers of interleukin (IL), tumor necrosis factor (TNF), interferons (INF), and activated Tc.

Hantavirus pathogenesis is a result of the host inflammatory response to the virus.



Schönrich et al 2018

In reservoirs

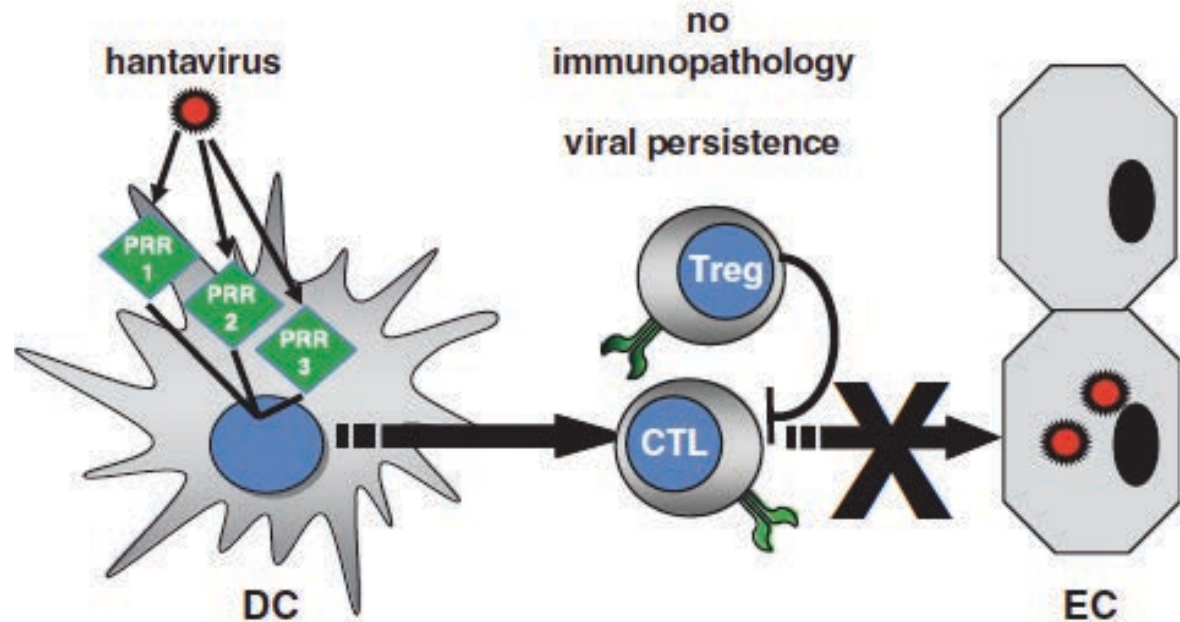


Long co-adaptation

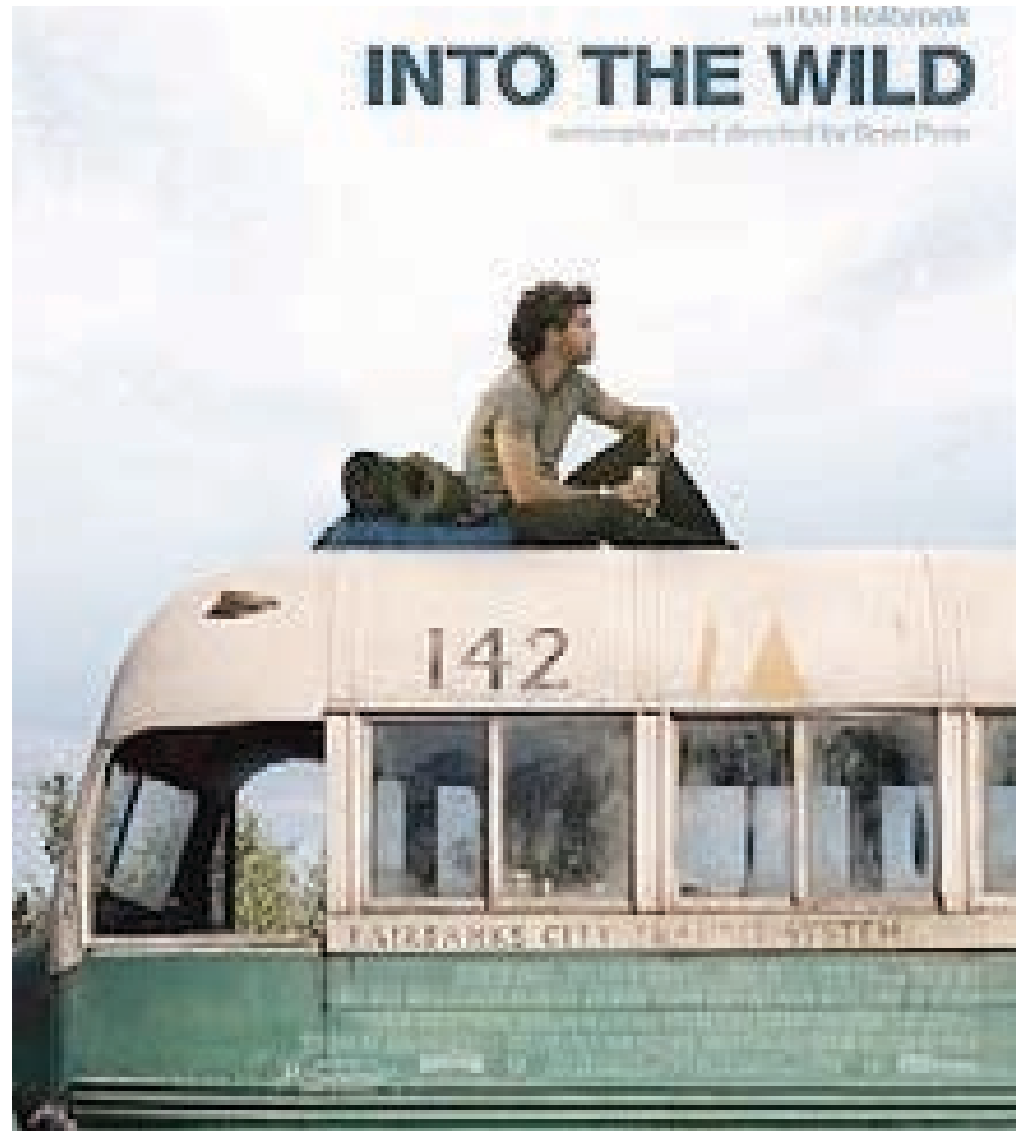


Tolerance to hantavirus ?

Chronic  
and  $\cong$  asymptomatic infection



# THE IMMUNE PERSPECTIVES ...





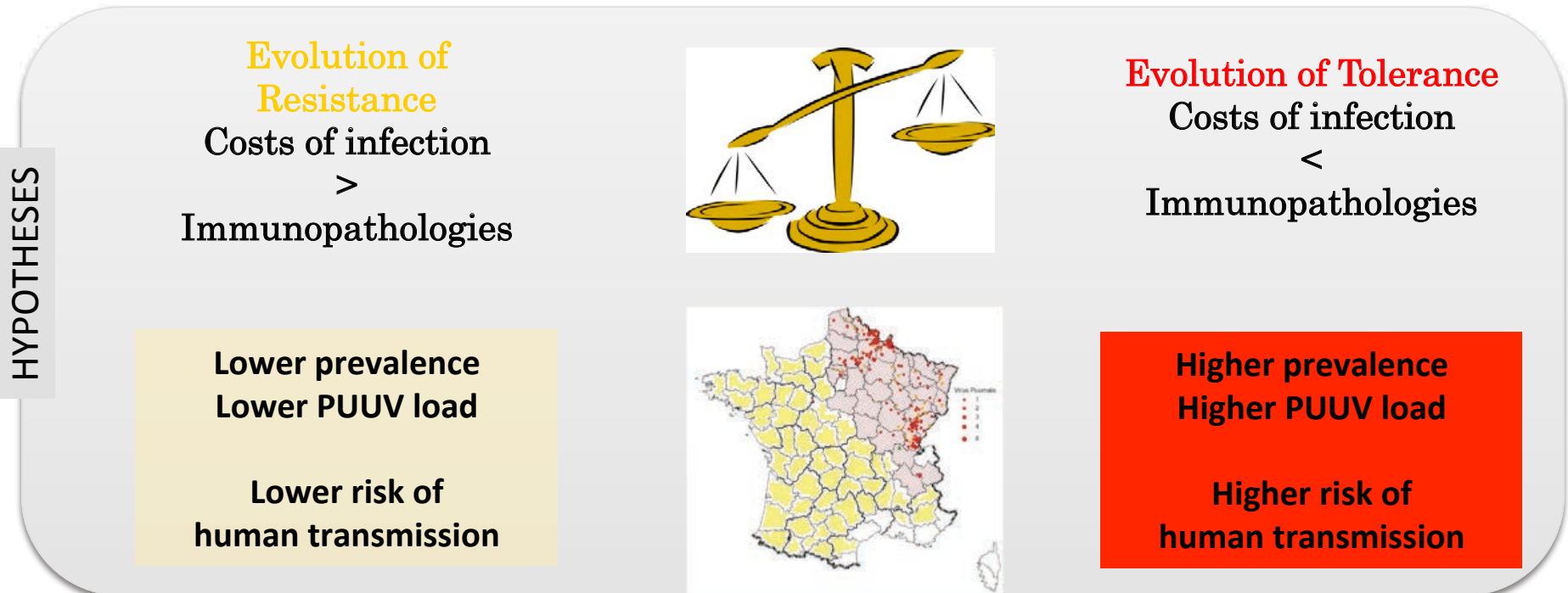
# THE IMMUNE PERSPECTIVES ...

Spatial variability in bank vole responses (tolerance) to PUUV ?  
Immunology

Signature of this variability in bank vole genomes ?  
Immunogenetics and genomics

Influence of virus genetic variability or pathobiome on bank vole responses ?  
Epidemiology

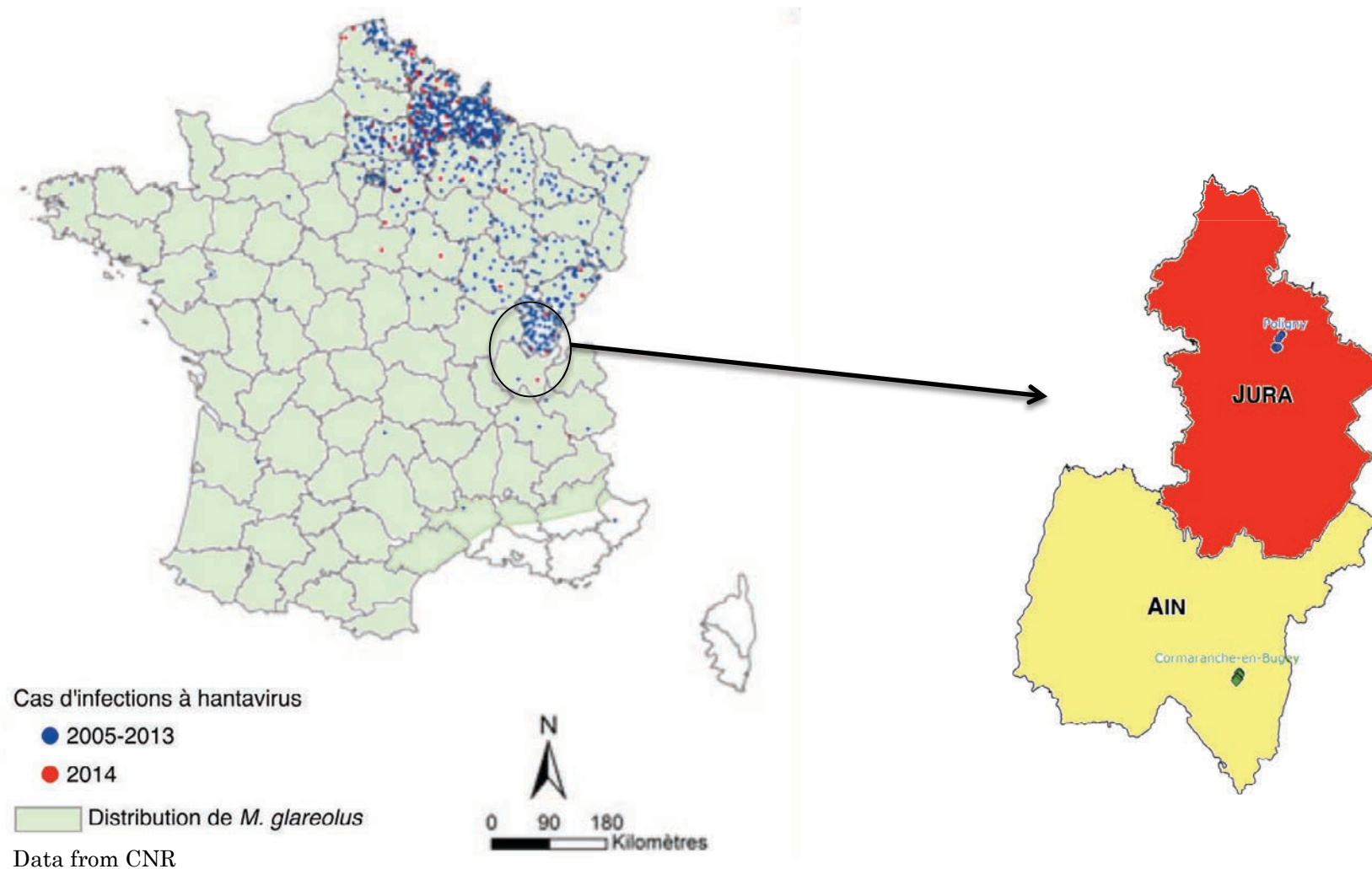
Links with  
PUUV distribution ?



A teal-colored brushstroke graphic with irregular, feathered edges, centered on a white background. The text is written in a bold, black, serif font within this graphic.

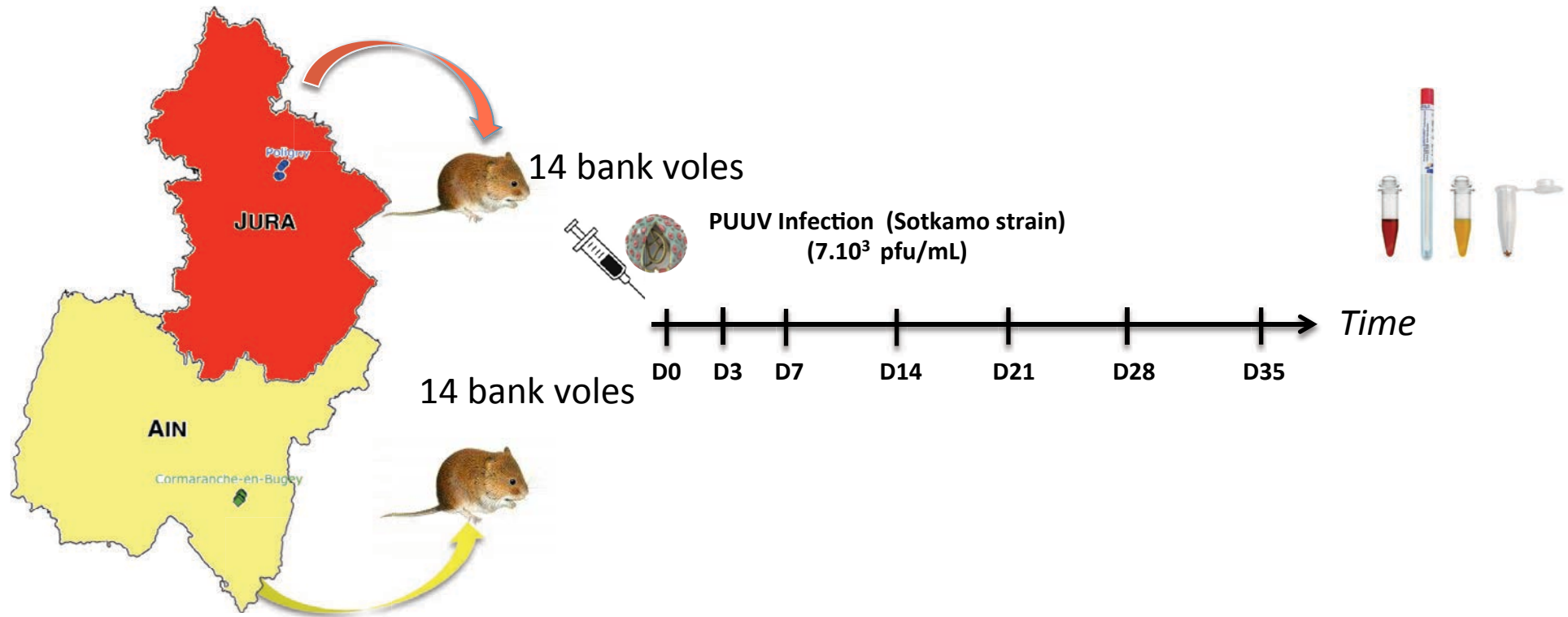
# **IMMUNOLOGICAL APPROACHES**

# COMPARISON OF BANK VOLE IMMUNE RESPONSES BETWEEN PUUV ENDEMIC AND NON ENDEMIC AREAS





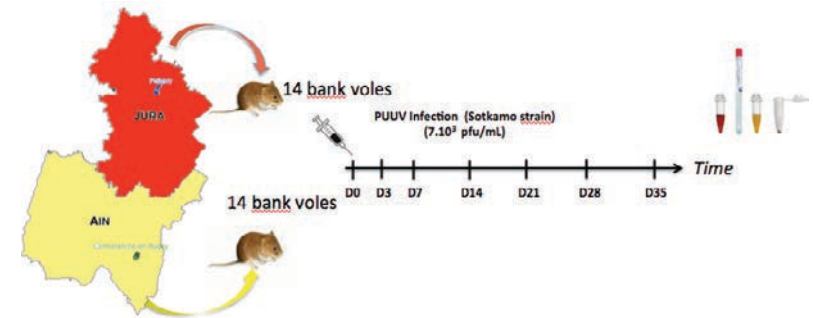
# EXPERIMENTAL INFECTIONS



Collab. ANSES Lyon

# EXPERIMENTAL INFECTIONS

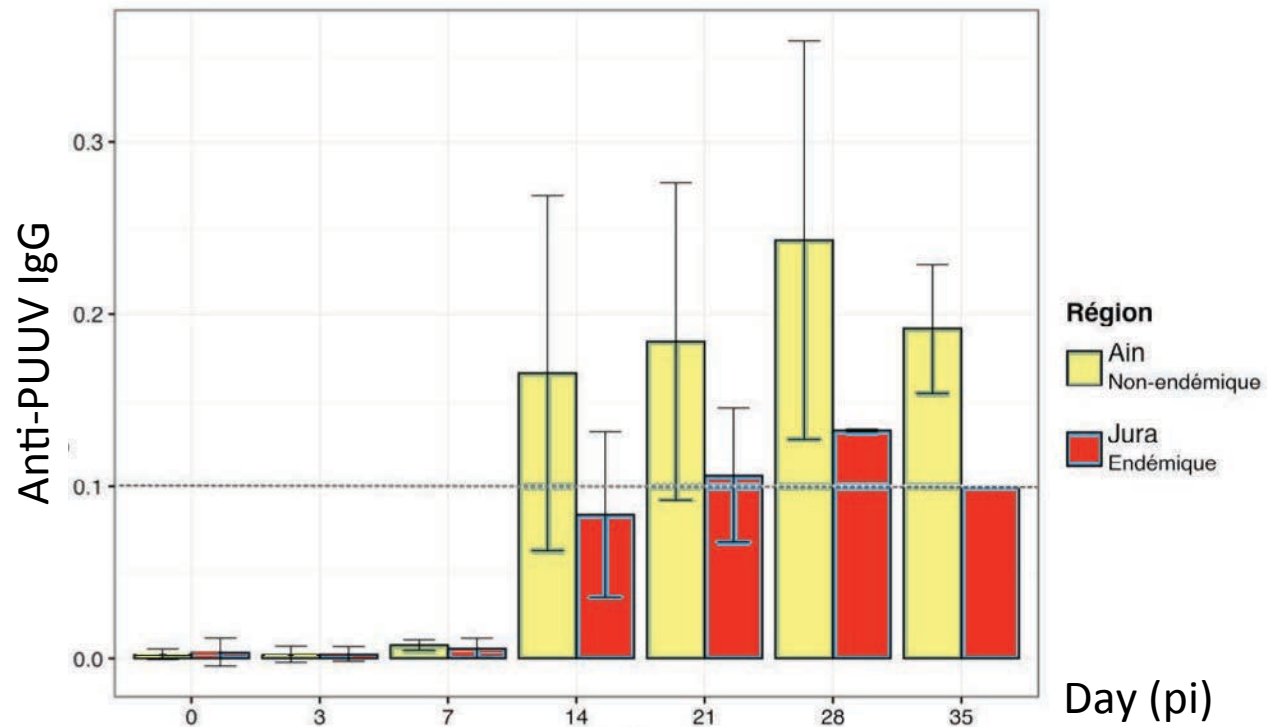
## 1- ANTIBODY PRODUCTION



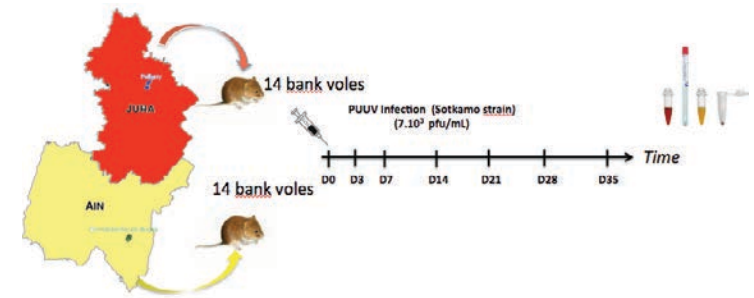
Inter-individual variability

All voles seroconverted  
-> Bank voles from the non endemic area are sensitive to PUUV

Higher & earlier production of antibodies in the non endemic region



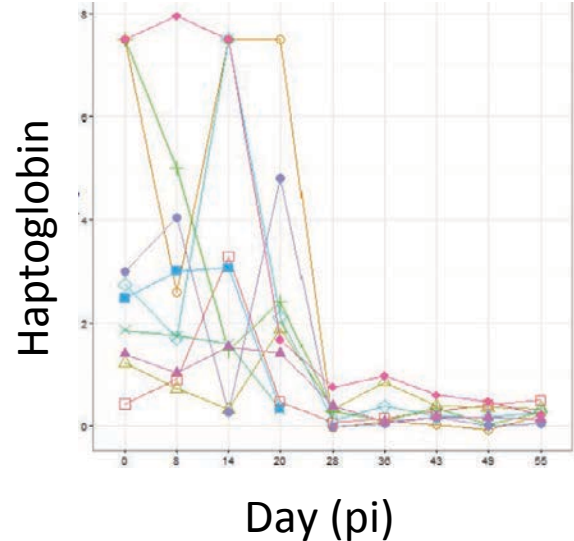
## 2- INFLAMMATORY RESPONSE



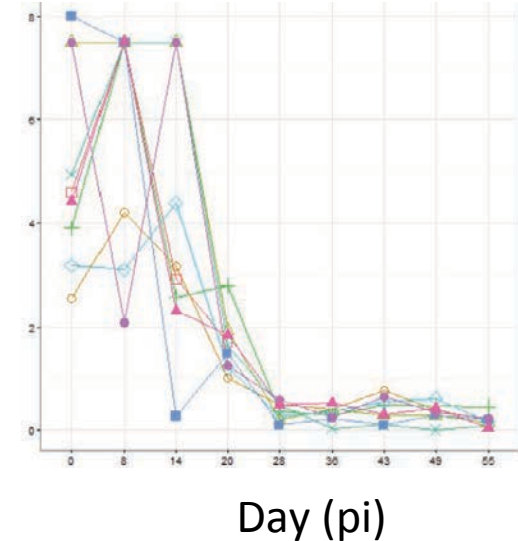
High inter-individual variability of the inflammatory response

Longer inflammatory response of voles from Ain

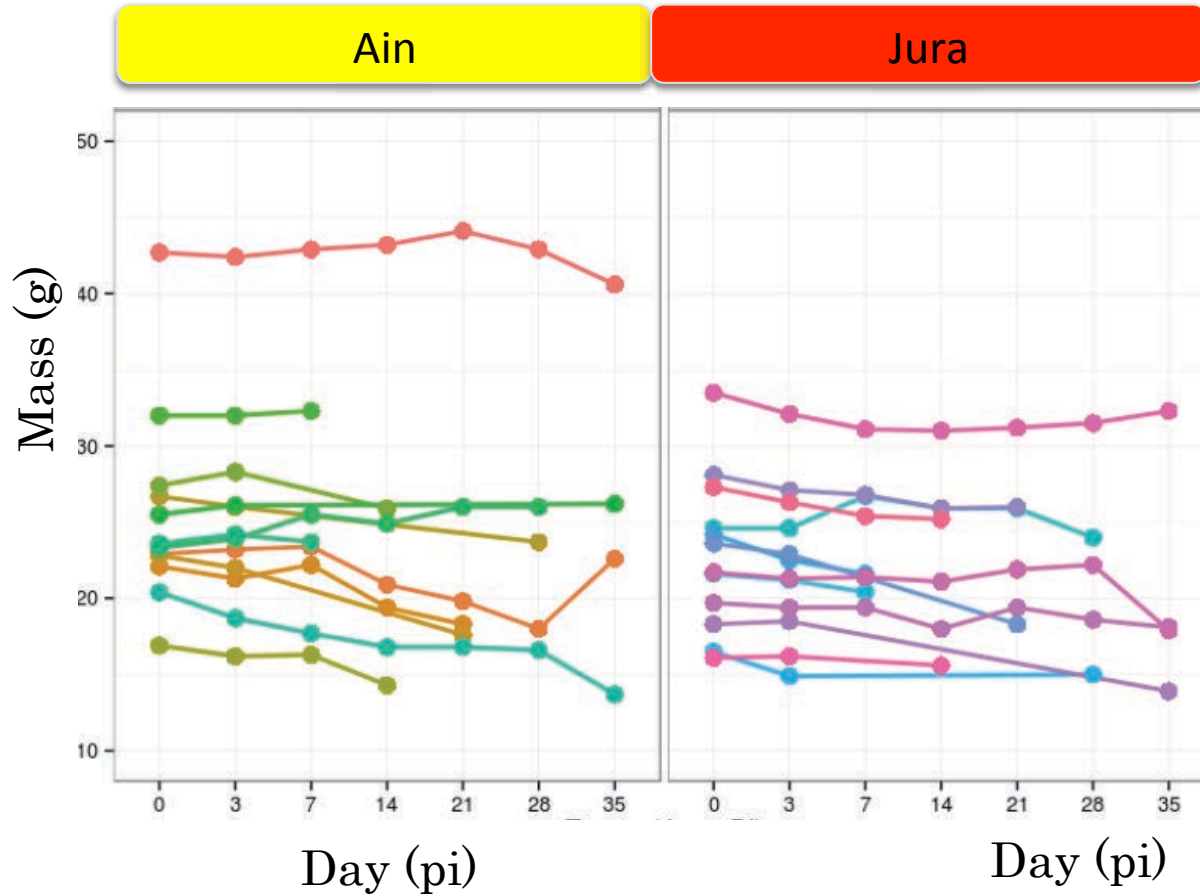
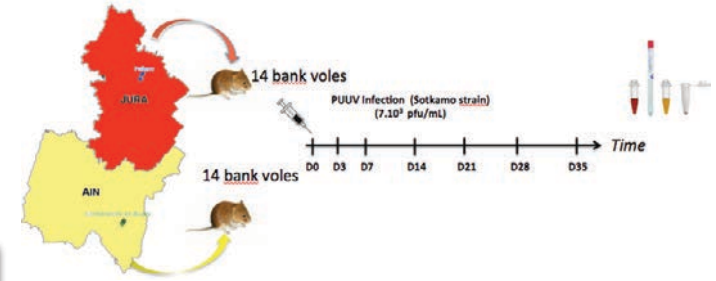
Ain



Jura



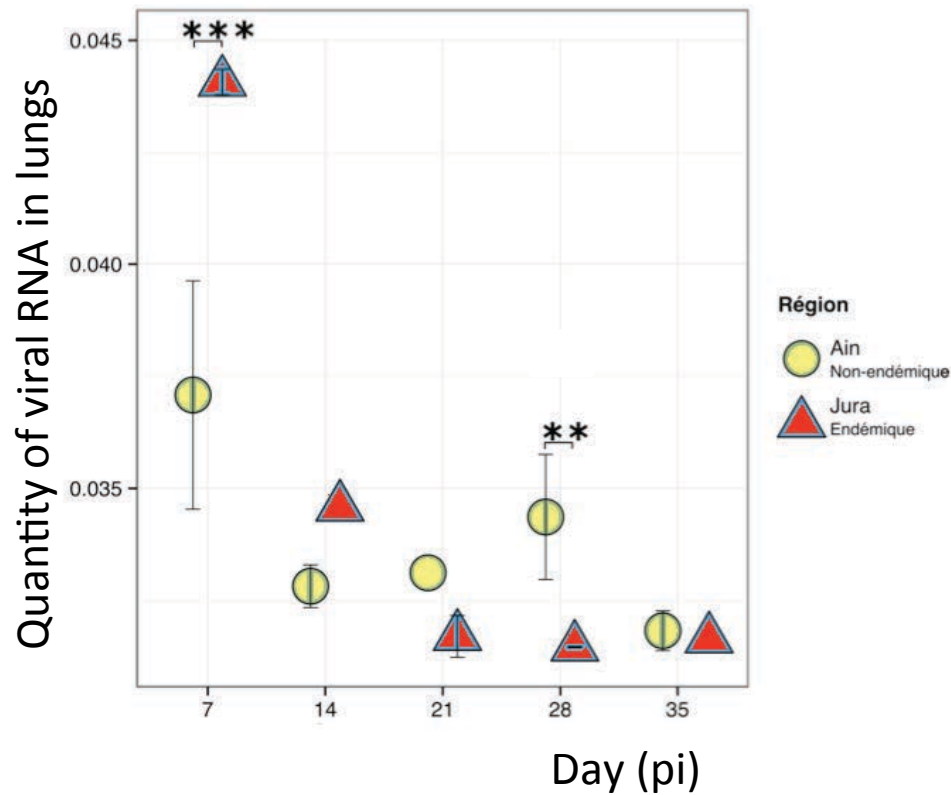
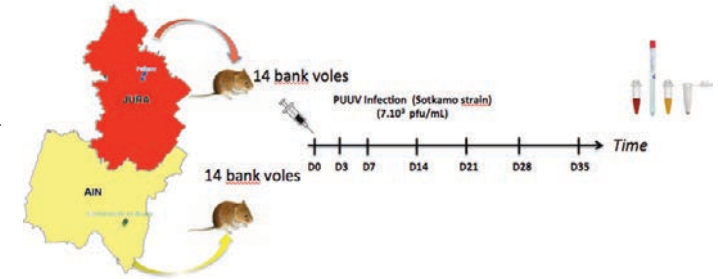
### 3- DIFFICULTY TO ASSESS TOLERANCE ...



No changes of temperature  
or body mass  
No evidence of tolerance  
with these parameters



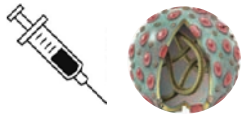
## 4- IMPACT OF BANK VOLE IMMUNE RESPONSE ON PUUV REPLICATION



High inter-individual variability of PUUV viral load detected in target organs

Higher replication of PUUV for infected bank voles of the endemic region

Differences in Resistance / Tolerance ?



# PUUV INFECTION WITH A WILD PUUV FRENCH STRAIN (ARDENNES)

## Ardennes

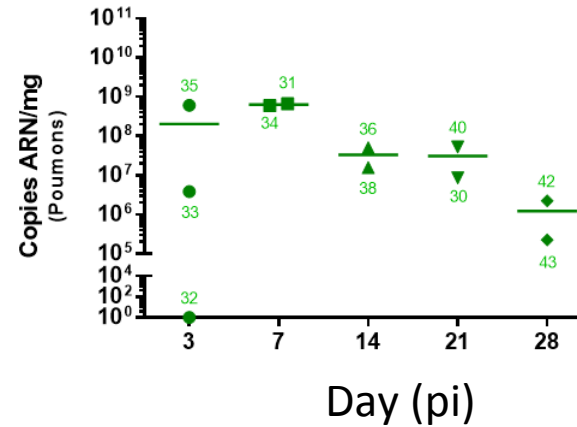
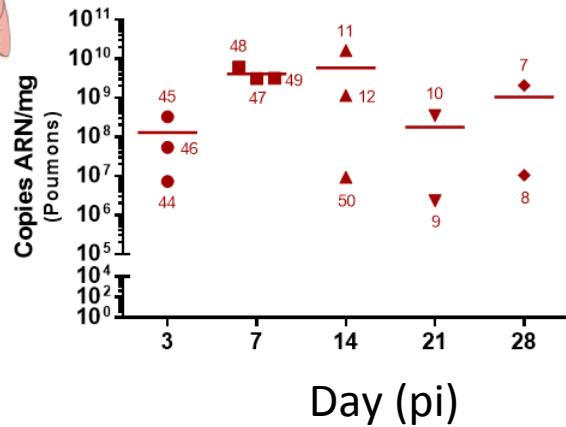
Endemic region

## Loiret

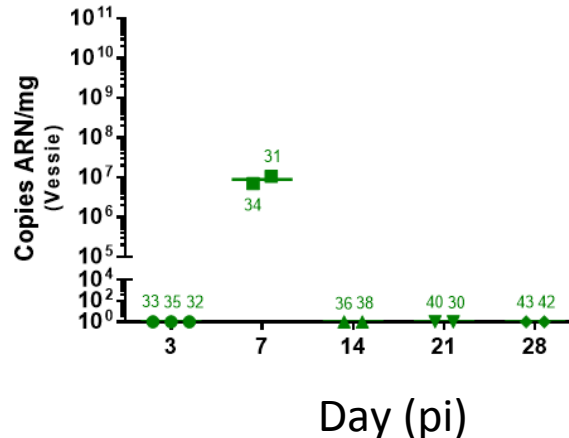
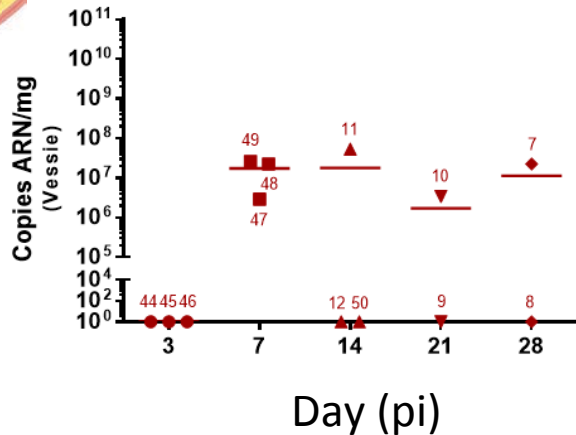
Peri endemic region



Lung  
Quantity of viral RNA



Bladder  
Quantity of viral RNA

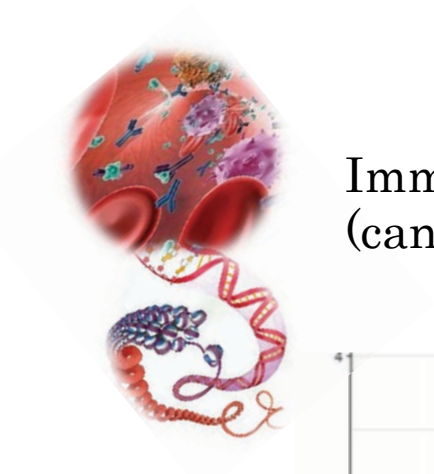


Higher and longer rate of PUUV replication for bank voles of the endemic region

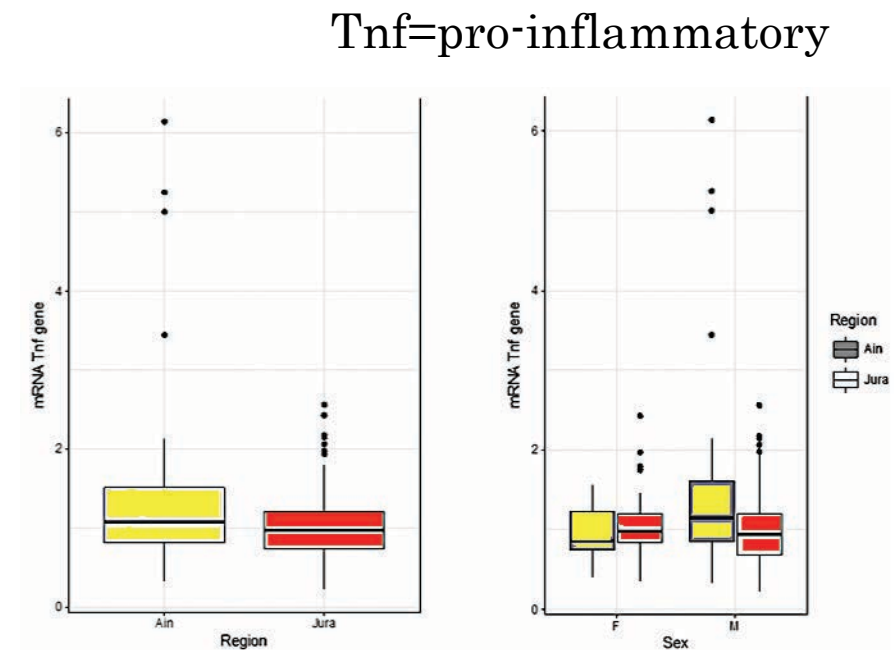
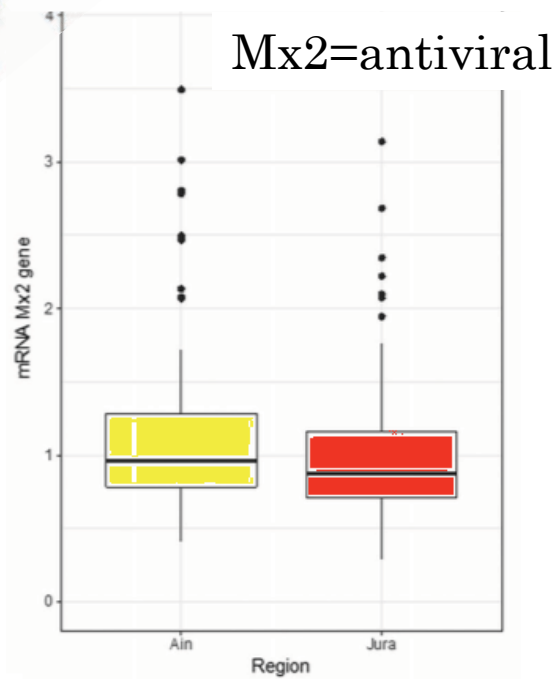
Lower Resistance ?  
Super-spreaders ?

Local adaptation of PUUV ?  
Cross-infection in progress

# VARIABILITY OF IMMUNE RESPONSES IN NATURAL POPULATIONS

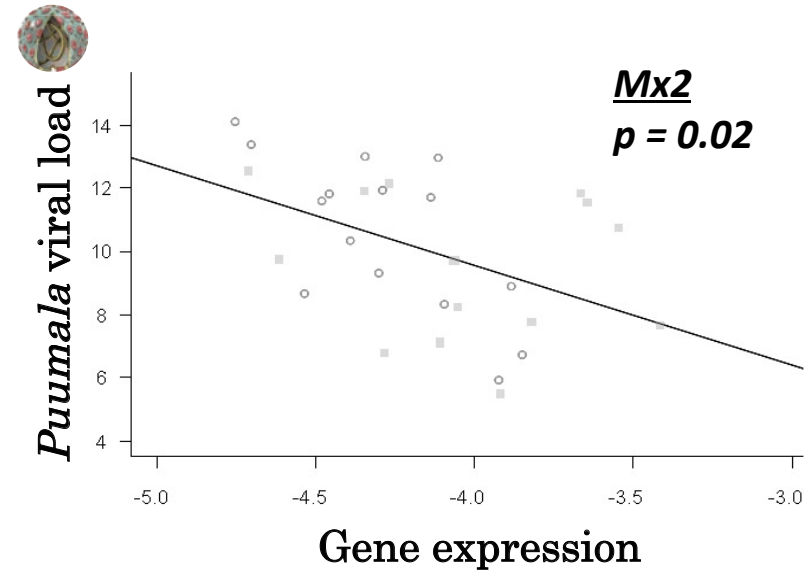
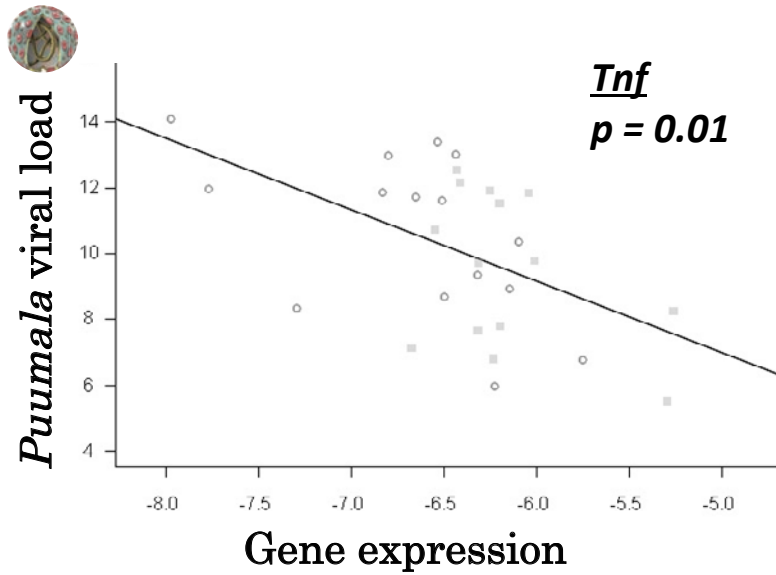
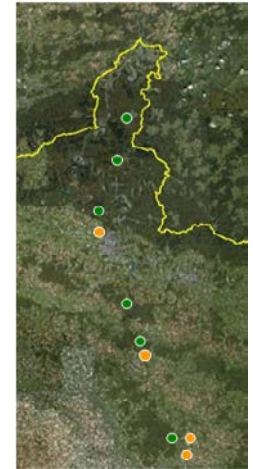


Immune phenotype = Immune gene expression  
(candidate genes chosen from the literature)



Higher immune gene expression in the non endemic compared to the endemic region

# French Ardennes Study



Strong negative relationships  
between immune gene expression (inflammation, antiviral)  
and PUUV replication

Tolerance



Resistance



## CONCLUSIONS FROM THE IMMUNOLOGICAL APPROACHES

Bank voles from non endemic areas are sensitive to infections with PUUV lab or wild strains

There is a high inter-individual variability in the response to PUUV


There is a slight inter-population variability in the response to PUUV

- Higher antibodies and longer inflammation in bank voles from non endemic area
- Weaker replication of PUUV in bank voles from non endemic area  
Higher **resistance** of bank voles from non endemic areas ?  
Higher **tolerance** of bank voles from endemic areas ? Super-spreaders ?

There is a risk for Hantavirus disease emergence  
in non endemic areas adjacent to endemic ones.

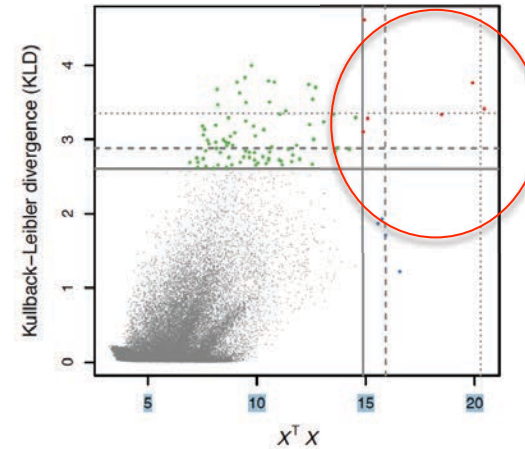
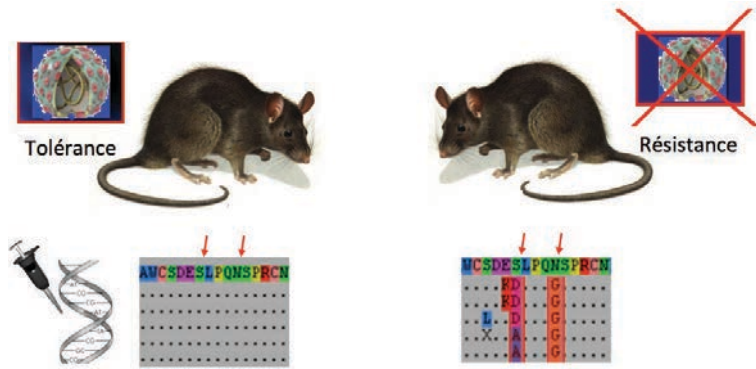
The risk might be limited by a strong immune response against PUUV,  
leading to its rapid elimination in bank voles,  
what should limit its persistence in these reservoir populations

Can tolerance to PUUV evolve in these non endemic areas adjacent to endemic ones ?

A teal-colored brushstroke graphic with irregular, feathered edges, centered on a white background. The text is written in a bold, black, serif font within this graphic.

**IMMUNOGENETICS  
& GENOMICS  
APPROACHES**

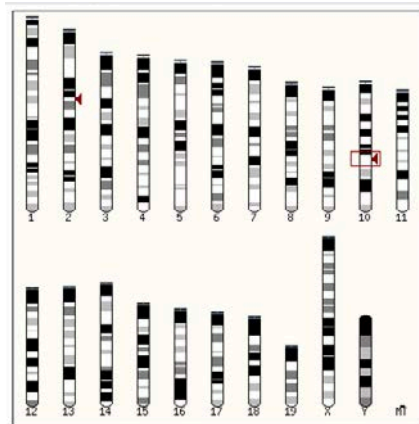
# PRINCIPLES OF ADAPTATION GENOMICS



Identification of 'outliers' loci

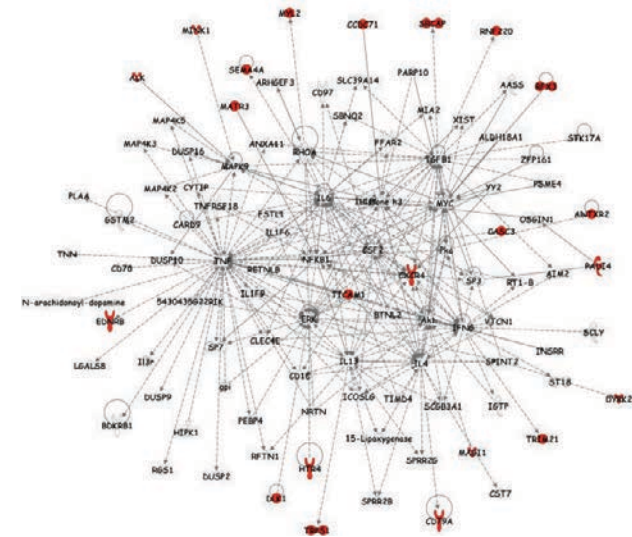
Higher genetic differentiation than expected under neutral models

Genome scan :  
Identification of genes  
evolving under positive directional  
selection between populations

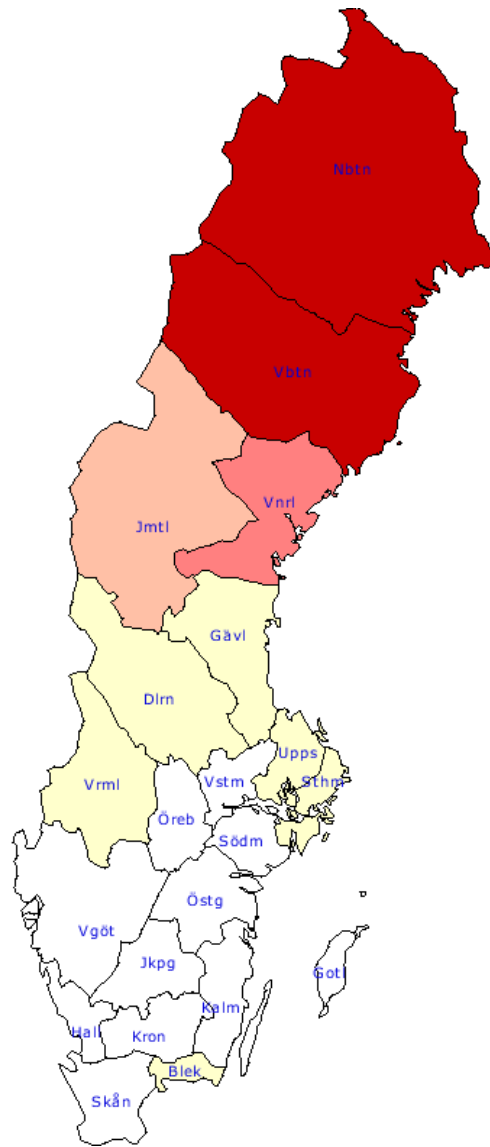


Annotation of outliers  
Gene ontology

Enrichment analyses  
Network analyses



# SPATIAL DISTRIBUTION OF NEPHROPATHIA EPIDEMICA IN SWEDEN



## Sweden

- High presence of human cases
- Low presence of human cases
- Absence of human cases

Human socio-economics

Probability of human exposure to NE

Environmental variables

PUUV persistence in the environment

Bank vole population dynamics

Bank vole population heterogeneity

Immunity, genomics







## Gene candidate approach

based on the medical literature

Tnf, Mx2, TLR4, TLR7 (SNP genotyping, Kaspar technology)

but restrictive approach (you find only what you search)

## Whole genome approach

Blind, no a priori - but *Myodes glareolus* has a large genome (3Gb)

Need for a reduced representation sequencing : RAD-sequencing



Enzyme : SbfI = 8 cutter

Pools of 35/40 individuals

6 pops, 4 replicates (Mid tags 5 & 6 bases, 2 libraries)

Paired-end sequencing, 2x100 bp

70,000 contigs, 500 bp

95,000 SNPs (> 40 reads, <5% singleton)



# DETECTION OF SIGNATURE OF SELECTION

## 1- Population Genomic Differentiation

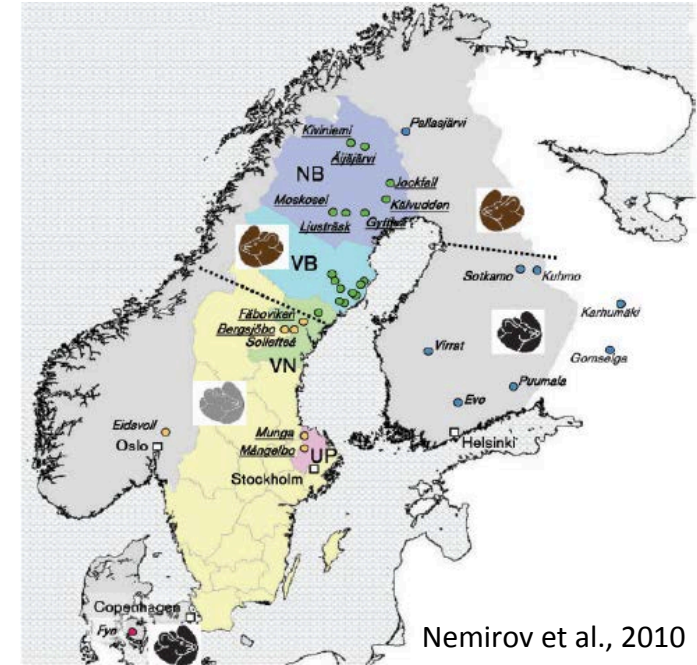
SelEstim

### Detecting and Measuring Selection from Gene Frequency Data

Renaud Vitalis,<sup>\*,1,†</sup> Mathieu Gautier,<sup>\*,†</sup> Kevin J. Dawson,<sup>‡</sup> and Mark A. Beaumont<sup>§</sup>

<sup>\*</sup>Institut National de la Recherche Agronomique, Unité Mixte de Recherche CBGP, (Inra, Ird, Cirad, Montpellier-SupAgro) 34988 Montpellier-sur-Lez Cedex, France, <sup>†</sup>Institut de Biologie Computationnelle, 34095 Montpellier Cedex, France, <sup>‡</sup>Cancer Genome Project, The Wellcome Trust Sanger Institute, Hinxton, CB10 1SA, United Kingdom, <sup>§</sup>Department of Mathematics and School of Biological Sciences, University of Bristol, Bristol BS8 1TW, United Kingdom

Genetics, Vol. 196, 799–817 March 2014



Nemirov et al., 2010

High  $F_{st}$  estimates,  
Hierarchical structure

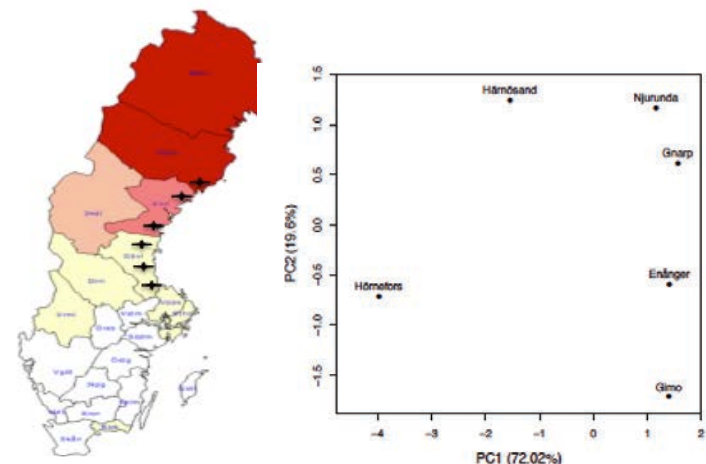
BayPass

### Genome-Wide Scan for Adaptive Divergence and Association with Population-Specific Covariates

Mathieu Gautier<sup>\*,§,1</sup>

<sup>\*</sup>INRA, UMR CBGP (INRA – IRD – Cirad – Montpellier SupAgro), Campus international de Baillarguet, CS 30016, F-34988 Montpellier-sur-Lez, France, <sup>§</sup>Institut de Biologie Computationnelle, 95 rue de la Galera, 34095 Montpellier, France

Genetics, Vol. XXX, XXXX–XXXX September 2015



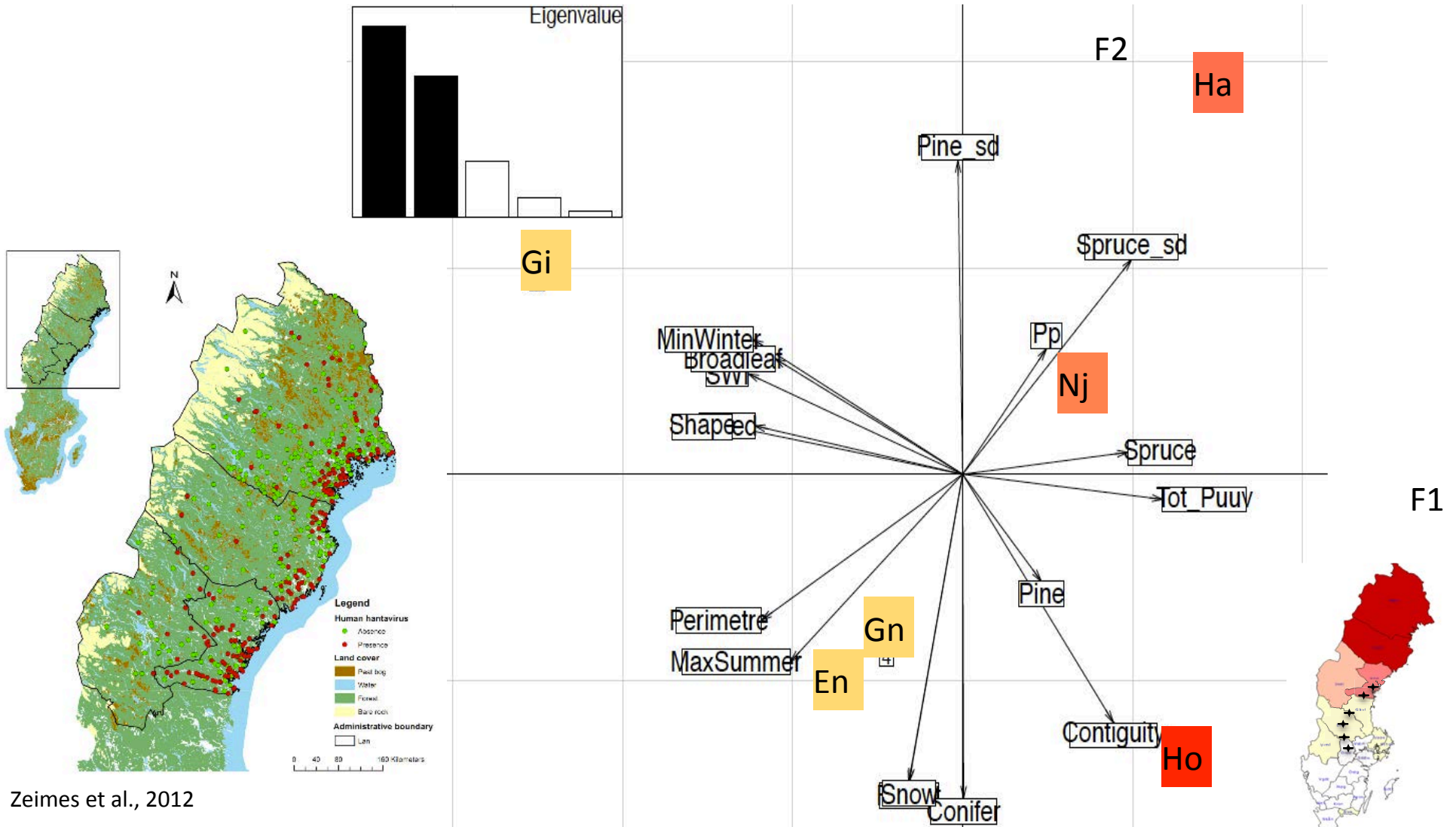
## 2- Environmental & climatic variables

## Genome-Wide Scan for Adaptive Divergence and Association with Population-Specific Covariates

Mathieu Gautier<sup>\*S,1</sup>

<sup>\*</sup>INRA, UMR CBGP (INRA – IRD – Cirad – Montpellier SupAgro), Campus international de Baillarguet, CS 30016, F-34988 Montpellier-sur-Lez, France, <sup>§</sup>Institut de Biologie Computationnelle, 95 rue de la Galera, 34095 Montpellier, France

### BayPass

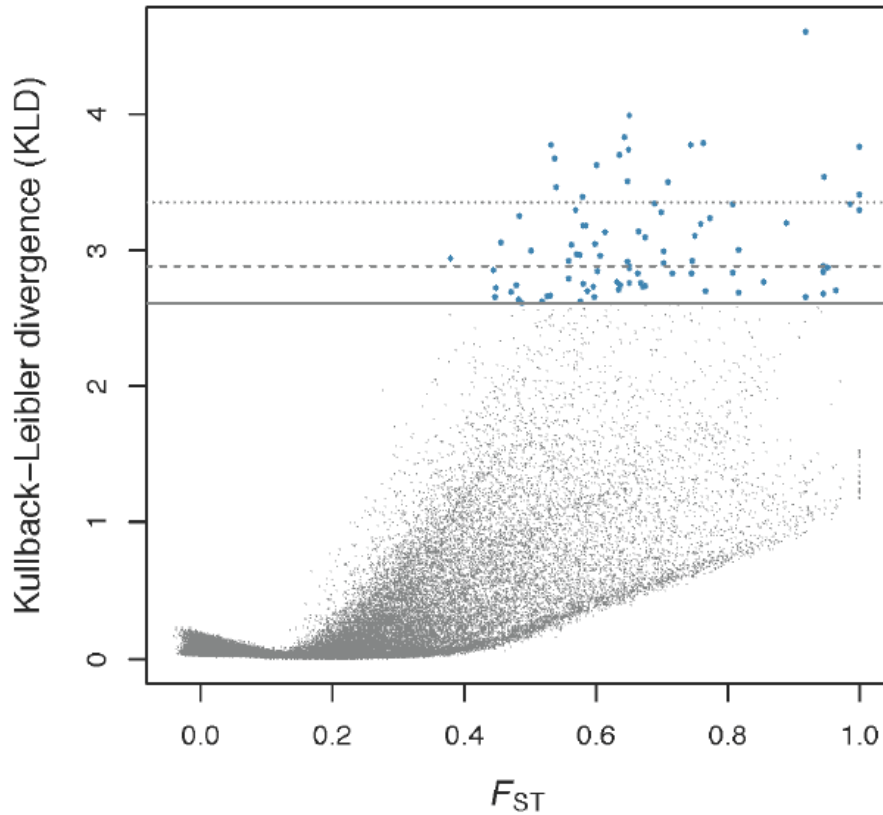


Zeimes et al., 2012



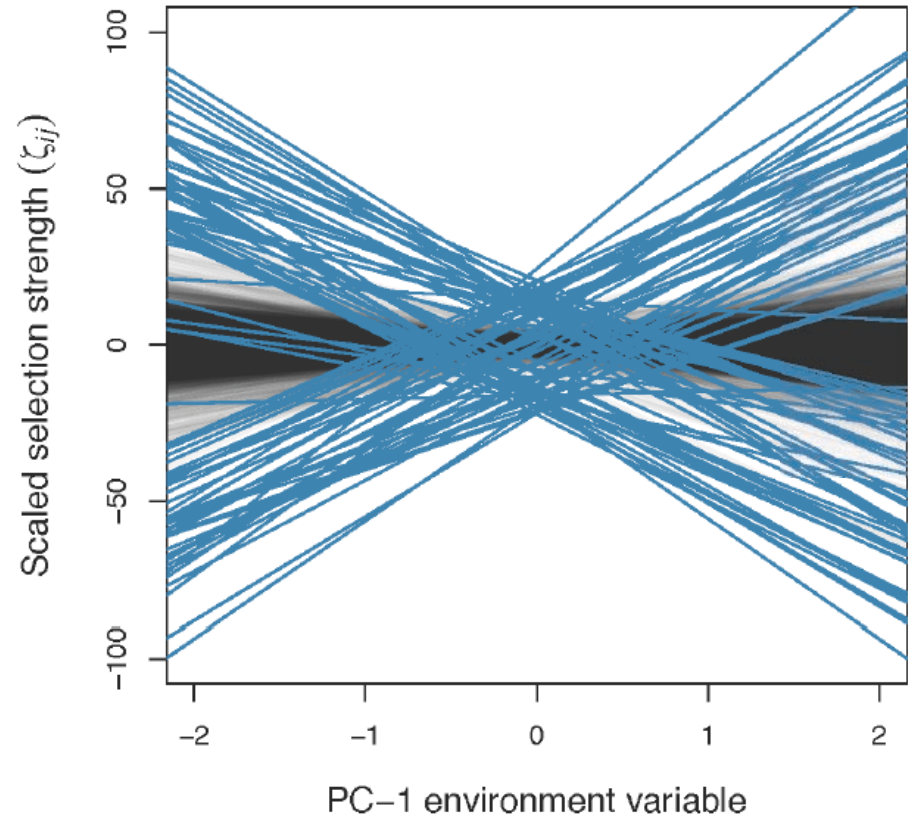
# RESULTS

## 1.a- Genomic Differentiation, SelEstim



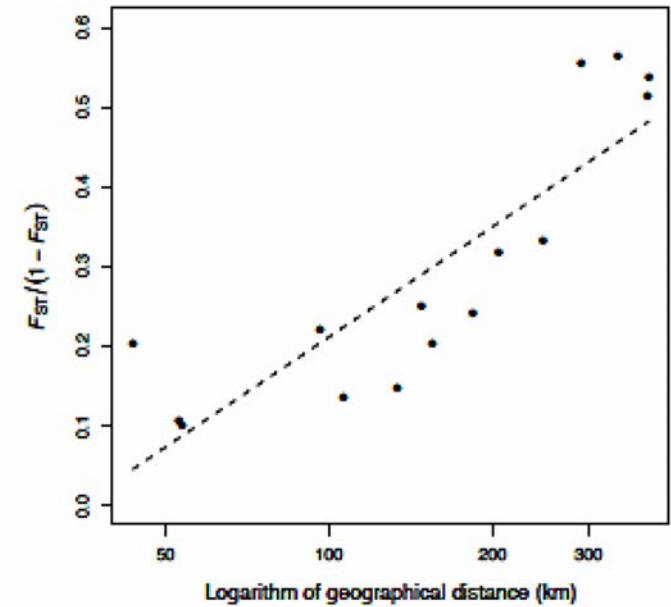
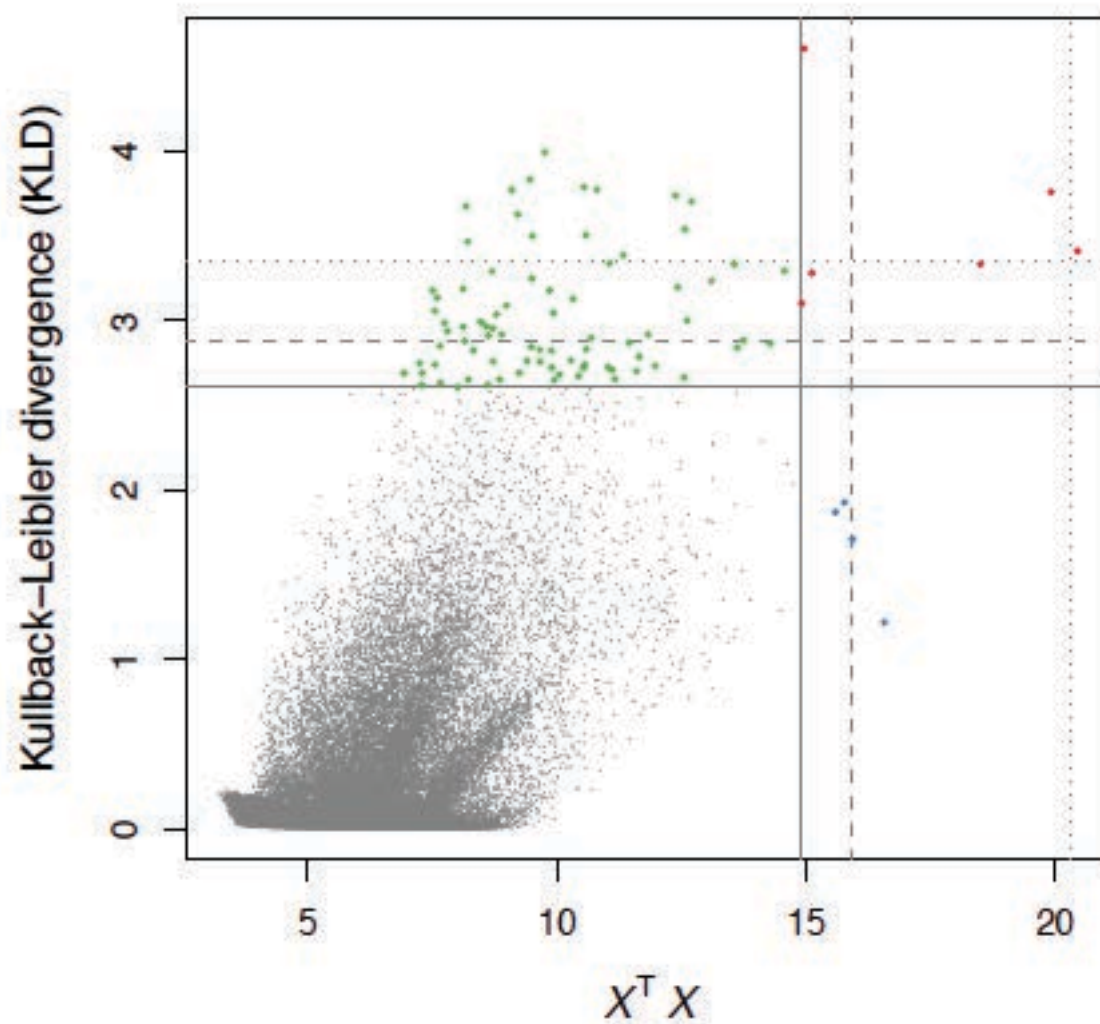
Threshold 99.9%

86 outliers detected  
Including the candidate *Tlr7*



Strong correlation between  
local selection coefficients and  
environmental variables associated  
with PUUV

## 1.b- Genomic Differentiation, BayPass

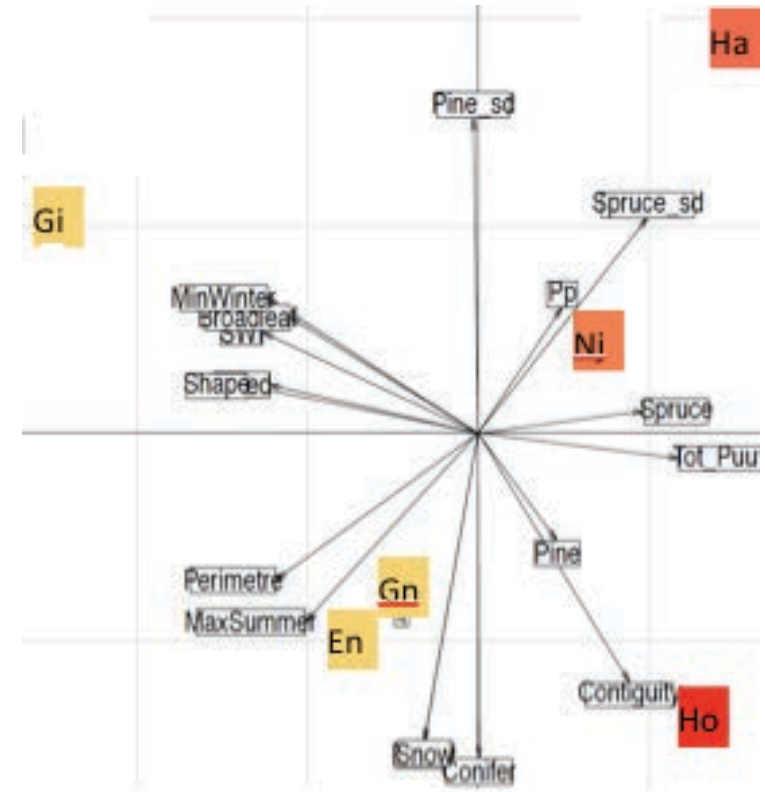
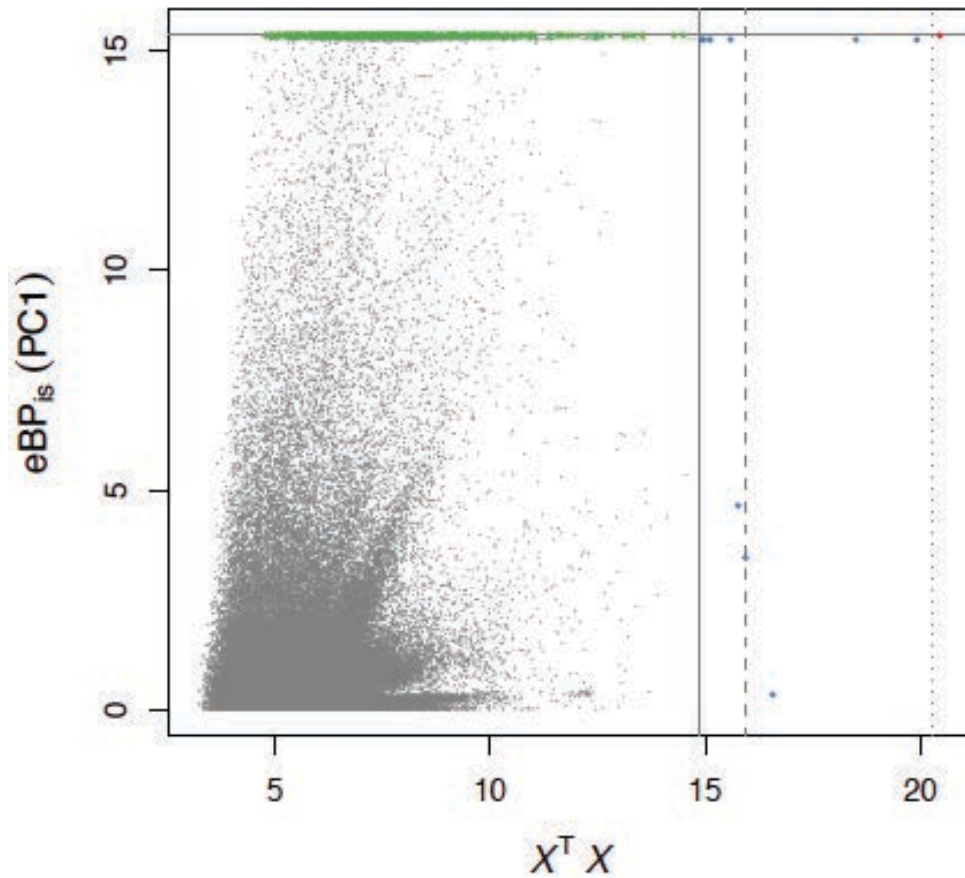


Strong isolation  
by distance pattern

Threshold 99.9%

10 outliers detected,  
6 are common with SelEstim

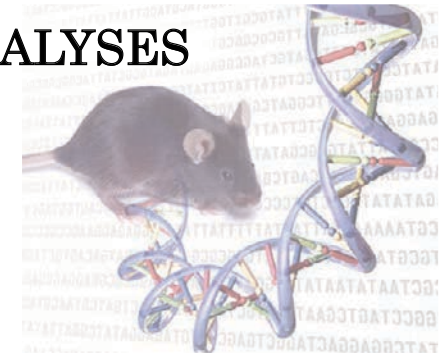
## 2. Environmental covariates, BayPass



Threshold 99.9%

395 outliers detected with PC1,  
26 are common with SelEstim or  
BayPass

# ANNOTATIONS OF OUTLIERS AND GENE ENRICHMENT ANALYSES



Among the 70.000 contigs and using  
*M. musculus genome*

44 % are annotated using BlastN  
21 % are annotated using BlastX  
14 % matched with coding sequences  
of proteins

22 chromosomes are covered

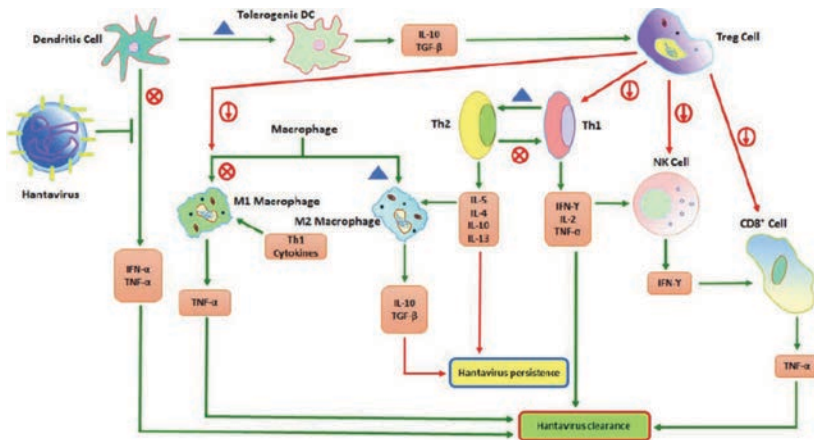
Annotation of outliers

468 RAD – contigs

108 outliers – including *Tlr7* – matched to  
104 mouse protein-coding genes

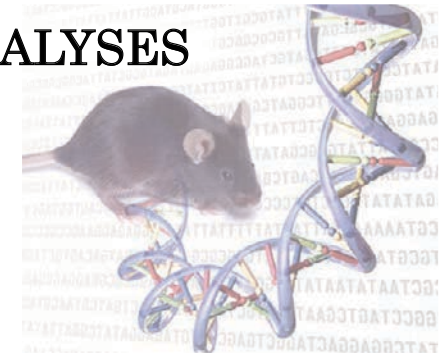
GO annotations for 95 of them

14 genes had GO related with immunity





# ANNOTATIONS OF OUTLIERS AND GENE ENRICHMENT ANALYSES

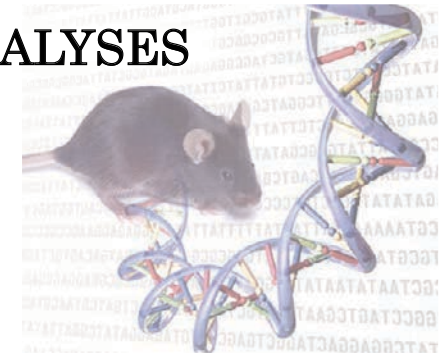


The third most represented biological process was the **‘positive regulation of cytokine production’**

primary metabolic process	cellular metabolic process	protein metabolic process	phosphorus metabolic process	regulation of cellular biosynthetic process	macromolecule biosynthetic process	regulation of biosynthetic process	regulation of cellular process	cellular response to stimulus	negative regulation of biological process	signal transduction	positive regulation of multicellular organismal process
organic substance metabolic process	macromolecule metabolic process	macromolecule modification	regulation of metabolic process	cellular macromolecule biosynthetic process	carbohydrate derivative metabolic process	organic cyclic compound biosynthetic process	regulation of biological process	regulation of response	intracellular signal	regulation of cell communication	regulation of signaling
	cellular macromolecule metabolic process	cellular biosynthetic process	cellular glycoprotein metabolic process	glycoprotein metabolic process	nitrogen-containing compound biosynthetic process	cellular nitrogen compound biosynthetic process		positive regulation of biological process	response to stress	cell surface receptor signaling pathway	regulation of multicellular organismal process
nitrogen compound metabolic process	organonitrogen compound metabolic process	biosynthetic process	heterocycle biosynthetic process	organic cyclic compound metabolic process	regulation of transcription from RNA polymerase II promoter	transcription from RNA polymerase II promoter	cellular protein modification process	regulation of biological quality	response to chemical	memory	behavioral fear response
	aromatic compound biosynthetic process	cellular nitrogen compound metabolic process	heterocycle metabolic process	gene expression	RNA metabolic process	RNA metabolic process		regulation of cellular metabolic process	regulation of nitrogen compound metabolic process	negative regulation of cellular process	
anatomical structure development	cell differentiation	multicellular organism development	cellular component organization	nervous system development	positive regulation of RNA metabolic process	hematopoietic or lymphoid organ development	phosphate-containing compound metabolic process	peptidyl-serine autophosphorylation	regulation of primary metabolic process	regulation of nucleobase-containing	regulation of name
	cellular developmental process	cell development	positive regulation of biosynthetic process	cellular component morphogenesis	positive regulation of developmental process	immune system development		cellular protein metabolic process	regulation of macromolecule metabolic process	negative regulation of metabolic process	peptidyl-amino acid modification
positive regulation of cellular process	anatomical structure morphogenesis	system development	animal organ development	positive regulation of signaling	cellular component assembly	tissue development	response to organic substance	response to oxygen-containing compound	response to mechanical stimulus	cellular response to organic substance	response to endogenous stimulus
	positive regulation of cellular biosynthetic process	positive regulation of metabolic process	positive regulation of gene expression	positive regulation of response to stimulus	glycerolipid metabolic process	bone development		response to abiotic stimulus	response to external stimulus	ethanol	cellular response to chemical stimulus



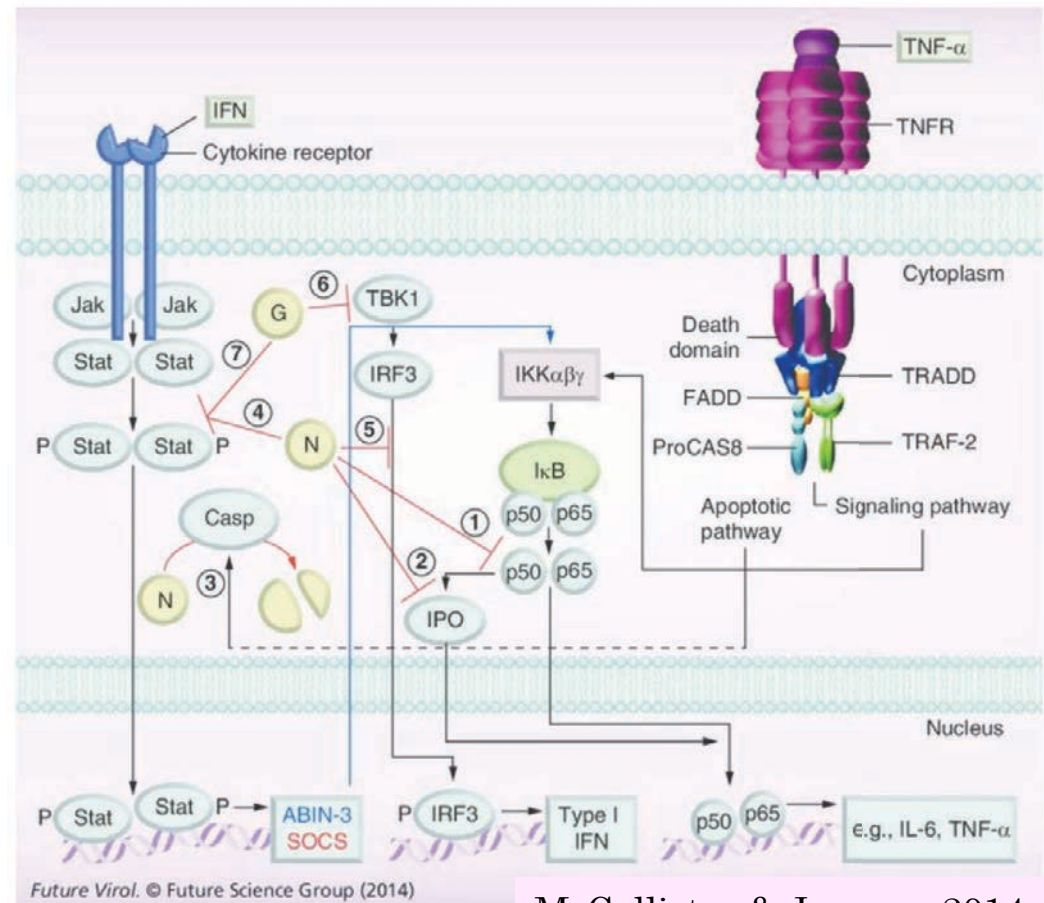
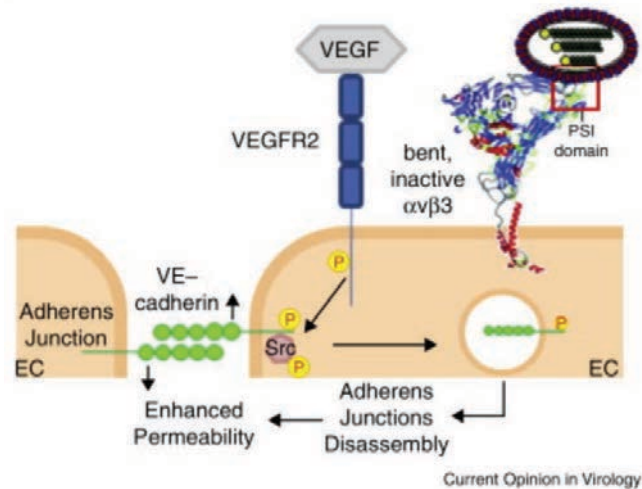
# ANNOTATIONS OF OUTLIERS AND GENE ENRICHMENT ANALYSES



125 pathways were identified,  
out of which 13 were significantly enriched

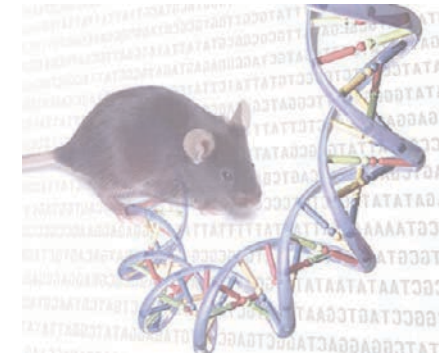
Toll-Like Receptors cascades  
JAK STAT pathway regulation  
IL-7 signaling

+ VEGF signalling pathway



McCallister & Jonson 2014

# CONCLUSIONS FROM THE GENOMICS APPROACHES



## Limits of the study

- Low number of populations
- High levels of genomic differentiation (contact zone)
- Potential confounding effects (correlation latitude and PUUV distr.)
  
- Lack of power to annotate outliers

## But

- Outliers showing congruent 'spatial' patterns of selection
- Candidate genes & expected genes with regard to immunity

## Perspectives

- Large spatio-temporal surveys of bank vole populations in Sweden
- Analyses of other european samplings (e.g. France, Germany)
- Improvement of bank vole genome / transcriptome annotation
- Collaborations with immunologists, clinicians to interpret outliers







# FROM HOST-PARASITE TO HOST PATHOBIOME

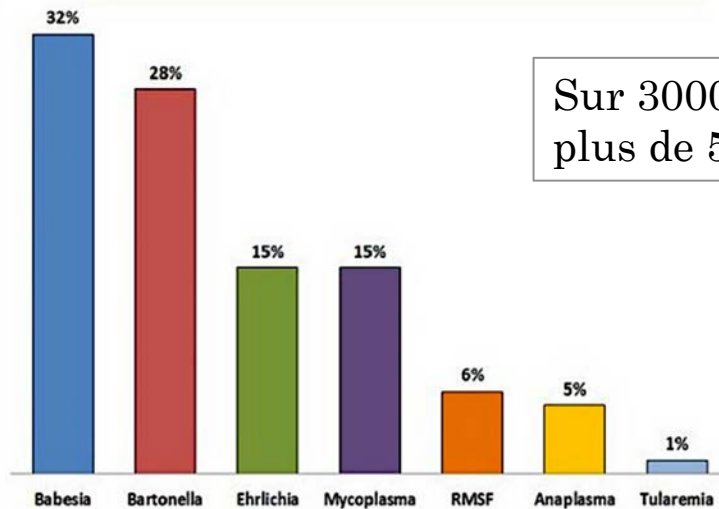






Exposition simultanée à de nombreux pathogènes = fréquent  
Influence épidémiologique (sensibilité, pathogénicité ...)

The Most Commonly Reported Co-Infections  
U.S. Patients with Lyme Disease



Sur 3000 patients diagnostiqués pour 'Lyme', plus de 50% ont des coinfections bactériennes

Microbiome = très diversifié, fort impact sur l'immunité



Changement de paradigme

1 hôte / 1 pathogène

Hôte / **Pathobiome** = pathogène et son environnement biotique

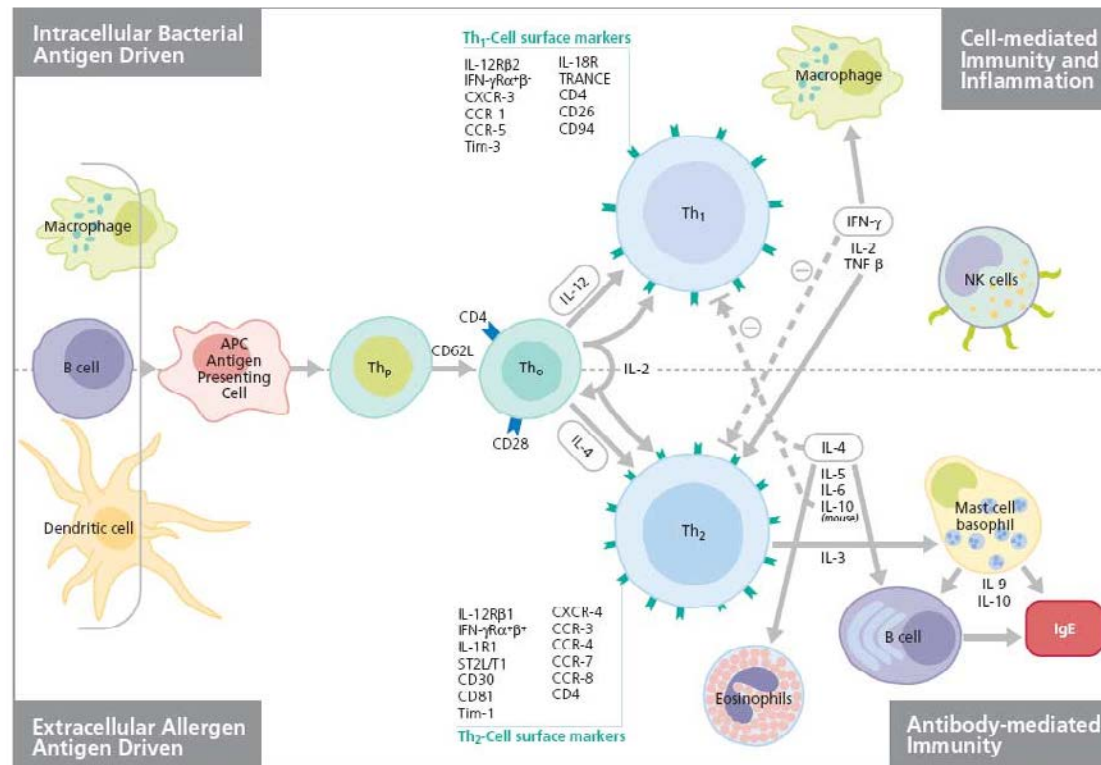




# CO-INFECTIONS BETWEEN PUUV AND HELMINTHS

Evolutionary or physiological trade-offs between immune responses

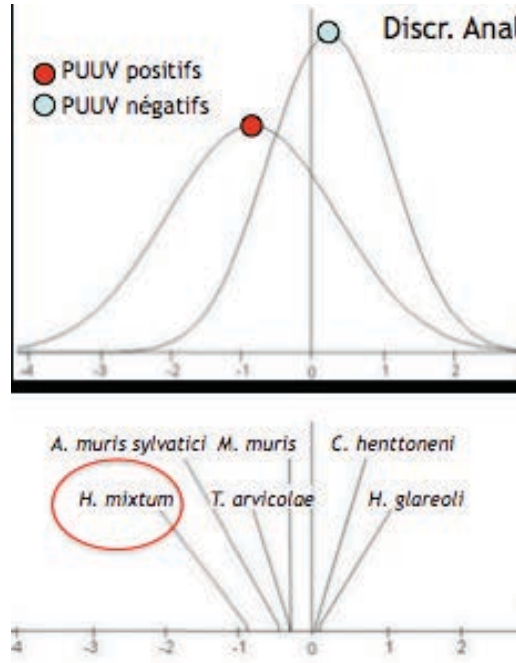
Ex : Antagonistic immune responses to intra & extra cellular pathogens (Th1/Th2)



Hyp : High Th1 (eg inflammation) = low risk of viral infection  
but high risk of helminth inf.



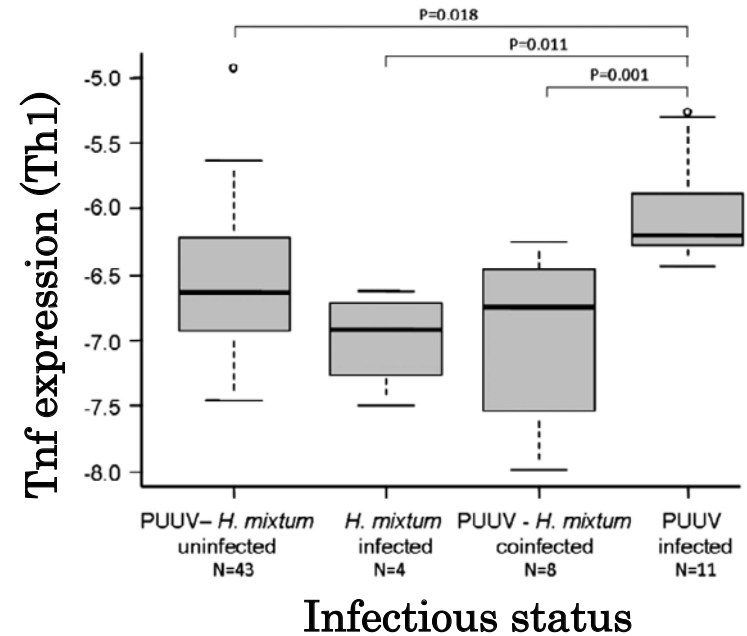
French Ardennes



Co-infections *H. mixtum* / PUUV

Co-infection increases *H. mixtum* load

Th1 could mediate these patterns



# INTERACTIONS BETWEEN PUUV THE BACTERIOME AND THE MYCOME



Methodological challenges in molecular biology, bioinformatics and biostatistics

Solved for bacteriome studies

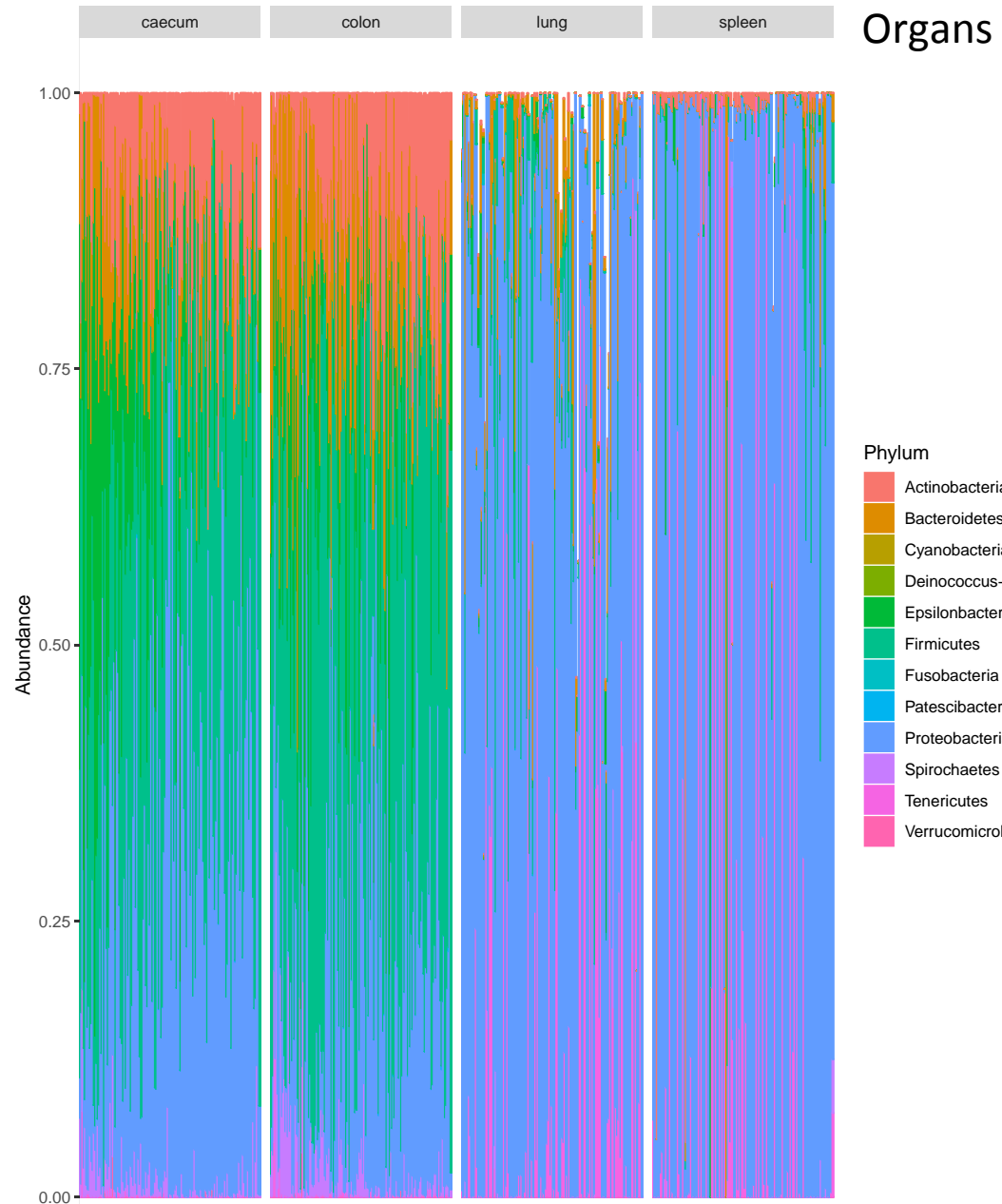
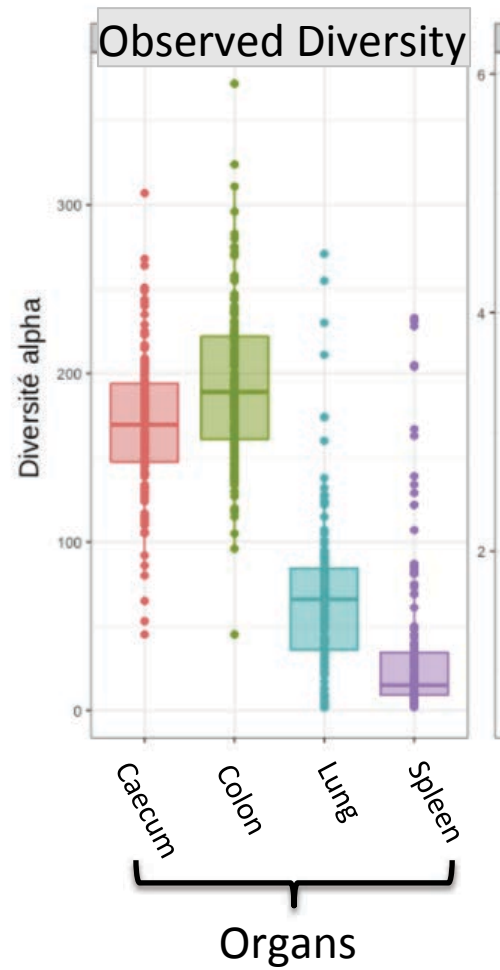
In progress for mycobiome study

(but see FrogsFungi : <https://github.com/geraldinepascal/FROGS/releases>)

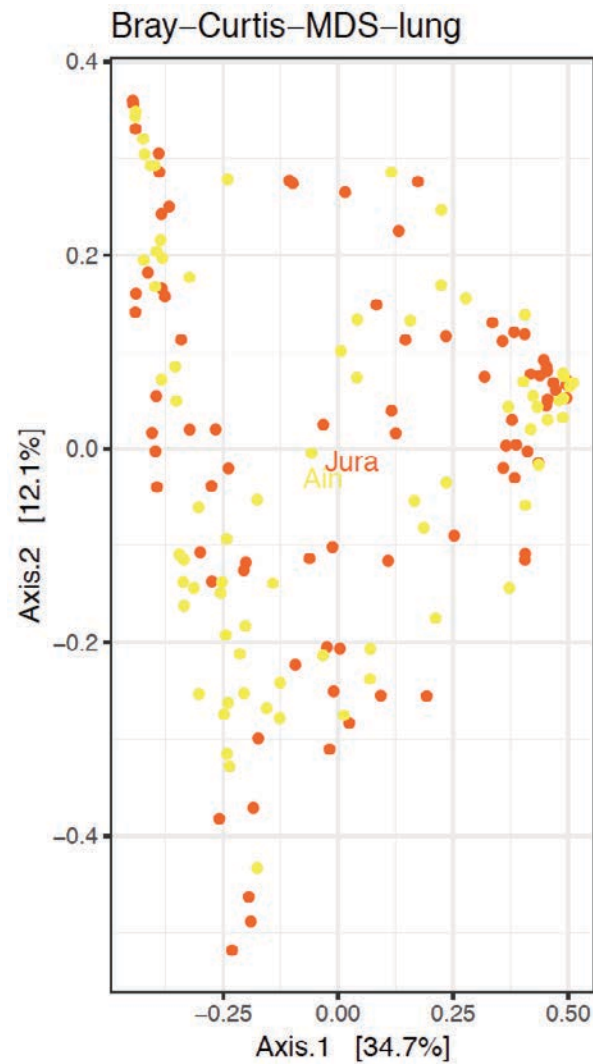
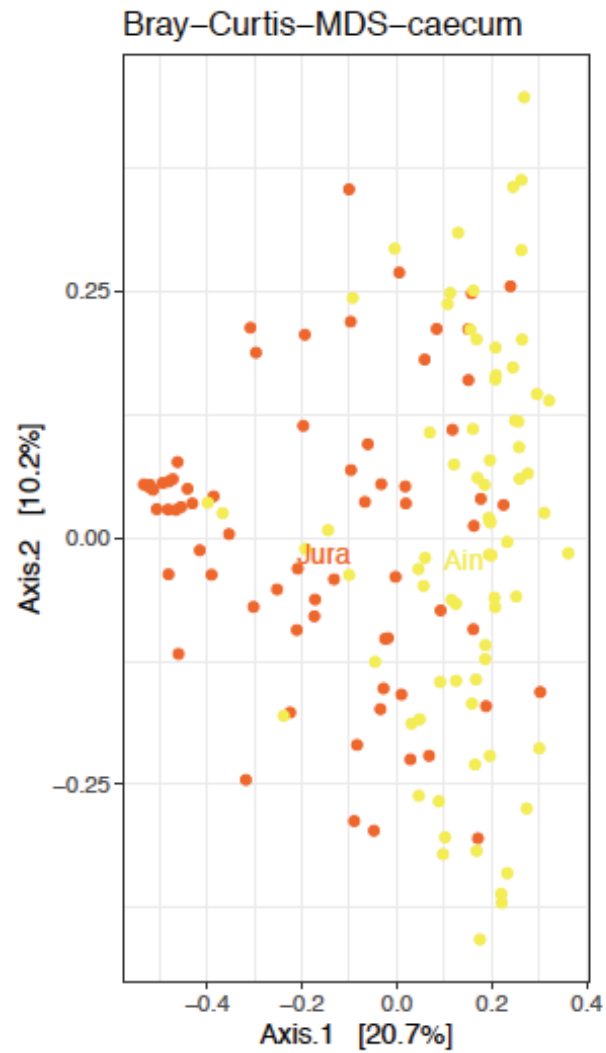
In progress for PUUV intra-host diversity analyses (see Guillaume Castel)

No Hypothesis : ‘catch me if you can’ strategy !

# BACTERIOME DIVERSITY AND COMPOSITION



# Beta diversity analyses



Differences of bacterial communities between Jura and Ain (Caecum & Colon not Lung or Spleen)



# INTERACTIONS BETWEEN BACTERIA AND PUUV

## Gut bacterial communities

Analyse how gut bacterial communities change during experimental infections

Analyse interactions between helminths and gut bacteria

## Lung and splenic bacteria

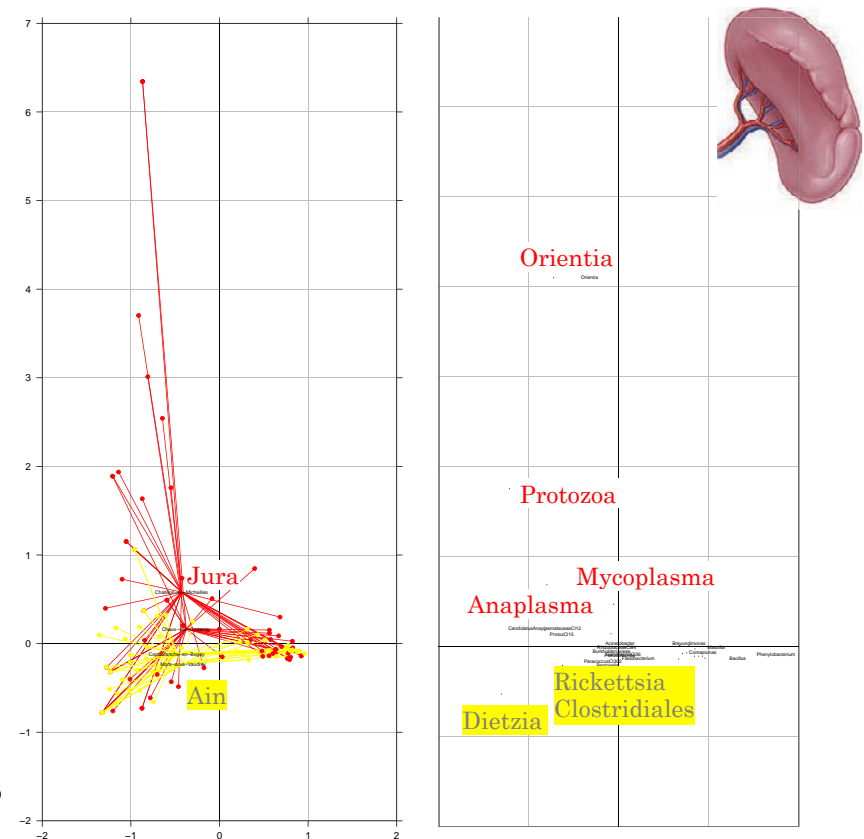
Focus on pathogenic bacteria  
&  
Detection of associations

## Jura / Ain - Spleen

*Bartonella, Anaplasma, Rickettsia ...*

## Discriminant analyses on spleen

\*\*\* Significant differences between regions



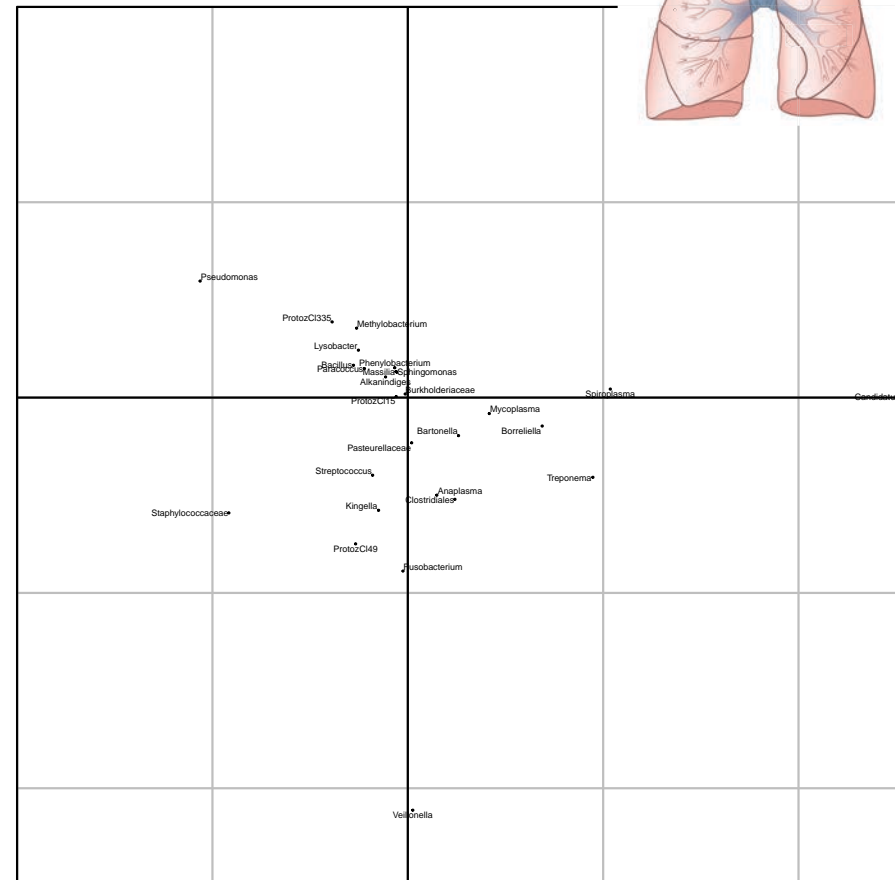
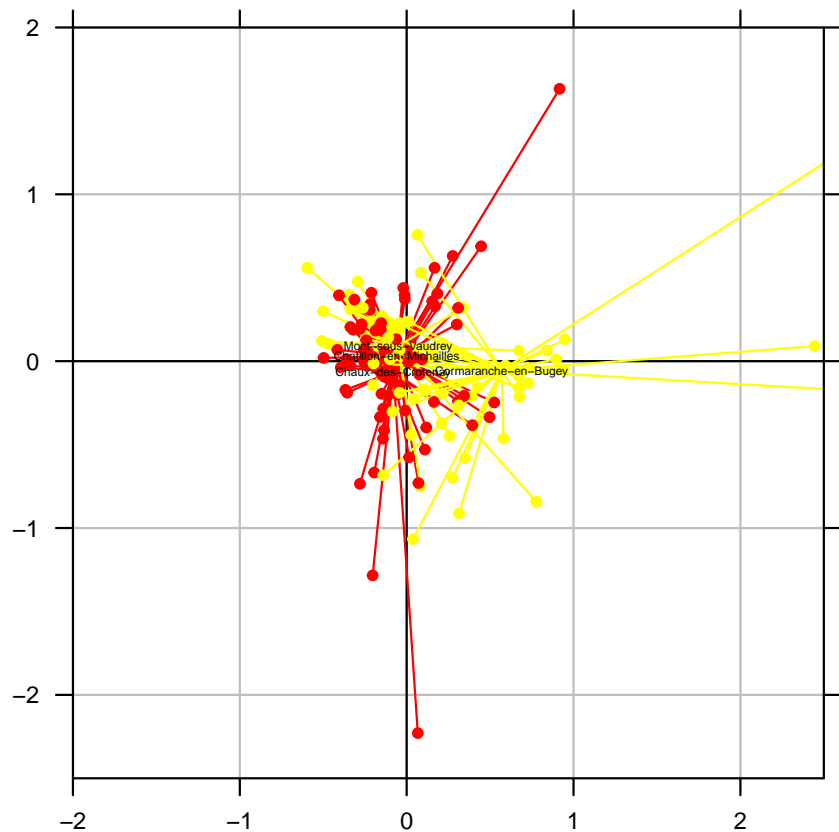
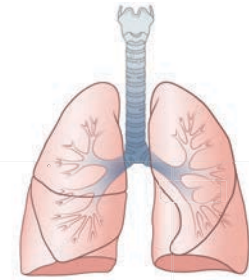
# INTERACTIONS BETWEEN BACTERIA AND PUUV

Jura / Ain – Lung

Pasteurellaceae, Pseudomonas, Paracoccus...

Discriminant analyses on lung

No significant differences between regions



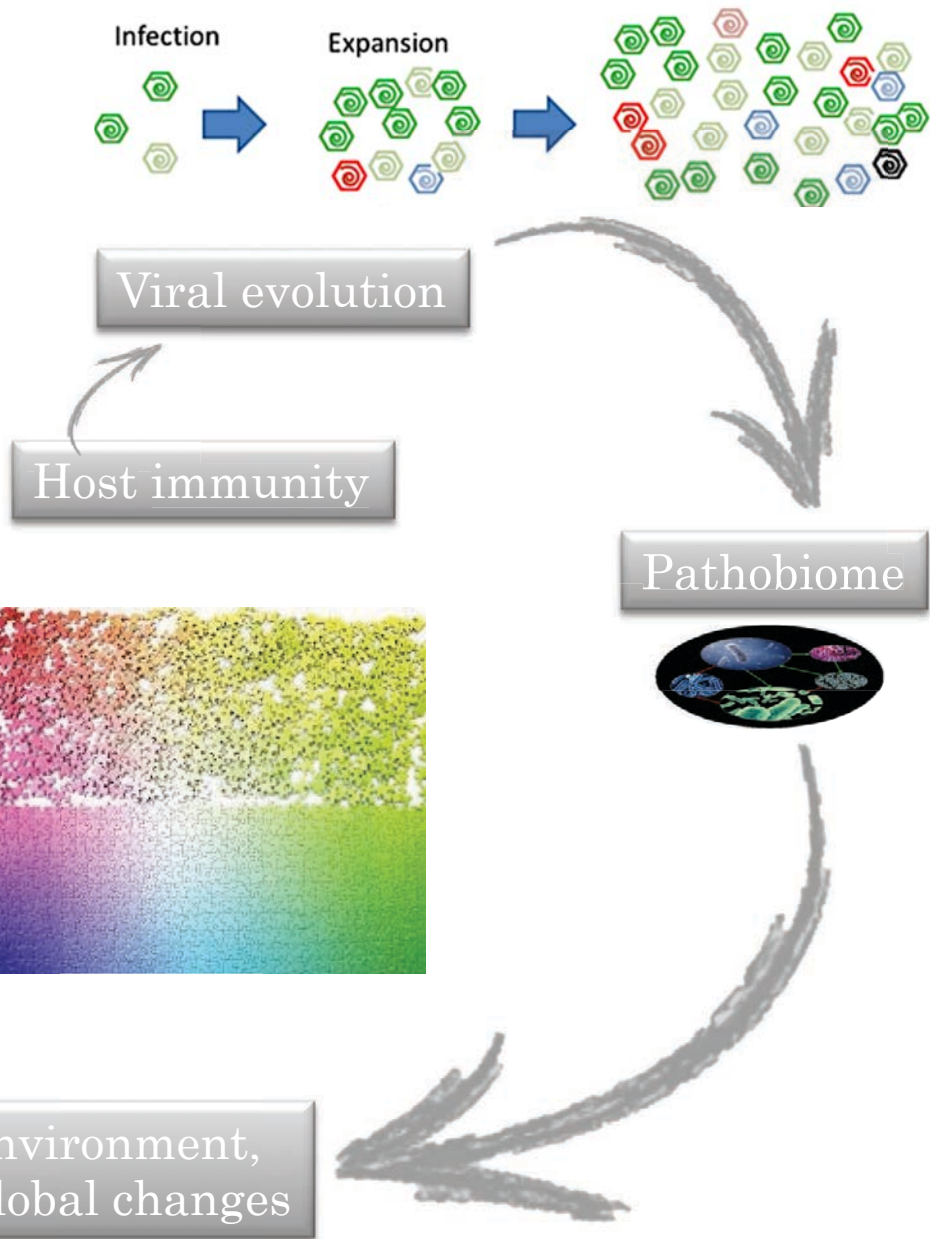
# CONCLUSIONS

## More information to gather

- Need to evaluate **tolerance**
- Need for a better knowledge of bank vole genome and transcriptome
- ...

More **integration** is required

Need epidemiologists to **understand** the consequences of the immune/viral variability on PUUV distribution





Anses Lyon

SLU Sweden

Haartman Finland

INRA Gabi, Genphyse

FP6 Eden  
FP7 Edenext

INRA EFPA

INRA MEM

ANSES

THANKS FOR YOUR ATTENTION

Photos L. Benoit





## Some related publications

- Dubois, A., Castel, G., Murri, S., Pulido, C., Pons, J.B., Benoit, L., Loiseau, A., Lakhdar, L., Galan, M., Charbonnel, N., Marianneau, P., 2017a. Experimental infections of wild bank voles (*Myodes glareolus*) from nephropatia epidemica endemic and non-endemic regions revealed slight differences in Puumala virological course and immunological responses. *Virus Research* 235, 67-72.
- Dubois, A., Castel, G., Murri, S., Pulido, C., Pons, J.B., Benoit, L., Loiseau, A., Lakhdar, L., Galan, M., Marianneau, P., Charbonnel, N., 2018. Bank vole immunoheterogeneity may limit nephropatia epidemica emergence in a French non-endemic region. *Parasitology* 145, 393–407.
- Dubois, A., Galan, M., Guivier, E., Henttonen, H., Voutilainen, L., Gauffre, B., Vitalis, R., Cosson, J.F., Charbonnel, N., 2017b. Microevolution of bank voles (*Myodes glareolus*) at neutral and immune-related genes during a multi-annual complete dynamic cycle : consequences for Puumala hantavirus epidemiology. *Infection Genetics Evolution* 49, 318–329.
- Guivier, E., Galan, M., Henttonen, H., Cosson, J.F., Charbonnel, N., 2014. Landscape features - fragmentation and helminth coinfection - shape bank vole immunoheterogeneity, with consequences for Puumala virus epidemiology. *Heredity* 112, 274-281.
- Razzauti-Feliu, M., Galan, M., Bernard, M., Maman, S., Klopp, C., Charbonnel, N., Vayssier-Taussat, M., Eloit, M., Cosson, J.F., 2015. Comparison of next-generation sequencing approaches surveying bacterial pathogens in wildlife. *PLoS Neglected Tropical Diseases* 9, e0003929.
- Ribas Salvador, A., Guivier, E., Chaval, Y., Xuéreb, A., Henttonen, H., Voutilainen, L., Poulle, M.L., Cadet, P., Cosson, J.F., Charbonnel, N., 2011. Concomitant influence of helminth infection and landscape on the distribution of Puumala hantavirus in its reservoir, *Myodes glareolus*. *BMC Microbiology* 11, 1-13.
- Rohfritsch, A., Galan, M., Gautier, M., Gharbi, K., Olsson, G.E., Gschloessl, B., Zeimes, C., VanWambeke, S., Vitalis, R., Charbonnel, N., 2018. Preliminary insights into the genetics of bank vole tolerance to Puumala hantavirus in Sweden. *Ecology and Evolution* 8, 11273–11292.
- Rohfritsch, A., Guivier, E., Galan, M., Chaval, Y., Charbonnel, N., 2013. Apport de l'immunogénétique à la compréhension des interactions entre le campagnol roussâtre *Myodes glareolus* et l'hantavirus Puumala. *Bulletin de l'académie vétérinaire de France* 166, 165-176.
- Vaumourin, E., Vourc'h, G., Telfer, S., Lambin, X., Salih, D.A., Seitzer, U., Morand, S., Charbonnel, N., Vayssier-Taussat, M., Gasqui, P., 2014. To be or not to be associated: Power study of four statistical modeling approaches to identify pathogen associations in cross-sectional studies. *Frontiers in Cell Inf Microbiol* 4, 62.